# Innovative Water Management Technology to Reduce Environmental Impacts of Produced Water

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### FINAL REPORT

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### **ABSTRACT**

Clemson University with Chevron as an industry partner developed and applied treatment technology using constructed wetland systems to decrease targeted constituents in simulated and actual produced waters to achieve reuse criteria and discharge limits. Pilot-scale and demonstration constructed wetland treatment system (CWTS) experiments led to design strategies for treating a variety of constituents of concern (COCs) in produced waters including divalent metals, metalloids, oil and grease, and ammonia. Targeted biogeochemical pathways for treatment of COCs in pilot-scale CWTS experiments included divalent metal sulfide precipitation through dissimilatory sulfate reduction, metal precipitation through oxidation, reduction of selenite to insoluble elemental selenium, aerobic biodegradation of oil, nitrification of ammonia to nitrate, denitrification of nitrate to nitrogen gas, separation of oil using an oilwater separator, and sorption of ammonia to zeolite. Treatment performance results indicated that CWTSs can be designed and built to promote specific environmental and geochemical conditions in order for targeted biogeochemical pathways to operate. The demonstration system successfully achieved consistent removal extents even while inflow concentrations of COCs in the produced water differed by orders of magnitude. Design strategies used in the pilot-scale and demonstration CWTSs to promote specific conditions that can be applied to designing full-scale CWTSs include plant and soil selection, water-depth selection, addition of amendments, and hydraulic retention time (HRT). These strategies allow conditions within a CWTS to be modified to achieve ranges necessary for the preferred biogeochemical treatment pathways. In the case of renovating a produced water containing COCs that require different biogeochemical pathways for treatment, a CWTS can be designed with sequential cells that promote different conditions. For example, the pilot-scale CWTS for post-reverse osmosis produced water was designed to promote oxidizing conditions within the first wetland cell for nitrification of ammonia, and the subsequent three cells were designed to promote reducing conditions for denitrification of nitrate. By incorporating multiple wetland cells in a CWTS, the conditions within each cell can be modified for removal of specific COCs. In addition, a CWTS designed with multiple cells allows for convenient sample collection points so that biogeochemical conditions of individual cells can be monitored and performance evaluated. Removal rate coefficients determined from the pilot-scale CWTS experiments and confirmed by the demonstration system can be used to calculate HRTs required to treat COCs in full-scale CWTSs. The calculated HRTs can then be used to determine the surface area or "footprint" of a full-size CWTS for a given inflow rate of produced water.

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### **EXECUTIVE SUMMARY**

Clemson University with Chevron as an industry partner developed and applied technology to decrease targeted constituents in simulated and actual produced waters to achieve reuse criteria and discharge limits. Phase I of the project included assessment of environmental factors associated with produced waters that impact development of U.S. oil and gas reserves, particularly factors pertinent to application of constructed wetland systems for the treatment of produced waters. Phase I began with completion of a Project Management Plan (Task 1). Chemical, physical, and risk characteristics of produced waters were identified in Task 2. A literature search was completed, and characterization data for specific types of produced waters were compiled. The data indicate a wide range of characteristics, with produced waters containing various metals, organics (e.g. oil and grease), and other constituents. Reuse and discharge criteria and specific treatment performance goals were determined in Task 3. The results of Tasks 2 and 3, including identification of constituents of concern in produced waters and development of treatment performance goals, were important for moving ahead to Phase II and designing systems to provide treatment for water discharge or reuse.

Phase II involved investigation of prototype (pilot-scale) constructed wetland treatment systems. The pilot-scale systems were designed and constructed in Task 4. Treatment performance of the pilot-scale systems was measured (Task 5) and assessed (Task 6). Performance measurements from the pilot-scale constructed wetland systems demonstrated successful design strategies for treating a variety of constituents of concern including divalent metals, metalloids, oil and grease, and ammonia. Targeted pathways for treating these constituents in the pilot-scale experiments included divalent metal sulfide precipitation through dissimilatory sulfate reduction, metal precipitation through oxidation, reduction of selenite to insoluble elemental selenium, aerobic biodegradation of oil, nitrification of ammonia to nitrate, denitrification of nitrate to nitrogen gas, separation of oil using an oil-water separator, and sorption of ammonia to zeolite.

Experiments in support of designing demonstration and full-scale constructed wetland treatment systems were performed to gain a better understanding of conditions affecting treatment pathways and hybridized design strategies that can be incorporated into constructed wetland treatment. Experiments included comparing seedling growth of corn, cabbage, and soybean between untreated and treated simulated produced water. The difference in early seedling growth between the waters was determined to be statistically significant, with higher growth rate in the treated water. Toxicity tests using Ceriodaphnia dubia and Pimephales promelas compared the potential between untreated and treated simulated produced water to cause adverse effects to receiving system biota. Statistical analysis indicated a significant difference in the responses of C. dubia and P. promelas between the untreated and treated water, with greater survival in the treated water. Evapotranspiration data and comparison of theoretical lysimeter outflow determined using the Penman Monteith equation with measured lysimeter outflow demonstrated application of the equation to estimating evapotranspiration in the pilot-scale constructed wetland treatment systems used in this investigation. Additional experiments were performed to determine the effect of plant transpiration on transport of constituents through the hydrosoil and the effects of seasonal changes on wetland performance. Wetland cells containing Typha latifolia were more effective at transporting constituents through the hydrosoil than wetland cells that did not contain *Typha* as indicated by shorter tracer breakthrough times.

The influence of water depth on treatment performance of the pilot-scale constructed wetlands for renovating simulated fresh oilfield produced water was investigated. Divalent metal concentrations were greater in outflow from wetland cells having 15 and 23 cm water depths than from cells having 46 and 56 cm water depths. The cells having 46 and 56 cm water depths provided reducing conditions conducive for dissimilatory sulfate reduction, which contributed to removal of divalent metals from the simulated produced water. Wetland cells with 15 and 23 cm water depths were more effective for removing oil in the simulated produced water than cells with greater water depths because the shallower depth promoted conditions favorable for aerobic degradation of oil.

