

Photosynthesis-driven microalgal system to mitigate carbon dioxide emission from power plant flue gases

DE-FE0032188

Yantao Li, Feng Chen, and Russell Hill, University of Maryland Center for Environmental Science;
Robert Mroz, HY-TEK Bio, LLC;
Troy Hawkins, Argonne National Laboratory
Wen Zhang, New Jersey Institute of Technology

2023 Carbon Management Research Project Review Meeting
August 28 – September 1, 2023

Project Overview

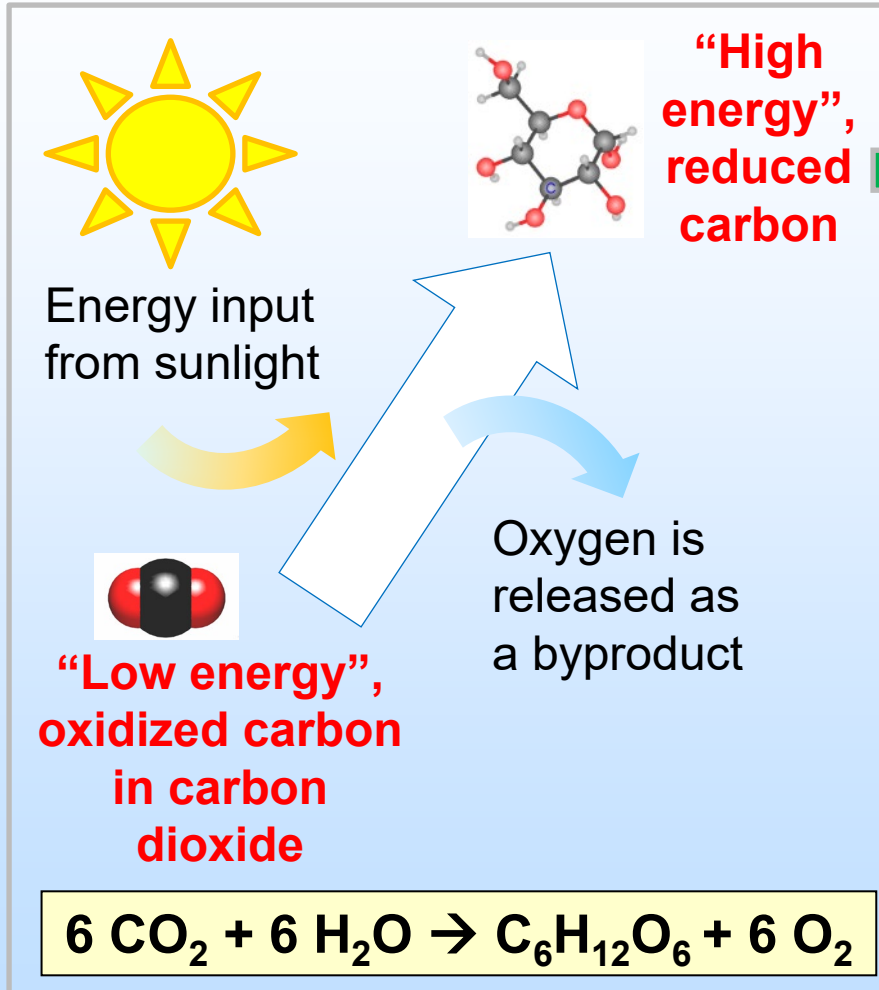
- Funding
 - DOE: \$2,000,000 and Cost Share: \$500,000
- Overall Project Performance Dates:
 - Feb. 2023 to Feb. 2026
 - BP1 (2/15/2023 to 8/14/2024), BP2 (8/15/2024 to 2/14/2026),
- Project Participants:
 - Yantao Li, Feng Chen, Russell Hill**, University of Maryland Center for Environmental Science;
 - Robert Mroz**, HY-TEK Bio, LLC;
 - Troy Hawkins**, Argonne National Lab
 - Wen Zhang**, New Jersey Institute of Technology
 - DOE NETL Program Manager: Zachary Roberts**

Project Overview

- Overall Project Objectives

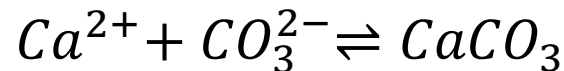
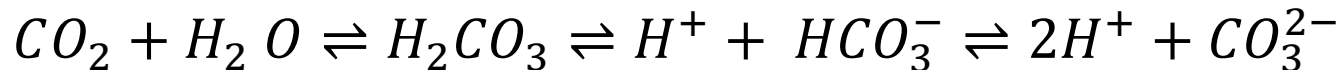
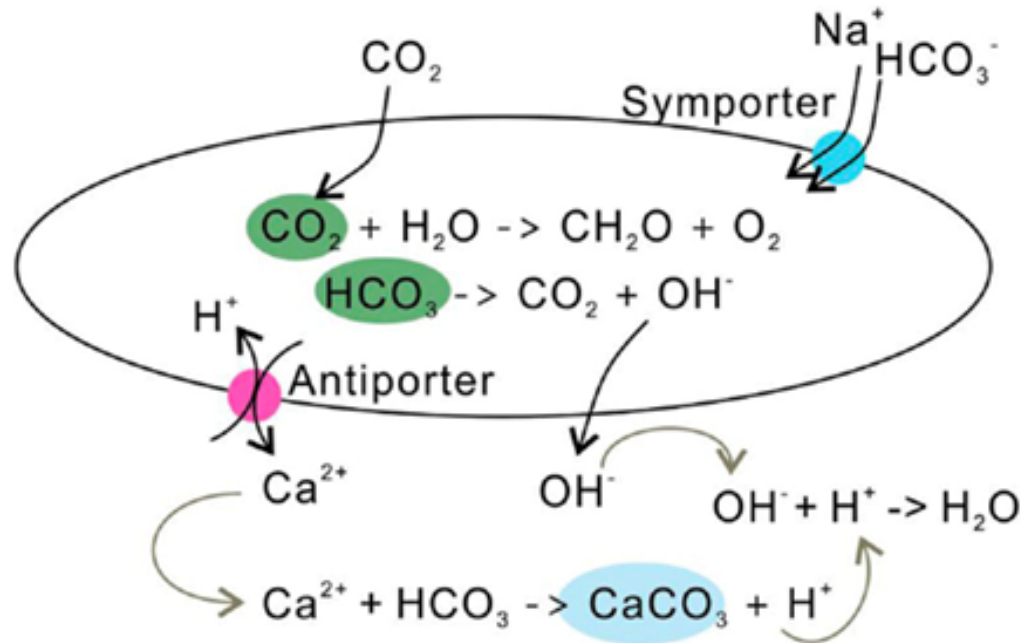
The objective of this project is to engineer microalgal polycultures through a photosynthesis-driven process to capture and sequester carbon dioxide (CO₂) from power plant flue gases in the form of algae biomass and carbonate precipitates.