Phase III included on-site treatment of produced water using a demonstration constructed wetland system. In collaboration with Chevron, a site for constructing the demonstration system was selected near Berry, Alabama. This site was within a producing coalbed methane field. Data from the pilot-scale experiments were used in Task 7 to design and construct the demonstration system. The system consisted of four wetland series designed to treat the following constituents of concern in water produced from the field: ammonia, barium, cadmium, iron, manganese, nitrate, and selenium. The system included sequentially arranged oxidizing and reducing cells designed to promote conditions identified during Phase II for targeted treatment pathways. Monitoring of treatment performance of the demonstration constructed wetland system (Task 8) from September 2012 through March 2013 indicated effective removal of constituents of concern, even as inflow concentrations of these constituents in the produced water varied. Sampling and analytical methods were used that allowed for calculation of removal efficiency, removal rate, and removal extent in order to assess treatment performance.

Results were applied to developing strategies for water management and technology replication in Task 9. Design strategies used in the pilot-scale and demonstration wetland treatment systems that can be applied to designing full-scale constructed wetland systems for treating waters ranging from fresh to saline include plant and soil selection, water-depth selection, addition of amendments, hydraulic retention time, and hybrid components. These strategies allow conditions within a wetland treatment system to be modified to achieve ranges necessary for the preferred biogeochemical treatment pathways. In the case of renovating a produced water containing constituents of concern that require different biogeochemical pathways for treatment, a constructed wetland system can be designed with sequential cells that promote different conditions. Removal rate coefficients determined from the pilot-scale constructed wetland experiments and confirmed by the demonstration wetland treatment system can be used to calculate hydraulic retention times required to treat constituents of concern in full-scale constructed wetland systems. The calculated hydraulic retention times can then be used to determine the surface area or "footprint" of a full-size constructed wetland treatment system for a given inflow rate.

#### INTRODUCTION

The project consists of three Phases, with each Phase corresponding to a Project Year. Phase I included an assessment of characteristics associated with produced waters. Phase II involved design, construction, and monitoring of a pilot-scale wetland treatment system. In Phase III, a demonstration system was designed and constructed, and performance in providing onsite treatment of produced water was measured. Rates of treatment in the demonstration system were determined under actual conditions, such as climate and variable water composition and volumes. Data were collected and analyzed for replicating the technology for full-scale treatment at other field sites. In addition, data were collected to compare the technology with other treatment options.

### **METHODS**

# Project Management Plan (Task 1)

A Project Management Plan (PMP) was completed by Clemson University in collaboration with DOE and approved during the first reporting period.

### Identify Chemical, Physical, and Risk Characteristics of Produced Waters (Task 2)

The approach in Task 2 involved characterizing produced waters to identify constituents that must be treated to achieve criteria for reuse or discharge. A literature review was conducted on natural gas produced water using published sources and databases from the United States Geological Survey (USGS) and DOE. The data, which include both analytical data and geological source information, were compiled into an Excel spreadsheet. A produced water inventory of these data was created, listing the produced water data by geologic basin and formation. To obtain data on produced waters, we contacted various potential data sources including state geological surveys, state oil and gas boards, and industry personnel.

The characterization included conventional natural gas produced water (NGPW), shale gas produced water (SGPW), coal bed methane produced water (CBMPW), and tight gas sand produced water (TGSPW). Data for some types of unconventional produced water (e.g. SGPW, TGSPW, etc.) can be scarce. Key data of produced water characteristics from shale gas wells were provided by a major producer of shale gas. However, we encountered other operators who were hesitant to provide produced water data from shale gas wells because of their concerns regarding regulatory issues and permitting of new shale gas wells. We received produced water samples from several field sites for analysis.

### Determine Reuse and Discharge Criteria and Develop Treatment Performance Goals (Task 3)

Post-treatment options for produced waters were assessed and compared by examining reuse and discharge criteria. These criteria for reuse were compiled from appropriate published sources,

including USEPA publications and NPDES (National Pollutant Discharge Elimination System) permits. Reuse criteria included those for livestock, irrigation, and discharge to receiving aquatic systems.

# Design and Construct Pilot-Scale Constructed Wetland Treatment Systems (Task 4)

Pilot-scale constructed wetland treatment systems were designed and constructed at Clemson University based on fundamental reactions and processes needed to treat (transfer and transform) the targeted constituents identified in the previous tasks. Four separate systems were constructed for representative constituents with specific water compositions selected based on the results of Tasks 2 and 3.

# Measure Treatment Performance of Pilot-Scale Constructed Wetland Systems (Task 5)

Performance from the four pilot-scale constructed wetland treatment systems designed and constructed during Task 4 was measured using sampling methods that allow for the determination of both the rate and extent of treatment. In conjunction with performance parameters, explanatory parameters were monitored to determine the conditions needed for efficient removal.

### Assess Pilot-Scale Performance (Task 6)

In Task 6, rate coefficients, extents, and efficiencies for pilot-scale treatment were calculated using data collected during Task 5. Measured performance parameters were compared with treatment goals to assess the success of each pilot-scale system.

# <u>Design and Construct Demonstration Constructed Wetland Treatment System (Task 7)</u>

Data from Task 6, including rate coefficients, extents, and efficiencies, were used in Task 7 to design a demonstration constructed wetland treatment system (CWTS). In addition, specific experiments in support of designing CWTSs were performed using pilot-scale systems to gain a better understanding of conditions affecting treatment pathways and hybrid design strategies that can be incorporated into constructed wetland treatment. The experiments included investigating the following: addition of zeolite for sorption, biogeochemical conditions and processes, effect of water depth, effect of seasonal variation, addition of a nutrient amendment, incorporation of an oil/water separator, and evapotranspiration.

The demonstration CWTS was constructed to treat CBMPW from the Black Warrior basin in central Alabama. The CWTS was designed with sequential oxidizing and reducing cells to treat constituents of concern, including ammonia, barium, cadmium, iron, manganese, nitrate, and selenium, identified through sample analyses.

# Measure Treatment Performance of Demonstration Constructed Wetland System (Task 8)

Performance of the demonstration wetland treatment system designed and constructed during Task 7 was monitored using sampling and analytical methods that allow for calculation of removal efficiency, removal rate, and removal extent of constituents of concern. Explanatory parameters were also measured to ensure conditions were suitable for effective treatment.

# Apply Results to Water Management and Technology Replication (Task 9)

Performance data from the pilot-scale and demonstration CWTSs were applied to evaluating the use of CWTSs at other sites. Suitability of treated waters for reuse or discharge was determined for different areas, and results and treatment costs using CWTSs were compared to other technologies for treating conventional and unconventional produced waters. Finally, the results were applied to developing water management plans and design.

# <u>Technology Transfer (Task 10)</u>

Technology transfer activities, including presentations and journal articles, were conducted to disseminate project results and findings.

### RESULTS AND DISCUSSION

# Project Management Plan (Task 1)

The revised Project Management Plan was competed, submitted to DOE, and approved during the first reporting period as scheduled.

### Identify Chemical, Physical, and Risk Characteristics of Produced Waters (Task 2)

Based on the data gathered, a wide range of chemical, physical, and risk characteristics occurs among the produced waters being investigated. Targeted constituents in the various produced waters include anions, cations (including metals), organics, dissolved solids, and suspended solids. Data on the water volumes produced are scarce in the sources investigated.

Many of the conventional gas produced water data were obtained from published sources for basins that co-produce both oil and natural gas. The compilation contains records for 635 samples of co-produced water from 36 geologic basins. Of these 635 samples, all have organics data and 223 have metals data.

The produced water data for unconventional gas include tight gas sand, shale, and coal bed formations. Data were compiled from 377 coal bed methane (CBM) records from several geologic basins, including Uinta, Black Warrior, Raton, Powder River, and Arkoma. Of these 377 samples, all have records for metals, 88 have records for organics, and 243 have records for

nitrogen-associated compounds (i.e. nitrate, nitrite, and ammonia). Data from 541 shale gas records were compiled from four basins; of these, 499 have metals data. Data from 137 tight gas sand records were compiled from eight basins; of these, one has metals data and one has organics data.

### Characterization

### Produced Water

Produced water is generated during processes such as fossil fuel extraction and fossil energy production. As water trapped in sediments and sedimentary rocks moves through the subsurface, it tends to accumulate a unique array of ionic constituents (Tucker, 2001). These constituents may then occur in waters produced with oil and gas. Based upon their origins, produced waters can be categorized into two different classes: conventional produced waters and unconventional produced waters. Conventional produced waters are generally considered to be waters that are produced during traditional fossil fuel extraction. Unconventional produced waters are waters produced during extraction of non-traditional fossil fuels or during industrial processes. Conventional produced waters include: oilfield produced water (OFPW), natural gas produced water (NGPW), and natural gas storage produced water (NGSPW). Unconventional produced waters include: coal bed methane produced water (CBMPW), tight gas sand produced water (TGSPW), shale gas produced water (SGPW), oil sand produced water, industry cooling produced water, and industry sluicing produced water. Some of these waters are brought to the surface during recovery of fossil fuels from underground deposits, and others have origins on the surface and are used by industry as process water or for internal cooling and sluicing operations for compliance with environmental regulations. Produced water can range from fresh to hypersaline; for further characterization, produced waters are compared to surface waters with regard to mean constituent concentrations (Table 1).

As global fresh water resources are depleting, consumptive use of fresh water is increasing, as well as costs associated with disposal of produced water. The notion of produced water as a valuable resource through reuse is becoming a more feasible and necessary practice. With adequate characterization and treatment of produced water, applications may be implemented for either internal reuse (e.g. cooling, sluicing, etc.) or external reuse (e.g. surface discharge, ground water recharge, irrigation, etc.) for consumptive and/or non-consumptive purposes. Consumptive reuse of produced water is defined as the reuse of water in a manner that removes it from a source and releases it in a different location that cannot recharge the original system. Non-consumptive reuse is defined as removing water from a source and returning it to the same source, thus replacing what was taken (Agarwal et al., 2000). Non-consumptive reuse is clearly a more desirable practice; however, if the water returning to the originating system is not adequately treated, the environmental repercussions to the receiving system may outweigh the benefit of the influx of water (Agarwal et al., 2000).

Produced waters contain both inorganic and organic constituents. The major inorganic constituents include chloride, sodium, calcium, magnesium, potassium, mercury, iron, lead, and selenium as well as many other metals, metalloids and non-metals. Produced waters can contain many organic constituents as well, such as: oil and grease, phenols, *n*-alkanes, and low

molecular-weight aromatic hydrocarbons such as benzene, toluene, ethylbenzene, and xylene (BTEX) (Veil et al., 2004). Depending on the designated reuse for the produced water, both inorganic and organic constituents may be targeted for treatment.

**Table 1.** Mean constituent concentrations (mg/L) for fresh, brackish, saline and hypersaline surface waters

	Fresh	Brackish	Saline	Hypersaline
Ca	$3.8^{1}$	$320^{7}$	$410^{2}$	16900 <sup>9</sup>
K	$0.6^{1}$	45 <sup>7</sup>	$389^{2}$	$7300^{9}$
Mg	4.5 <sup>1</sup>	135 <sup>7</sup>	$1303^{2}$	$40700^9$
Na	$23.0^{1}$	$460^{7}$	$10813^{2}$	$39200^9$
В	$0.01^{5}$	nd	$5.0^{3}$	nd
Cl	$32.0^{1}$	$1,140^{7}$	$19440^2$	$212400^9$
$HCO_3$	$19.0^{1}$	$100^{7}$	$143^{2}$	nd
$SO_4$	$50^{1}$	$730^{7}$	$2713^{2}$	nd
$NO_3$	$0.07^{1}$	nd	$0.6^{4}$	nd
Fe	$0.002^{1}$	nd	$0.003^2$	nd
F	$0.26^{5}$	3 <sup>7</sup>	$13^{6}$	nd
Se	nd*	$0.0001^{8}$	$0.00009^8$	nd
Zn	$0.007^{5}$	nd	$0.005^2$	nd

nd = not determined

### Chemical Characterization

Produced waters typically contain a range of concentrations of many constituents of concern (COCs), including chlorides, hydrocarbons, metals, and metalloids (Ray and Engelhardt, 1992; Johnson et al., 2008a). Chemical characteristics measured for produced water also include: total dissolved solids (TDS), pH, hardness, alkalinity, five-day biochemical oxygen demand (BOD<sub>5</sub>), chemical oxygen demand (COD), conductivity, and dissolved oxygen concentration (DO) (Table 2). The predominant constituents in produced water are dissolved salts (Fillo et al., 1992) expressed as salinity, conductivity, or TDS (Veil et al., 2004). The majority of these salts consist of sodium chloride; however, the ionic composition of produced water can range widely (Fillo et al., 1992). Cations found in produced water commonly include sodium, calcium, magnesium, and potassium; anions present often include chloride, sulfate and bicarbonate (Fillo et al., 1992). The inorganics (i.e. metals, metalloids, and nonmetals) present in produced water can include any or all of the following: barium, iron, manganese, potassium, strontium, lithium, zinc, cadmium, cobalt, copper, lead, mercury, chromium, nickel, and several others (Fillo et al., 1992). Organic constituents can be dissolved or dispersed in the produced water and belong to one of