Technology Background



Microalgal Carbon Capture and Biomass Production: Microalgae-driven carbon dioxide mitigation (MadCom)

Photosynthesis



Microalgae-driven carbon dioxide mitigation in nature: Whiting events



13-year data, Florida Whitings visible in MODIS images on about 14% of time each year

<https://earthobservatory.nasa.gov/images/91807/making-sense-of-whiting-events>

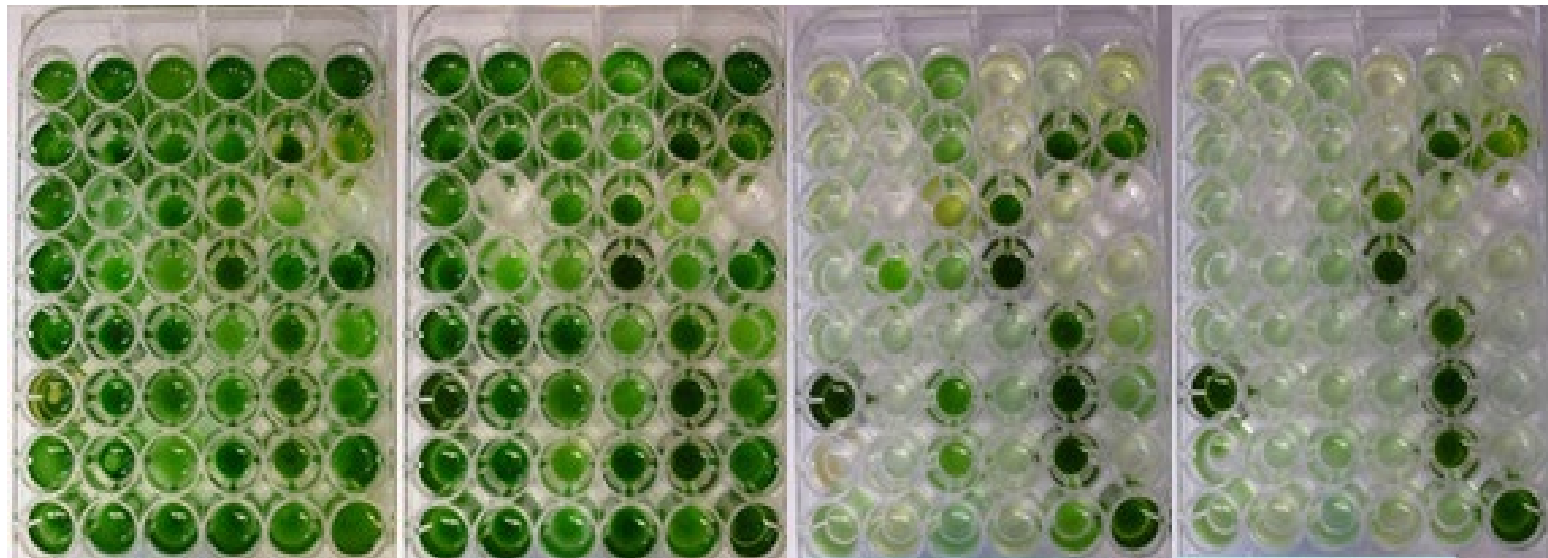
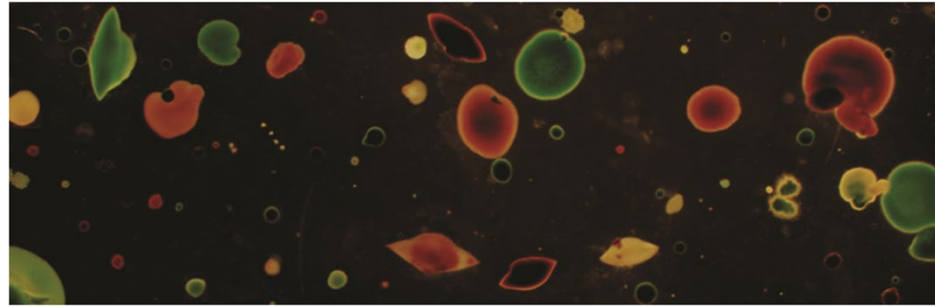
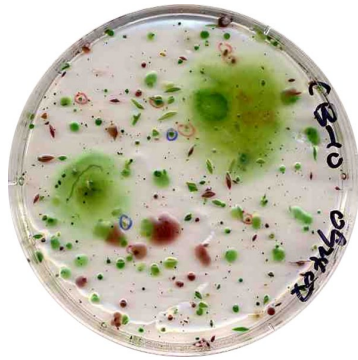
Escoffier, N. et al. 2022. Journal of Geophysical Research: Biogeosciences.



Marl from lake bottom (Courtesy of Larry Bean, rock collector, Livonia, Michigan.), <https://www.michigan.gov/>