<sup>1.</sup> Hecky and Kilham, 1973

<sup>2.</sup> Lyman and Fleming, 1940

<sup>3.</sup> Taniguchi, 2004

<sup>4.</sup> Wood et al., 1967

<sup>5.</sup> Wetzel, 2001

<sup>6.</sup> Warner, 1971

<sup>7.</sup> Amor et al., 2001

<sup>8.</sup> Zawislanski et al., 2001a

<sup>9.</sup> Nissenbaum, 1975

the following classes: aliphatic hydrocarbons, phenols, carboxylic acids (e.g. naphthenic acids), and low-molecular weight aromatic compounds (Stephenson, 1992). Dispersed organic constituents (e.g. oil and grease) often consist of small suspended droplets (Stephenson, 1992). Gases (e.g. carbon dioxide, hydrogen sulfide, methane) can be formed by chemical reactions within the produced water. Naturally occurring radioactive material (NORM) originates in some geological formations and can be brought to the surface with produced water. NORM in produced water includes radium-226 and radium-228, which are derived from the radioactive decay of uranium and thorium associated with formations in the hydrocarbon reservoir (Utvik, 1999). As the water approaches the surface, temperature changes can cause radioactive elements to precipitate. The resulting scales and sludges may accumulate in water separation systems (Veil et al., 2004).

In addition to chemicals that occur naturally in produced water, chemicals may be added during production processes. These production chemicals are added to treat or prevent operational problems (e.g. scaling, biofouling, and corrosion). Production treatment chemicals include scale inhibitors, corrosion inhibitors, biocides, emulsion breakers, and water treatment chemicals (coagulants/flocculants) (Stephenson, 1992).

### Physical Characterization

Physical characteristics of produced water include color, turbidity, temperature, and odor. The physical characteristics of produced water are often due to reservoir geology, hydrocarbon composition, geographical location, and water injection history (VWS, 2009).

# **Biological Characterization**

Characterization of a specific produced water for a potential reuse may include a biological component as well. Biological characterization would include questions such as: "is it toxic?" and "how toxic is it?" These two questions of aquatic life sustainability are answered in different ways. To determine if toxicity exists in a specific water, a range-finding toxicity experiment can be conducted to determine a range of concentrations of COCs that elicit a response in a test organism with regard to an exposure (Table 3). Once the range of critical concentrations has been narrowed, a toxicity identification evaluation (TIE) may be conducted to discern the specific concentrations at which selected COCs are toxic. Three parameters are typically measured in toxicity testing: mortality, growth, and reproduction.

There are several methods of aquatic and terrestrial toxicity testing that can evaluate the ability of a specific water to support a particular organism and its survival, growth, or reproduction. Phytotoxicity evaluation is of special concern in areas where fresh water is scarce and toxic constituents in a water may prohibit its use for irrigation. Phytotoxicity testing evaluations can be conducted in a fashion similar to the aquatic animal toxicity evaluations, with the exception that seed germination or sustained crop growth over time is the toxicity identifier instead of mortality or failure of either growth or reproduction.

 Table 2. Analytical methods

Parameter	Method	Detection Limit
Turbidity	Closed reflux colorimetery (Standard Methods: 2130 B)	0.1 NTU
pН	Direct Instrumentation: Orion Model 420A	0.01 S.U.
Conductivity	Direct Instrumentation: YSI 30	0.1 μS/cm
Alkalinity	Standard Methods: 2320 B	2 mg/L as CaCO3
Hardness	Standard Methods: 2340 B	2 mg/L as CaCO3
$DO^1$	Direct Instrumentation: YSI Model 52	0.1 mg/L
$COD^2$	Closed reflux colorimetery (HACH- modified from Standard Methods: 5220D)	3 mg/L
$BOD_5^3$	Standard Methods: 5210 B	0.1 mg/L
$TSS^4$	Standard Methods: 2540 D	0.1 mg/L
TDS <sup>5</sup>	Standard Methods: 2540 C	0.1 mg/L
Ammonia	Direct Instrumentation: Orion Model 95-12 (Standard Methods: 4500-D)	0.03 mg/L
Nitrate	Standard Methods: HACH-modified from Standard Methods: 4500-NO3	0.01 mg/L
Chloride	HACH tritration method	10-10,000 mg/L
Toxicity	Ceriodaphnia dubia and Pimephales promelas	
Oil and grease	EPA Method 1664 A (Environmental Express StepSaver Modification)	~5 mg/L
		(mg/L)
Barium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0023
Iron	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0062
Manganese	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0014
Nickel	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.015
Arsenic	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.053
Chromium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0061
Cobalt	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.007
Copper	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0054
Lead	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.042
Mercury	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0025
Silver	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.007
Cadmium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0034
Selenium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.075
Magnesium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.03
Tin	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.025
Aluminum	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.045
Zinc	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0018
Sodium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.029
Calcium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.03
Potassium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.7
Boron	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.057
Lithium	Inductively Coupled Plasma-Atomic Emissions Spectrometry (ICP-AES): 200.7	0.0037

(USEPA 1991; USEPA, 1994; APHA 2005)

**Table 3.** Summary of toxicity measurements for *Ceriodaphnia dubia, Daphnia magna*, and *Pimephales promelas* for elements or compounds present in non-traditional waters.