Algal Strain Selection under Simulated Flue Gas

Nannochloropsis oceanica IMET1 and *Scenedesmus obliquus* HTB1 were selected



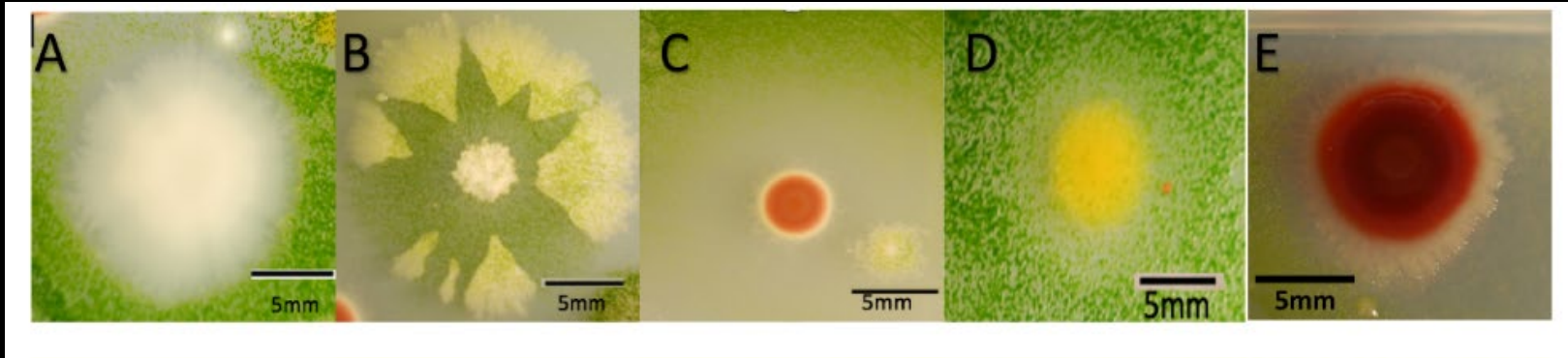
2% CO₂

5% CO₂

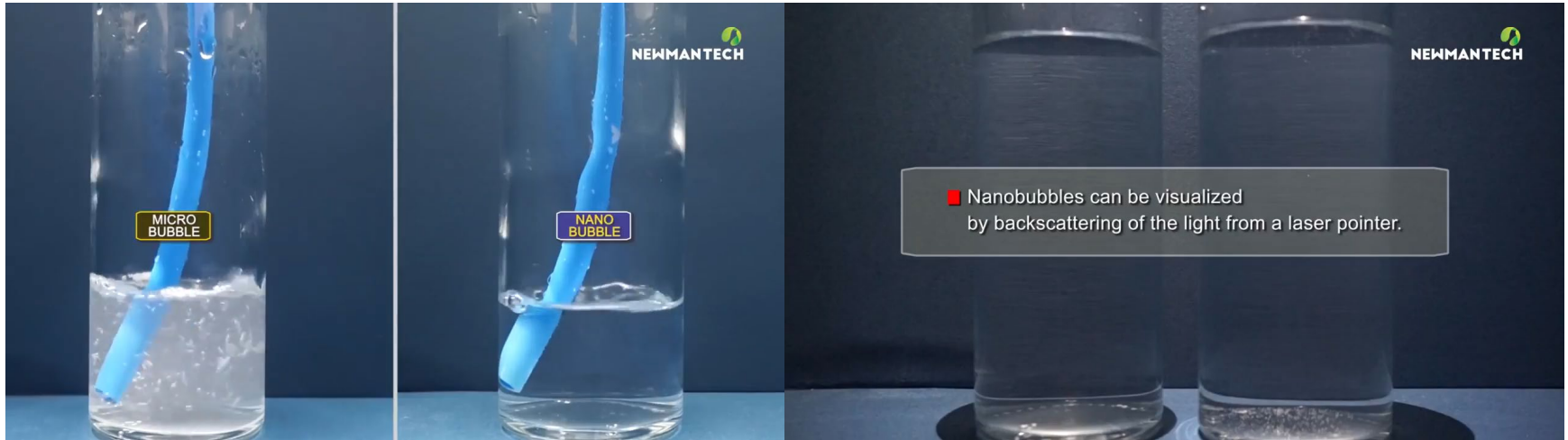
10% CO₂

20% CO₂

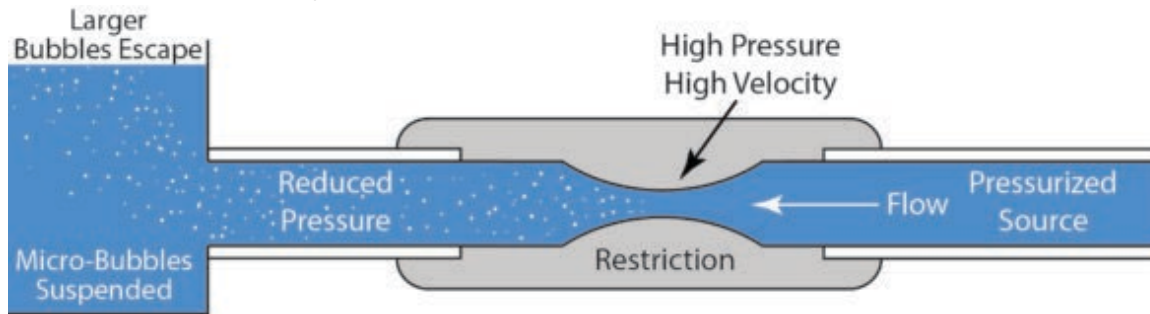
Microbial interactions in non-axenic microalgal cultures



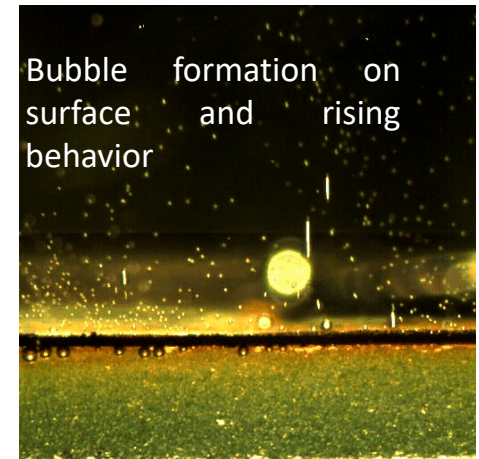
Microbubbles and nanobubbles to improve carbon utilization efficiency (CUE)



Video sources: <http://newmantech.co.kr/nanoe/>

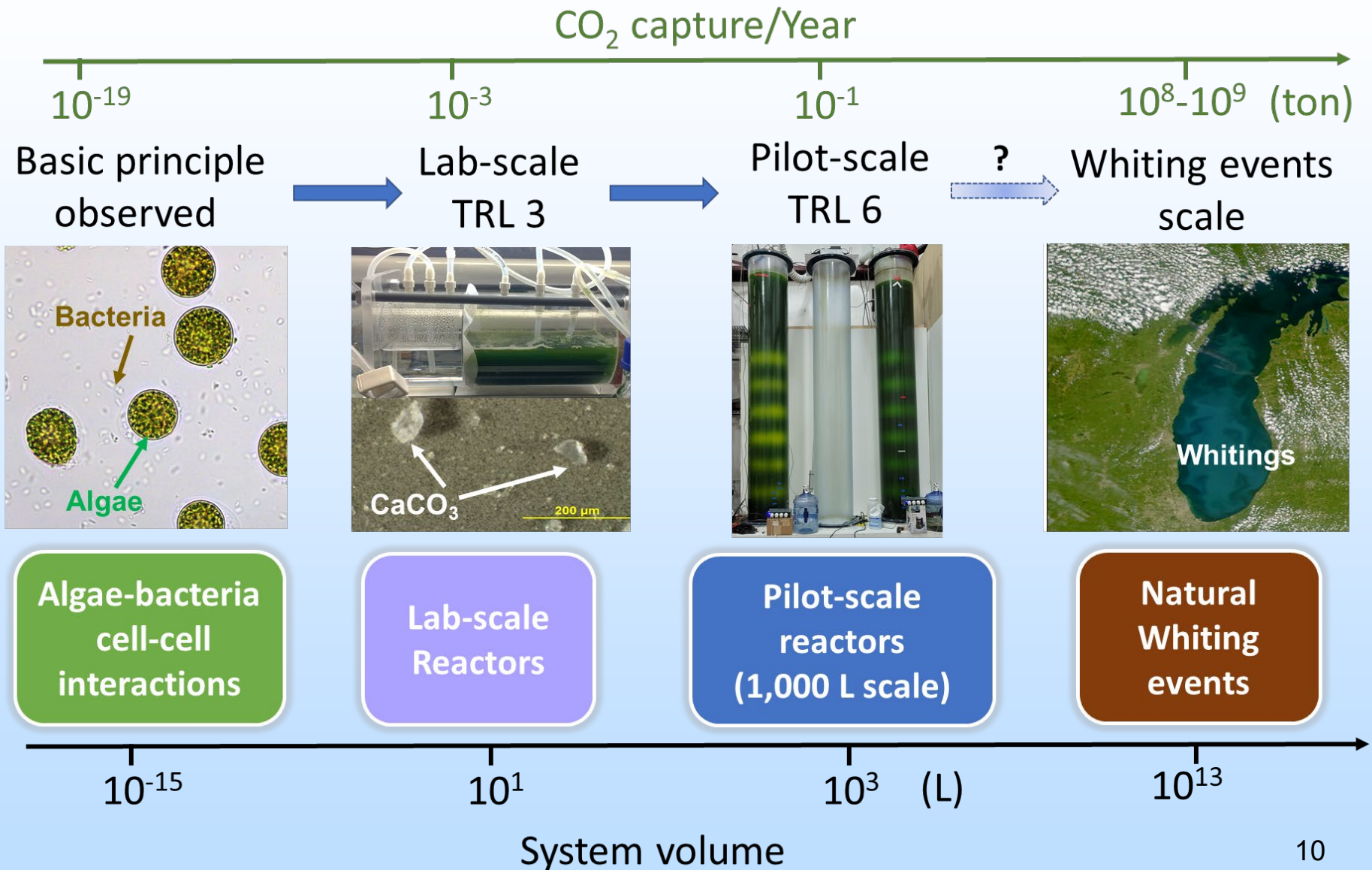


<https://techblog.ctgclean.com/2019/01/the-case-for-micro-bubbles-in-cavitation/>

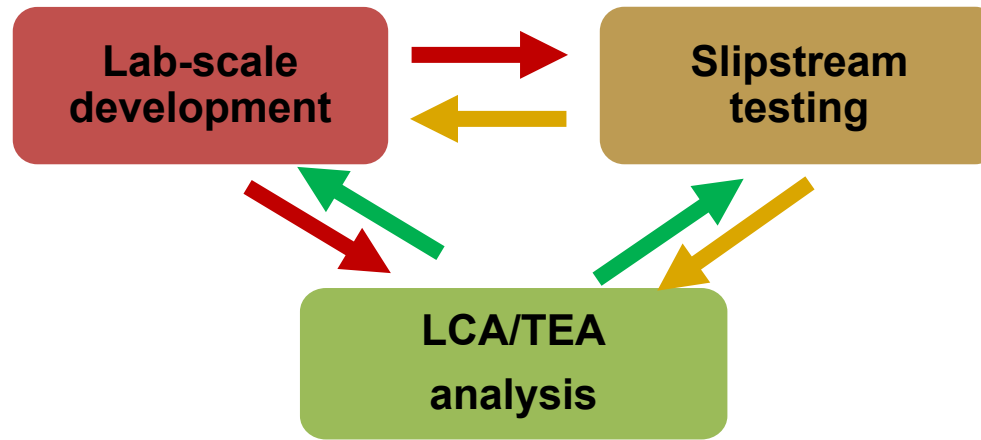


Wen Zhang and NJIT team

Proposed Microalgae-driven carbon dioxide mitigation TRL



Project overview



Lab-scale development of algal system and culture microbiome optimization (UMCES)

- Subtask 2.1; 3.2: Saltwater algal system and microbiome optimization (**Li and Hill**)
- Subtask 2.2; 3.3: Freshwater algal system and microbiome optimization (**Chen and Hill**)

Development and testing of bubblers in the lab and upscaled algal systems (NJIT)

- Subtask 2.3, 3.4: Develop and optimize micro-/nano- bubblers (**Zhang**)

Slipstream testing of the algal carbon sequestration system (HY-TEK Bio)

- Subtask 2.4, 3.1: Slipstream test at 9 L and 1,000 L (**Mroz**)

Development of TEA and LCA models (Argonne)

- Subtask 4.0: Perform TEA and LCA analysis (**Hawkins**)

Progress: Budget Period 1 Bench-scale and 9 L scale test

Subtask 2.1 - Laboratory development of seawater *Nannochloropsis* system

- Analyze the culture microbiome to assess changes in microbial community;
- Isolate and test urease-producing and probiotic bacterial strains;
- Measure CaCO₃ precipitates, culture alkalinity, and biomass yield.

Milestone 2.1 Isolate and confirm the identity of >5 urease-producing bacteria and >5 probiotic bacterial strains for *Nannochloropsis oceanica* IMET1; Date: M15



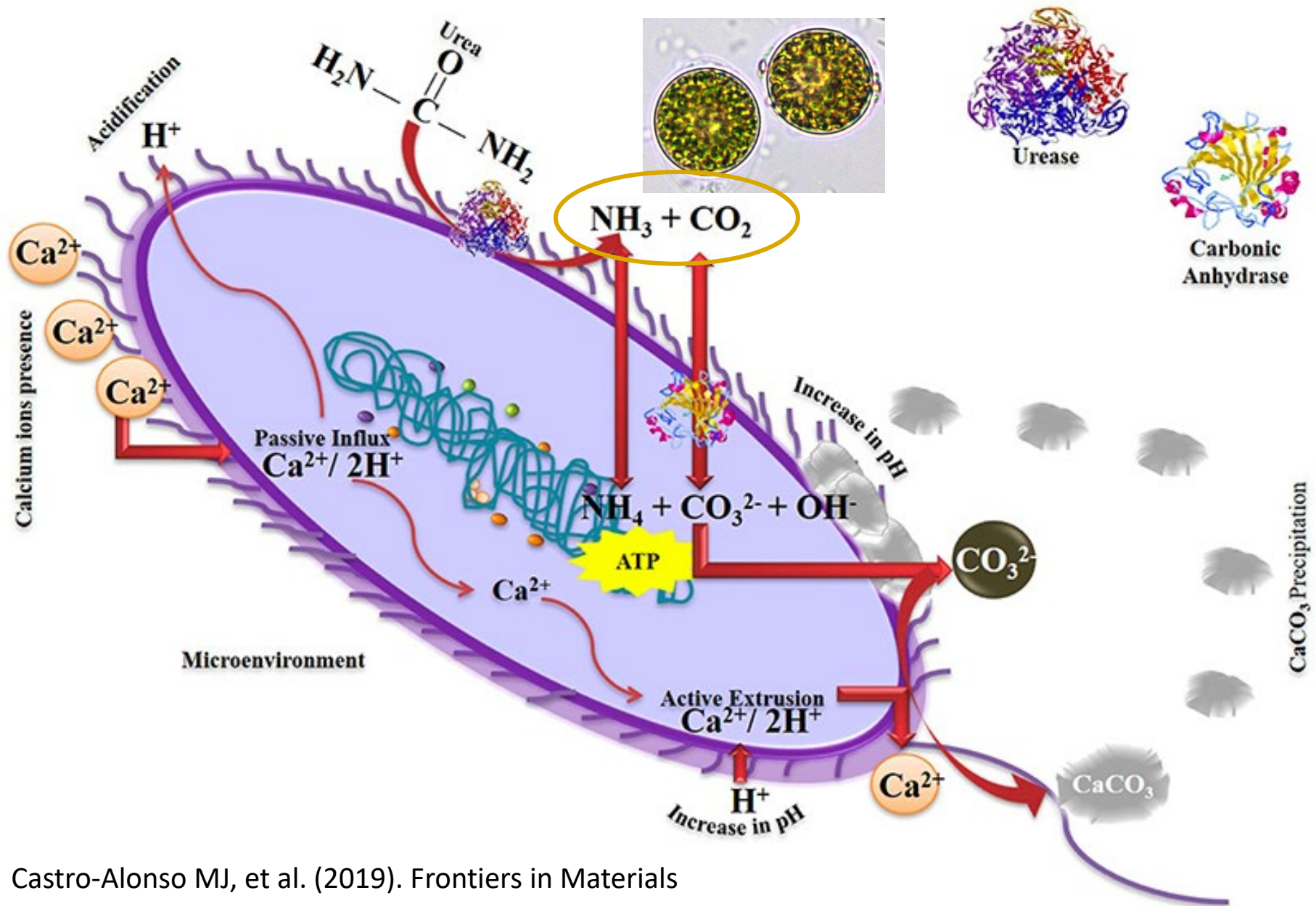
Progress: Budget Period 1 Bench-scale and 9 L scale test