Constituent	Species	Experiment	Effect	NOEC (mg/L)	LOEC (mg/L)	LC <sub>50</sub> (mg/L)	IC <sub>50</sub> (mg/L)	Reference
Aluminum	Cd	48h	Mort			2.88		6,10
Aluminum	Pp	96h	Mort			4.25		11
Calcium	Cd	48h	Mort			1830		11
Calcium	Pp	96h	Mort			4630		11
Magnesium	Cd	48h	Mort			880		11
Magnesium	Pp	96h	Mort			2120		11
Sodium	Dm	48h	Mort			1820		7,10
Sodium	Pp	7d	Mort		8000			11
Chloride	Cd	7d	Mort			1042		8
Chloride	Pp	7d	Mort		8000			11
Bromine	Dm	48h	Mort		0.46			9,10
Bromine	Pp	7d	Mort			0.78		11
Boron	Cd	48h	Reprod		18			11
Lithium	Cd	7d	Mort			0.92		11
Lithium	Pp	26d	Mort			1.2		11
Barium	Cd	48h	Mort			13.5		7,10
Manganese	Cd	7d	Reprod				9	1
Manganese	Pp	8d	Mort			38.5		11
Iron	Cd	48h	Mort			30.06		11
Iron	Pp	96h	Mort			2.54		11
Potassium	Cd	48h	Mort			630		11
Potassium	Pp	96h	Mort			880		11
Copper	Cd	48h	Mort			0.027		3
Copper	Cd	7d	Reprod	0.012	0.032	0.027		2
Copper	Pp	96h	Mort	0.012	0.002	0.49		5
Zinc	Cd	48h	Mort			0.101		11
Zinc	Pp	96h	Mort			0.87		5
Fluoride	Dm	48h	Mort			304		12
Fluoride	Pp	96h	Mort			205		11
Nitrate	Cd	48h	Mort			374		4
Nitrate	Рp	96h	Mort			1341		4
Phosphate	Pp	96h	Mort			18		11
Strontium	Dm	48h	Mort			125		7,10
Sulfate	Cd	48h	Mort			1770		11
Sulfate	Pp	96h	Mort			2820		11
Lead	Cd	24h	Mort	0.051	0.099	2020		11
Arsenic	Cd	48h	Mort	0.051	0.077	29.1 μΜ		11
Arsenic	Pp	96h	Mort			9.9		11
Selenium	Pp	96h	Mort			1.0		11
Chromium	Pp	7d	Growth		6.4	1.0		11
Chromium	Рр Рр	7a 96h	Mort		0.4	37		11
Mercury	гр Cd	48h	Mort			.05 μM		11
Mercury	Pp	48h	Mort			.03 μlVI 0.05		11
Cobalt	Рр Cd	7d	Mort	0.05		0.03		11
				0.03		4.014		
Cobalt	Cd	24h	Mort			4.014		11

Cobalt	Pp	96h	Mort		12.6	11
Cobalt	Pp	7d	Mort	1.232		11
1. Lasier et al.	, 1999.		5. Buhl, 1	998.	9. Dorman et al., 2010.	
<ol><li>Carlson et a</li></ol>	1., 1986.		<ol><li>Soucek</li></ol>	et al., 2001.	10. EPA EcoTox, 2009.	
3. Toussaint, e	t al., 1995.		<ol><li>DeGrae</li></ol>	eve et al., 1992.	11. Fieser et al., 1986.	
4. Scott and Ca	runkilton, 200	0.	8. LeBlan	ic, 1980.	<ol><li>Biesinger and Christer</li></ol>	nsen, 1972

### Produced Water Types

#### Shale Gas Produced Water

SGPW characterization data from several basins were compiled (Table 4). Shale gas produced water has chloride concentrations that range from freshwater (<5,000 ppm) to hypersaline (>50,000 ppm). Metals (e.g. aluminum, iron, and barium) are present in the well prior to fracturing of the shale; other metallic and metalloid constituents (e.g. cobalt, chromium, molybdenum, nickel, vanadium, and arsenic) in shale gas produced water can originate in the fracturing fluids (Sumi, 2008).

### Coal-bed Methane Produced Water

Coal bed methane (CBM) may be produced from water-saturated coal seams. This process requires dewatering of coal in order to mobilize methane (Rice and Nuccio, 2000). The water extracted from the formation is brought to the surface as CBMPW (Rice and Nuccio, 2000; Veil et al., 2004). Because of the amount of water generated during dewatering and the rapid increase in CBM exploration and production, management of these waters is a concern (Nelson, 1999; Rice and Nuccio, 2000). Some of the major CBM producing basins include the Powder River Basin in Montana and Wyoming, the Raton Basin in Colorado and New Mexico, the Black Warrior Basin in Alabama, and the Uinta Basin in Utah (Rice and Nuccio, 2000; Benko and Drewes, 2008). The chemistry of the waters produced may vary due to the depth and depositional environment (Rice and Nuccio, 2000). CBM waters typically contain dissolved inorganic elements and dissolved organic compounds (Rice and Nuccio, 2000; Benko and Drewes, 2008).

To identify the water characteristics of CBMPW, several sources were reviewed including papers by the USGS on the Powder River Basin, the Arkoma Basin, and the Ferron Sand of the Uinta Basin, as well as CBMPW water chemistry reports from the Colorado Oil and Gas Conservation Commission (COGCC), the Geological Survey of Alabama (GSA), and the Montana Groundwater Information Center (GWIC).

In total, 377 records of CBMPW composition exist in our compilation consisting of data from the Powder River, Uinta, Raton, Black Warrior, and Arkoma basins. The data are sufficient to be used to identify specific COCs for CBMPW based on specific reuse criteria. The characteristics include general water chemistry parameters such as pH, temperature, conductivity, TDS, TSS, alkalinity, and hardness, as well as major cations and anions, trace metals, nitrogen species, and organic carbon. Statistical analyses have been conducted on the data to determine the range and

mean of each parameter (Table 5).

CBMPW production quantity data were recorded for each basin. The Powder River basin yields approximately 370,000 cubic meters per day (m³/d) of produced water, the Uinta Basin produces approximately 42,000 m³/d, the Raton Basin produces approximately 19,000 m³/d, and the Black Warrior Basin produces approximately 27,000 m³/d (Rice and Nuccio, 2000; Benko and Drewes, 2008). Water production rates from individual wells may vary greatly based upon drilling methods, geologic location, and age of the well (Rice and Nuccio, 2000; Veil et al., 2004; Benko and Drewes, 2008).

### Oil Field Produced Water

In addition to natural gas produced waters, constituents of some specific oilfield produced waters (OFPWs) were examined. The data used were provided by Chevron and are pertinent to Phases II and III of the project. Constituents in OFPW depend on depositional environment of the reservoir, trapping mechanism, and adjacent rock units within the basin. Produced waters from oil fields may contain groundwater or seawater that is injected to maintain reservoir pressure (Veil et al., 2004). Data included concentrations for cations and anions, oil and grease, specific polycyclic aromatic hydrocarbons (PAHs), and general water quality parameters (pH, total suspended solids, total dissolved solids, alkalinity, conductivity, dissolved oxygen, and hardness) (Table 6).