Subtask 2.2 - Laboratory development of freshwater *Scenedesmus* system

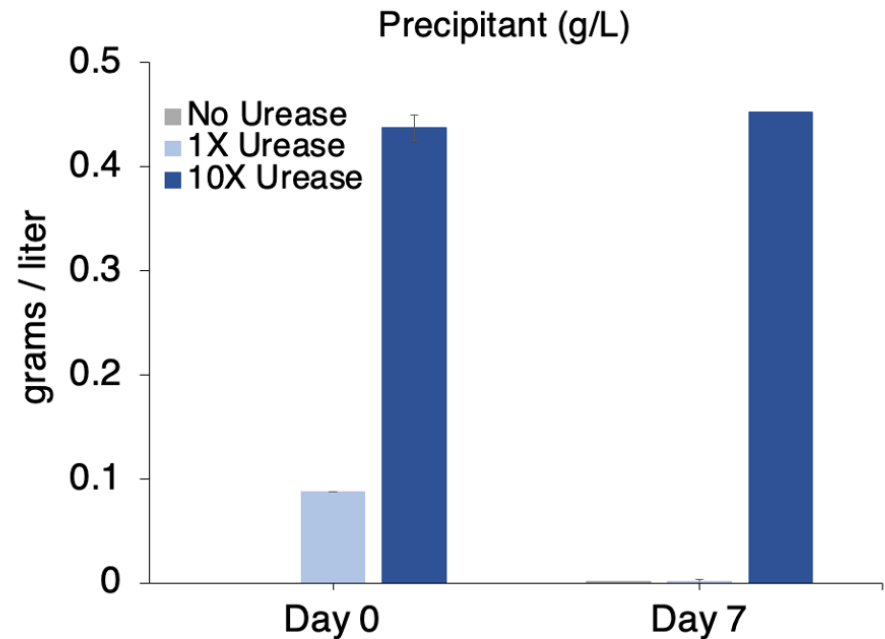
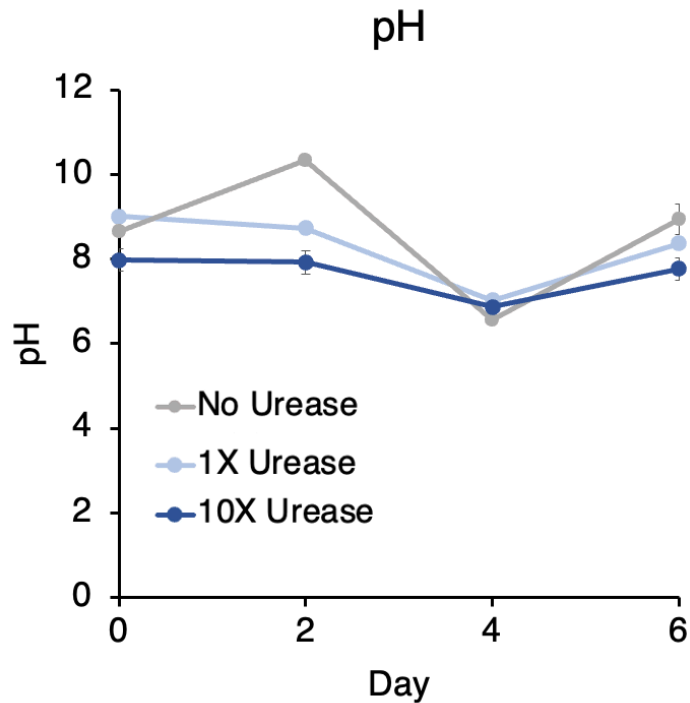
- Analyze the culture microbiome to assess changes in microbial community;
- Isolate and test urease-producing and probiotic bacterial strains;
- Measure CaCO₃ precipitates, culture alkalinity, and biomass yield.

Milestone 2.2 Isolate and confirm the identity of >5 urease-producing bacteria and >5 probiotic bacterial strains for *Scenedesmus* HTB1; Date: M15

Progress: Budget Period 1 Bench-scale and 9 L scale test

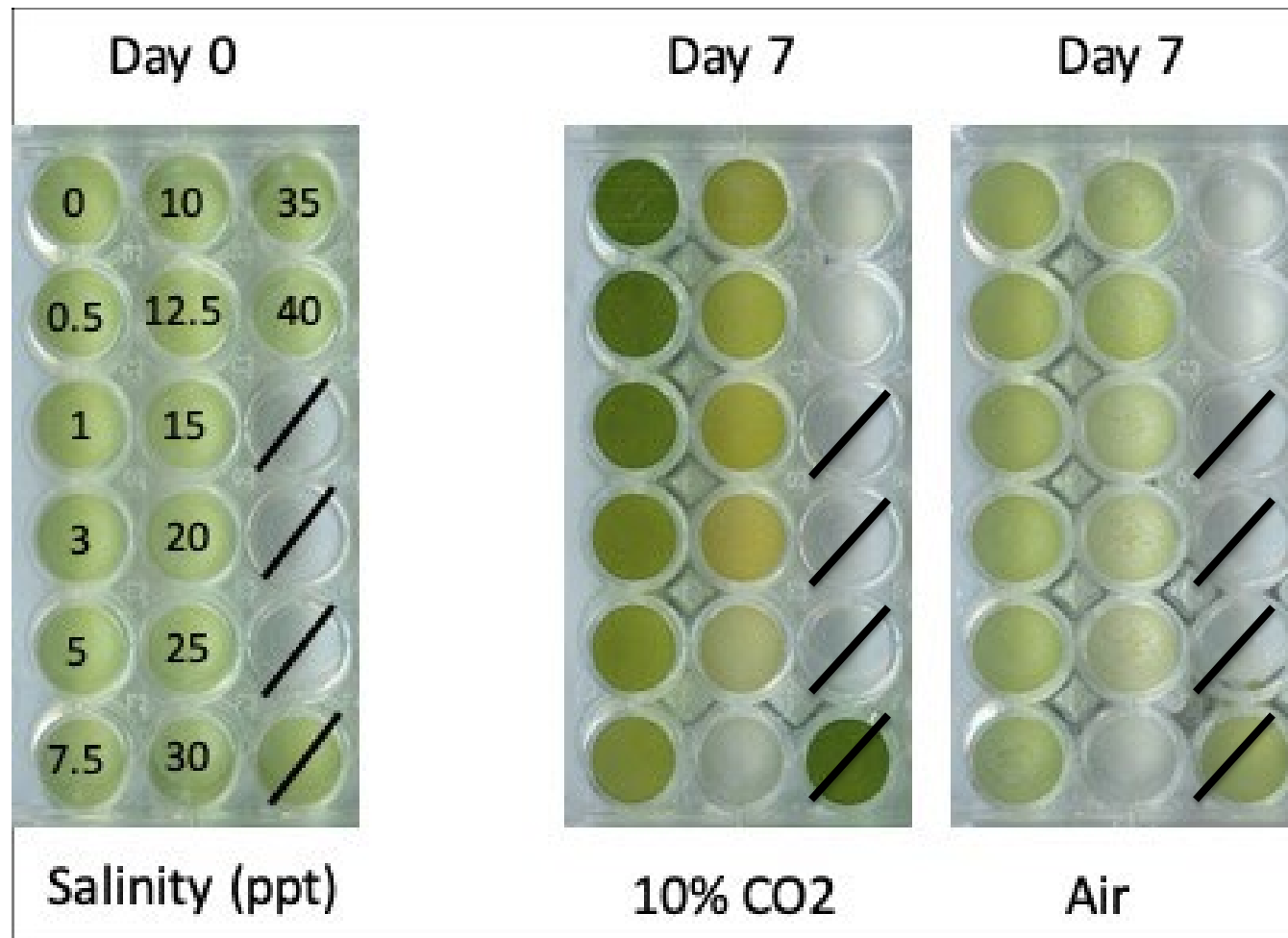


Progress: Budget Period 1 Bench-scale and 9 L scale test



Progress: Budget Period 1 Bench-scale and 9 L scale test

Scenedesmus HTB1 tolerates varying salinity gradients and produces carotenoids



Budget Period 1: Bench-scale and 9 L-scale field test

Subtask 2.3 - Development and testing of bubblers

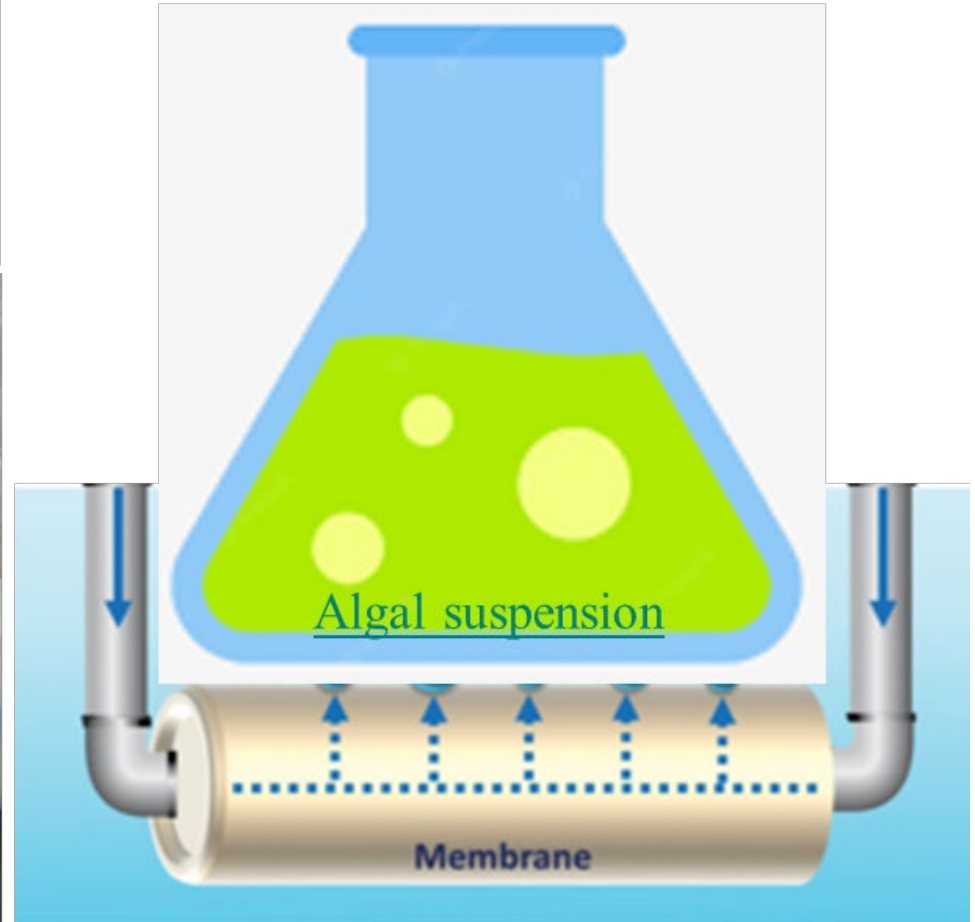
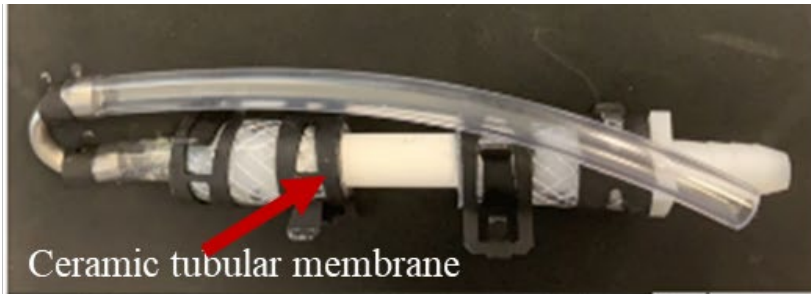
- Generate and optimize micro- and nano-bubbles of CO₂ dispersion;
- Determine key parameters such as bubble sizes and CO₂ flow or flux

Milestone 2.3 Generate CO₂ nanobubbles with concentrations of up to 3×10^{14} bubbles·L⁻¹ and sizes ranging from 300 nm to 1 μm in diameter and microbubbles with concentrations of up to 1×10^8 bubbles·L⁻¹ and sizes ranging from 10 to 100 μm;