**Table 4.** Characterization of shale gas produced water (mg/L except as indicated) (n=541)

Constituents	Mean	Range
Temperature (°C)	25	
pH (S.U.)	6.124	1.21-8.36
Alkalinity		160-188 (total as CO <sub>3</sub> )
TOC		.05-10%
Salinity	190392	19-337900
TDS	76130.6	1724-228971
TSS		200-15,000
Al	126.5	0-5290
В	0.45	0.12-2.18
Ba	318.09	0-4370
Br	684.76	0-10600
Ca	14856	0.65-83950
Cl	84273	48.97-212700
Cu	0.795	0-15
F	3.06	0-33
Fe	137.8	0-2838
$HCO_3$	244.36	0-4000
I	20.75	0.8-105
K	1091.99	0.21-5490
Li	39.39	0-611
Mg	1830.6	1.08-25340
Mn	7.73	0-96.5
Na	34672.2	10.04-204302
$NO_3$	226.1	0-2670
$PO_4$	0.276	0-5.3
$SiO_2$	52.57	0.03-5020
$SO_4$	241.43	0-3663
Sr	594.23	0.03-13100
Zn * As CaCOa	1.43	0-20

<sup>\*</sup> As CaCO<sub>3</sub>

(McIntosh et al., 2002; Hill, 2002; GPRI, 2003; Crowe, 2007; Burnett, 2007; Dodds, 2008; Sumi, 2008; Hill et al., 2008); Carter (PA Geol. Surv., pers. comm., 2009)

**Table 5.** Characterization of coal bed methane produced water

Constituent	Units	Range	Mean
pH	S.U.	6.56-9.87	-
Temperature	°C	1.2-41.7	18.71
Conductivity	mS/cm	94.8-145000	4419.57
Total Dissolved Solids	mg/L	270.18-114000	5197.25
Alkalinity	mg/L as CaCO3	54.9-9450	1858.84
Ba	mg/L	< 0.01-190	7.04
Br	mg/L	< 0.002-300	37.8
Ca	mg/L	0.8-5870	53.63
Cl	mg/L	0.7-70100	845.96
F	mg/L	< 0.05-15.22	2.52
Fe	mg/L	< 0.002-220	2.74
K	mg/L	0.3-186	10.63
Mg	mg/L	0.2-1830	34.59
Na	mg/L	8.8-34100	1183.08
Si	mg/L	<0.1-49.8	9.9
Sr	mg/L	0.032-565	16.08
Sodium Absorption Ratio	-	5.7-32	11.67
Ammonia	mg/L	1.05-59	3.53
Nitrate	mg/L	< 0.002-18.7	1.22
Phosphate	mg/L	< 0.050-1.5	0.23
Sulfates	mg/L	< 0.01-5590	335.14
Total Organic Carbon	mg/L	1-100	9.50
Ag	μg/L	< 0.5-375	63.10
Al	μg/L	0.2-1240	201.4
As	μg/L	0.1-60	3.21
В	μg/L	1.6-2400	151.36
Be	μg/L	< 0.1	< 0.1
Bi	μg/L	19-32	24.45
Cd	μg/L	<0.1-10	4.75
Ce	μg/L	<10.0	<10.0
Co	μg/L	< 0.1-0.729	0.17
Cr	μg/L	<1.0-53	11.49
Cs	μg/L	<0.1-0.78	0.18
Cu	μg/L	<0.2-60	8.75
Hg	μg/L	< 0.005-0.4	0.21
La	μg/L	<10.0	<10.0
Li	μg/L	0.21-6880	146.25
Mn	μg/L	<2.0-5400	59.12
Mo	μg/L	<0.5-100	28.84
Ni	μg/L	0.304-203	19.71
P	μg/L	<50-94	84.0
Pb	μg/L	<0.1-585	117.67
Rb	μg/L	4.1-38.2	11.79
Sb	μg/L	<0.1-950	575.0
Sc	μg/L μg/L	<0.1-3	1.34
50	μg/L μg/L	<0.1-70	7.44

Sn	μg/L	< 0.1-680	89.58
Th	μg/L	<20.0	<20.0
Ti	μg/L	<1.0-45	12.71
Tl	μg/L	<0.1-0.34	0.34
U	μg/L	< 0.5-50	12.82
V	μg/L	0.19-59	13.24
W	μg/L	<20.0	<20.0
Y	μg/L	<10.0	<10.0
Zn	μg/L	0.02-590	39.96
Zr	μg/L	0.21-131	27.68

(Hunter and Moser, 1990; Rice, 1999; Rice et al., 2000; ESN Rocky Mountain, 2003; Thordsen et al. 2007; Montana GWIC, 2009)

**Table 6.** Characterization of oil field produced water. n = number of samples.

Parameter	Mean (mg/L)	Minimum (mg/L)	Maximum (mg/L)	n
Alkalinity (mg/L as CaCO <sub>3</sub> )	326.2	300	380	13
Barium	4.1	0.07	7.4	7
Bicarbonate	704.7	433	976	2
Calcium	164.19	2.5	300	20
Carbonate	14.6	14.6	14.6	1
Chloride	12.33	5	48	15
Conductivity (µmhos)	1153.5	838	1469	2
Iron (III)	54.9	24.4	171	5
Iron (soluble)	0.14	0.1	0.5	13
Hardness (mg/L as CaCO <sub>3</sub> )	10.9	5	20	13
Magnesium	3.7	1.05	8.7	7
Manganese	6.3	1.4	8.1	5
Nickel	7.7	2.7	9.5	5
Nitrate	1.5	1	2	2
Oil and grease	92.0	92	92	1
Dissolved oxygen	0.00014	0.0001	0.0002	13
pH (standard units)	7.78	7.73	8.34	15
Potassium	10.5	1.6	42.6	7
Silicon	6.0	1.4	13.2	7
Sodium	115.3	8.8	339	7
Strontium	1.3	0.05	2.2	7
Sulfate	2.5	2	3	2
TDS	1037.0	704	1370	2
Tin	0.7	0.5	1.3	5
Zinc	12.8	6.3	17.4	5

### Conventional Natural Gas Produced Water

Conventional natural gas is extracted from underground formations typically comprised of permeable sandstone, and can be produced from other rock formations (e.g. limestone). Conventional natural gas accounts for approximately 61% of total natural gas production in the nation (EIA, 2008). During the extraction process, large quantities of formation water may be

brought to the surface as NGPW. Because of the extensive nature of conventional natural gas production and the inherent water production, the management of these NGPWs presents a challenge. Characteristics of NGPWs are listed in Table 7.