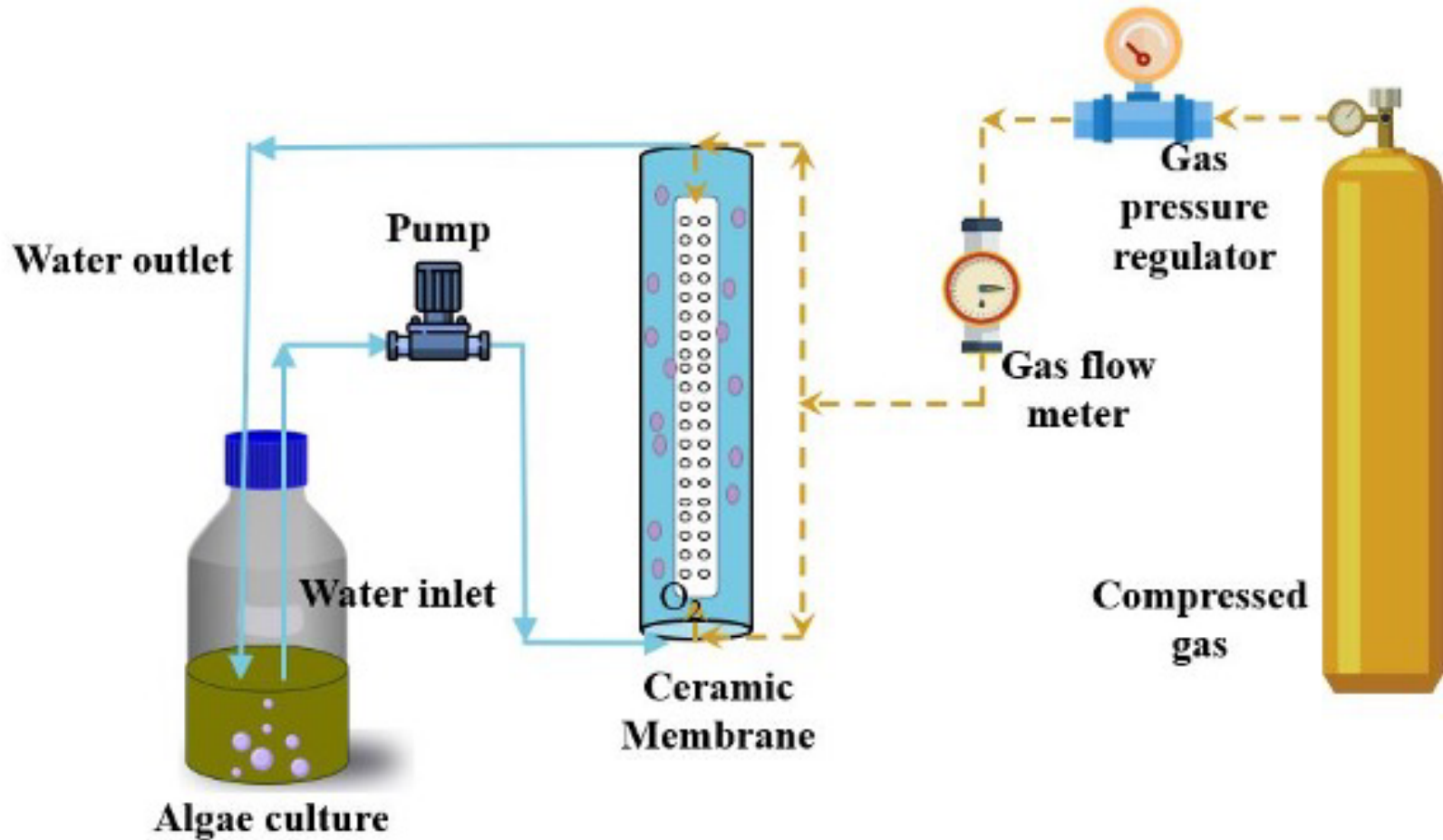
Date: M12



Progress: Budget Period 1 Bench-scale and 9 L scale test



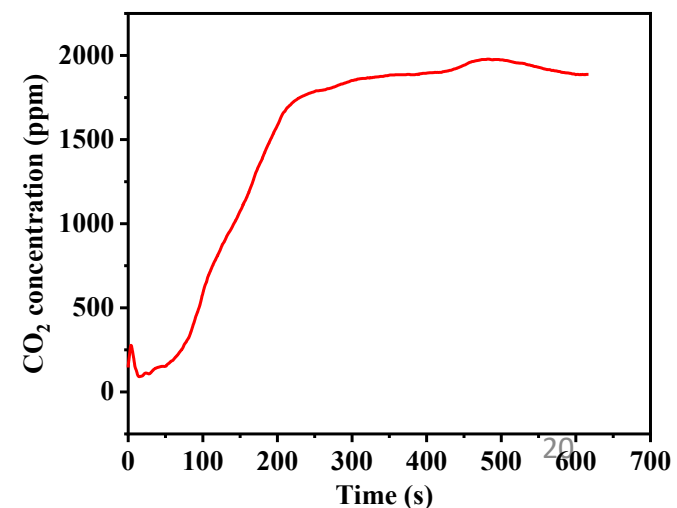
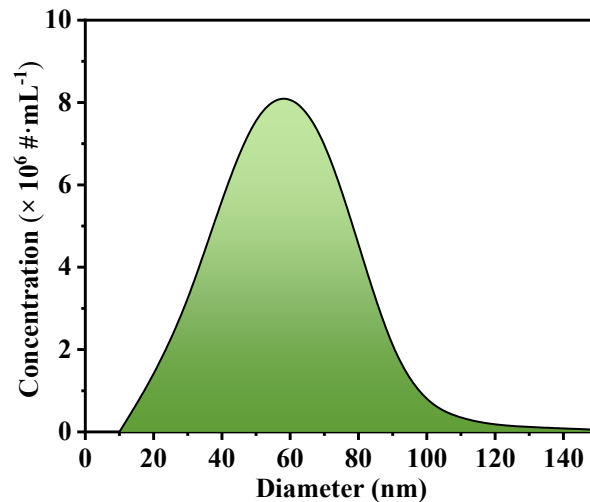
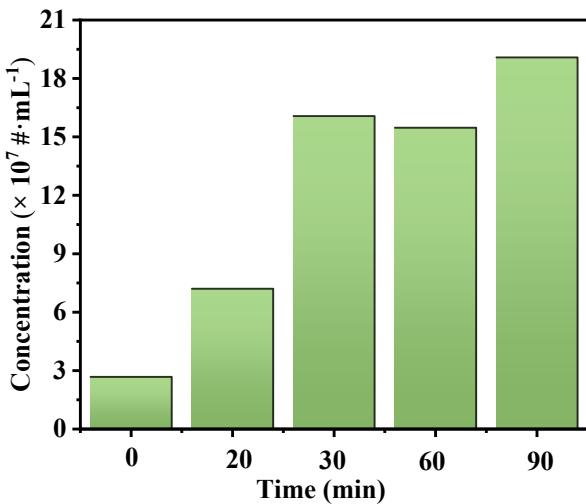
Progress: Budget Period 1 Bench-scale and 9 L scale test



Progress: Budget Period 1 Bench-scale and 9 L scale test



1. Using pure CO_2 , the observed dissolved CO_2 in the nanobubble water suspension is near 2,000 ppm, which corresponds to a partial pressure of **1.18 atm** (the injection pressure is 145 kPa or 1.43 atm).
2. The ambient air solubility under one atm can provide only **0.0004 atm** of the partial pressure of CO_2 . Thus, the nanobubbles of CO_2 increases the partial pressure up to 1.18 atm and resulted in nearly **3,000 times** of the CO_2 solubility under ambient air.



Progress: Budget Period 1 Bench-scale and 9 L scale test

Subtask 2.4 - Algal Culture Improvement at the flue gas site

- Use 9-L bioreactor systems to clean flue gas with algae culture.

Subtask 2.5 - Develop the frameworks for the TEA and LCA models

- Develop frameworks of LCA and TEA models for sunlight-driven seawater and freshwater algal carbon sequestration systems.

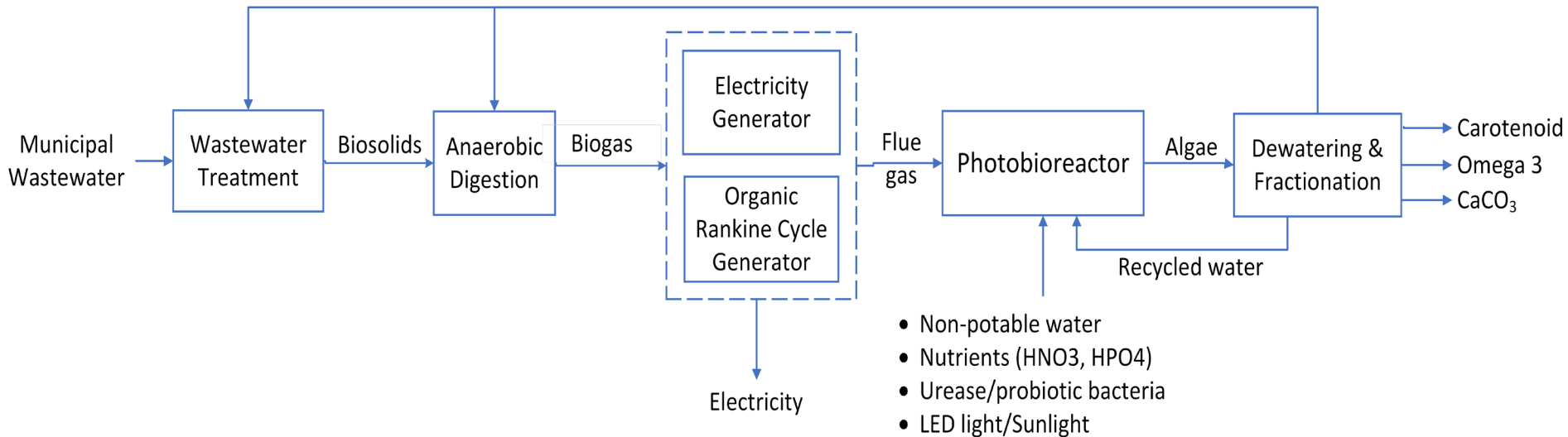
BP1 Success Criteria: Determine the best combinations of urease and probiotic bacterial strains and bubbling mechanisms for *S. obliquus* HTB1 and *N. oceanica* IMET1 that facilitate an average productivity of 30 g/m²/day and a CUE >50% (algae biomass) or >60% (algae and CaCO₃ precipitates) in lab culture and 9-L bioreactors. Date: M18

HY-TEK Bio's Facility at the Back River Waste Water Treatment Plant



Current HTB site in operation for more than 8yrs

Progress: Budget Period 1 Bench-scale and 9 L scale test



LCA and TEA hotspots: to identify environmental and cost hotspots and provide feedback to the team members to improve the process design

LCA metrics:

- Greenhouse gas (GHG) emissions (CO₂, CH₄, N₂O)
- Criteria air pollutant emissions (VOC, CO, NO_x, PM₁₀, PM_{2.5}, and SO_x)
- Fossil energy use
- Water consumption

TEA metrics:

- Cost
- Return on investment
- Marginal cost of GHG avoidance

Plans for future work- BP1

Milestone Title	Planned Completion date	Actual Completion date	Verification Method
<u>Milestone 2.1:</u> Isolate and confirm the identity of >5 urease-producing bacteria and >5 probiotic bacterial strains for <i>Nannochloropsis oceanica</i> IMET1	Month 15	N/A	Via Quarterly Reports submitted to DOE Project Officer
<u>Milestone 3.2:</u> Isolate and confirm the identity of >5 urease-producing bacteria and >5 probiotic bacterial strains for <i>Scenedesmus obliquus</i> HTB1	Month 15	N/A	Same as above
<u>Milestone 3.3:</u> Generate CO ₂ nanobubbles with concentrations of up to 3×10^{14} bubbles·L ⁻¹ and sizes ranging from 300 nm to 1 μm in diameter and microbubbles with concentrations of up to 1×10^8 bubbles·L ⁻¹ and sizes ranging from 10 to 100 μm	Month 12	N/A	Same as above

No change in the scope of work, and no extra fund requested.

Summary

- Urease effect on carbonate precipitation confirmed; further tests in progress.
- Micro-/Nano-bubble generation tests in progress; we will proceed with lab tests of seawater and freshwater cultures.
- Bioreactor parts ordered and in preparation for bioreactor tests with flue gas.

Appendix

- These slides will not be discussed during the presentation **but are mandatory.**

Organization Chart

No.	/Tasks	/Subtasks and PIs responsible for the task	Teams responsible
1.1 1.2	Project Management and Planning	<ul style="list-style-type: none"> • Project Management Plan (<i>Li working with all PIs</i>) • Project Reporting (<i>All PIs</i>) 	UMCES is the lead on this task.
2.1, 2.2, 3.2, 3.3.	Bench-scale development of a saltwater and a freshwater system and culture microbiome optimization	<ul style="list-style-type: none"> • Saltwater algal carbon sequestration system (<i>Li and Hill</i>) • Freshwater algal carbon sequestration (<i>Chen and Hill</i>) 	UMCES is the lead on this task.
2.3, 3.4	Development and testing of bubblers in the lab and upscaled algal systems	<ul style="list-style-type: none"> • Optimization of microbubbles and nanobubbles in lab cultures (<i>Zhang</i>) • Bubbler optimization at 1,000 L scale (<i>Zhang</i>) 	NJIT is the lead on this task.
2.4, 3.1	Slipstream testing of the algal carbon sequestration system at the Back River wastewater treatment plant	<ul style="list-style-type: none"> • Bioreactor test on site (at the wastewater treatment plant) at 9 L scale (<i>Mroz</i>) • Slipstream test at 1,000 L scale (<i>Mroz</i>) 	HY-TEK Bio is the lead on this task.
4.0	Development of TEA and LCA models to evaluate and guide research and testing activities.	<ul style="list-style-type: none"> • Develop the frameworks for the TEA and LCA models (<i>Hawkins</i>) • Perform contribution analysis, benchmarked against other conventional algae processes (<i>Hawkins</i>) 	Argonne National Lab is the lead on this task.

Gantt Chart

*Project Schedule (Gantt chart)**

Task	2023-24						2024-26					
	BP1						BP2					
2.1 Develop a seawater system						★						
2.2 Develop a freshwater system						★						
2.3 Engineer micro-/nano- bubblers				★								
2.4 Onsite lab-scale tests												
3.1 Slipstream testing at 1,000 L scale									★			
3.2 Optimize seawater culture										★		
3.3 Optimize freshwater culture												★
3.4 Optimize micro-/nano- bubblers												★
4.0 LCA/TEA												★

★ Milestone ★ Go-No Go

* Start date was Feb. 15, 2023; each block represents one quarter (3-month). At the end of the first BP, there is a Go-No Go decision point.