# Tight Gas Sand Produced Water

Tight gas sand is a low-permeability (less than 0.1 millidarcy) reservoir requiring natural and/or induced well stimulation for acceptable rates of gas flow (Kuuskraa 2004). Low-permeability reservoirs have the characteristics of small pore throats, strong water wettability, and strong capillary inhibition effect; thus they are very susceptible to damage during drilling, completion, production, and stimulation, which can lead to low production rates (Bennion et al. 1995). Chloride concentrations from the 137 records of TGSPW correspond to brackish to saline waters. Mean, maximum, and minimum concentrations of constituents are listed in Table 8.

**Table 7.** Characteristics of conventional natural gas produced water (units are same as in Table 5)

Min	Max	Constituent	Min	Max
4.4	7.0	Iron <sup>1</sup>	bdl	1100
3.1	6.47	Iron <sup>2</sup>	39	680
4200	180000	Lead <sup>2</sup>	< 0.2	10.2
136000	586000	Lithium <sup>2</sup>	18.6	235
0	285	Magnesium <sup>1</sup>	0.9	4300
2600	310000	Magnesium <sup>2</sup>	1300	3900
139000	360000	Manganese <sup>1</sup>	0.045	6.5
8	5484	Nickel <sup>1</sup>	bdl	0.02
75	2870	Nickel <sup>2</sup>	< 0.08	9.2
2600	120000	Potassium <sup>2</sup>	149	3870
bdl	0.4	Silver <sup>2</sup>	0.027	7
< 0.50	83	Sodium <sup>1</sup>	520	45000
0.004	1	Sodium <sup>2</sup>	37500	120000
< 0.005	1.51	Strontium <sup>1</sup>		6200
bdl	26	Sulfate <sup>2</sup>	<1.0	47
9.65	1740	Sulfide <sup>1</sup>	bdl	19
bdl	56	$Tin^1$	bdl	1.1
150	1149	$Zinc^1$	bdl	0.022
bdl	0.015	$Zinc^2$	< 0.02	5
< 0.02	1.21	$TOC^1$	67	38000
bdl	25000	Surfactants <sup>2</sup>	0.08	1200
9400	51300	Benzene <sup>1</sup>	1.8	6.9
1400	190000	Benzene <sup>3</sup>	< 0.01	10.3
81500	167448	Toluene <sup>1</sup>	0.857	3.37
bdl	0.03	Toluene <sup>3</sup>	< 0.01	18
bdl	0.02	Oil/Grease <sup>1</sup>	6	60
< 0.02	5	Oil/Grease <sup>2</sup>	2.3	38.8
	4.4 3.1 4200 136000 0 2600 139000 8 75 2600 bdl <0.50 0.004 <0.005 bdl 9.65 bdl 150 bdl <0.02 bdl 9400 1400 81500 bdl bdl	4.4 7.0 3.1 6.47 4200 180000 136000 586000 0 285 2600 310000 139000 360000 8 5484 75 2870 2600 120000 bdl 0.4 <0.50 83 0.004 1 <0.005 1.51 bdl 26 9.65 1740 bdl 56 150 1149 bdl 0.015 <0.02 1.21 bdl 25000 9400 51300 1400 190000 81500 167448 bdl 0.03 bdl 0.02	4.4         7.0         Iron¹           3.1         6.47         Iron²           4200         180000         Lead²           136000         586000         Lithium²           0         285         Magnesium¹           2600         310000         Magnesium²           139000         360000         Manganese¹           8         5484         Nickel¹           75         2870         Nickel²           2600         120000         Potassium²           bdl         0.4         Silver²           <0.50	4.4         7.0         Iron¹         bdl           3.1         6.47         Iron²         39           4200         180000         Lead²         <0.2

bdl = Below detection limit

<sup>&</sup>lt;sup>1</sup>Fillo and Evans, 1990

<sup>&</sup>lt;sup>2</sup>USEPA, 2000

<sup>&</sup>lt;sup>3</sup> Shepard et al., 1992.

**Table 8.** Characterization of tight gas sands produced water (mg/L except as indicated) (n=137)

Constituent	Mean	Range
pH (S.U.)	6.79	5-8.6
Hardness	780	
Conductivity	24.42	
Alkalinity	1424	
Temp (°C)	22	
Ammonia	2.74	
Bicarbonate	666.9	10-4040
Sulfate	701.98	0-8823
TDS	130393	1588-343300
TSS	85.7	
Oil/Grease	42.0	
Al	bdl	
As	0.17006	
Ba	0.30942	
Ca	14342.6	3-74185
Cd	0.03689	
Cl	79187	52-216000
Cr	0.26467	
Cu	0.53862	
Fe	0.01493	
K	983.8	5-2500
Mg	1661.5	2-8750
Mn	0.52457	
Na	33157.7	648-80000
Ni	0.12277	
Pb	0.48793	
Se	bdl	
Zn	0.07615	

Determine Reuse and Discharge Criteria and Develop Treatment Performance Goals (Task 3)

### Internal Reuse

### Reinjection

Reinjection into an underground formation is the most commonly used approach for managing produced water (Veil et al., 2004). Although some produced water is injected solely for disposal, most produced water (71%) is injected to maintain reservoir pressure and to hydraulically drive oil or gas toward a producing well. The produced water re-injection criteria focus on the concentration of halogenated and non-halogenated solvents decreased to less than 10% by volume of produced water as well as criteria for metals, metalloids, TSS, TDS, and dissolved oxygen (USEPA 2002a). Physical characteristics are not outlined in the re-injection criteria guidelines. Re-injection standards are governed by the Clean Water Drinking Act (CWDA) and the Underground Injection Control program (UIC).

### **Industrial Cooling**

According to the U.S. Geological Survey (Hutson et al. 2004), the U.S. power industry withdrew about 136 billion gallons per day of fresh water for power plant cooling; conventional surface and ground water resources are no longer sufficient to meet increasing power plant needs. Produced water represents a large-volume source of water that could potentially serve as makeup water for a power plant (DiFilippo, 2004). Water used for cooling operations should have a low TDS concentration; otherwise, scaling, corrosion, and biofouling of the cooling system may become problematic (WGA, 2006).

# **Industrial Sluicing**

Bottom ash contains non-combustible constituents with traces of combustibles produced during operation of a coal-, oil-, or gas-burning furnace (Sell, 1992; EIA 1995). Ash sluicing systems pump produced water to sluice the bottom ash to waste storage (Prewitt, 1995). To prevent problems with scaling and corrosion of the bottom ash system, water used for industrial sluicing must have low TDS and chloride concentrations.

### External Reuse

# Aquaculture

The physical, biological, and chemical reuse criteria for aquaculture are stringent. Water quality determines how well fish will grow and survive in an aquaculture operation (Buttner et al., 1993). Physical and chemical characteristics (e.g. DO, temperature, ammonia concentration) are commonly associated with fish kills, while other characteristics (e.g. pH, alkalinity, hardness, turbidity) can affect the health of a fish population, but are not consistently found to be toxic to the fish (Buttner et al., 1993). Biological characteristics for aquaculture are measured by examining growth, reproduction, and mortality of the aquatic organisms within an aquaculture environment (i.e. laboratory toxicity evaluation).

### Livestock

Water for livestock is an option for reuse of produced water, provided that the COC concentrations are below the EPA water quality criteria for livestock (API, 2004). Chemical analyses can be conducted to determine the COCs and discern which constituents should be targeted for treatment and/or removal. Toxicity tests can be conducted to determine if a specific produced water is toxic, and if so, how toxic it is. Using organisms such as *Ceriodaphnia dubia* and *Pimephales promelas*, which are sensitive indicator species, can facilitate determination of toxicities from COCs.

# Drinking

Drinking water quality criteria are stringent. If produced water were to be reused in this capacity, successful and simultaneous treatment of COCs would likely be necessary. However, achieving drinking water quality with produced water, especially in more arid regions, is desirable. The extent of treatment needed to achieve drinking water standards may vary depending on the type and characteristics of produced water. In order to analyze treatment system effluent to determine if drinking water criteria have been met, a full chemical characterization must be conducted. For evaluation of the specific effluent for potential use as drinking water, a biological and physical characterization must also be conducted, but water that is non-toxic physically or biologically does not always indicate that the water reached drinking water criteria, and vice versa.

## Irrigation

Irrigation presents a feasible reuse option for produced water. Salinity, sodicity, and toxicity are the critical parameters when considering reuse of a specific water for irrigation purposes (Veil et al. 2004). The salinity can be estimated by measuring chloride concentration in the sample. Biological characterization involves phytotoxicity tests which use seed germination or sustained crop growth over time as an identifier of toxicity. The seed germination or sustained growth test should involve a plant species that is analogous to a crop that may be irrigated with the specific water. A physical characterization focuses on temperature, total suspended solids (TSS), and turbidity.

# Groundwater Recharge

There are specific considerations for characterization of water to be reused for groundwater recharge. Because proximity to the surface is an important factor when determining water quality criteria, physical characterization must include depth of the aquifer to be recharged. With chemical characterization, it is important to examine local mineralogy of the aquifer. Also, certain purposes involved with groundwater recharge call for specific characterizations. For example, if the purpose is to establish a saltwater intrusion barrier in coastal aquifers, then chloride concentration should be considered.

# Surface Water Discharge

Discharging produced water to an existing surface water system presents stringent treatment goals similar to drinking water standards. In accordance with the Clean Water Act (CWA), all point source discharges to federal waters from municipal and industrial facilities must be permitted through the National Pollutant Discharge Elimination System (NPDES). The primary goal of the NPDES is to protect the health of the nation's surface waters by regulating the discharge of contaminants that may present a risk. Regulation is enforced primarily by setting effluent limitations.

NPDES permits are obtained either directly from the U.S. Environmental Protection Agency (USEPA) or from within states that have been given primary enforcement responsibilities (primacy) by the USEPA. Permits are based either on general permitting guidelines established

for specific industries located in a specified geographic area, or may be specific to a facility. The permitting process involves limits based upon both the potential impacts to the receiving system (water quality standards) as well as technology-based limitations for treating the water defined as effluent limitation guidelines (ELGs). In the absence of applicable ELGs for discharge water, the permit writer may be authorized to use best practical judgment to set an effluent limit. The final permit will include the most stringent limitations established by either the water quality standards or the ELGs including any values determined by best practical judgment.

Water quality standards may be determined using three different methods. In the chemical-specific approach, chemical analyses are performed on the specific discharge water and the receiving system to determine site-specific potential chemical impacts. In the whole effluent toxicology (WET) approach, the impacts of the discharge water are evaluated by testing the acute and chronic effects of exposure to aquatic organisms. In the biological assessment approach, the biological diversity and integration of a receiving system may be compared to a healthy system to determine the impacts of discharge.

ELGs for oil and gas extraction point source wastes including produced waters may be found in the Code of Federal Regulations (CFR) under 40 CFR Part 435. The ELGs are divided into different categories based upon the level of technology required to meet the guidelines including best conventional pollution control technology (BCT), best practical control technology currently available (BPT), best available technology economically achievable (BAT), and new source performance standards (NSPSs). According to 40 CFR Part 435, oil and grease is the only ELG available. Table 9 lists the levels for oil and grease required by the produced water ELGs based upon the technology levels. Existing dischargers of produced water must meet BCT, BPT, and BAT ELGs while new dischargers must meet the NSPS ELGs. In the case of oil and grease, the most stringent levels found under the BAT and NSPS ELGs are the same and therefore all discharges of produced water must meet the oil and grease ELG of 42 ppm maximum and 29 ppm 30 day maximum.

**Table 9.** Oil and grease effluent limitation guidelines (ELGs) for produced waters

Technology Level	Maximum Level ppm	30 Day Maximum ppm*
BCT	72	48
BPT	72	48
BAT	42	29
NSPS	42	29

<sup>\*</sup>Concentrations of oil and grease may not exceed this value for more than 30 consecutive days

# Produced Waters and Reuse Purposes

Reuse criteria for produced waters are listed in Table 10. Specific reuse purposes can be identified for types of produced waters based on the geographic areas of production and water needs associated with those areas.

<sup>40</sup> CFR Part 435

**Table 10.** Selected Reuse Criteria for Produced Waters. From Ayers and Westcot (1994), WDEQ (2001), USEPA (2004), Johnson et al. (2008a)

Livestock and Surface Drinking Parameter<sup>1</sup> Irrigation Aquatic Life Wildlife Discharge<sup>2</sup> Water<sup>3</sup> Conductivity (ms/m) 1700-13000 16000 TDS 500 500 4.5-9.0 6.5-8.5 6.5-9.5 6.0 - 9.0pH (range) (S.U.) 6.5-8.5 SAR (ratio) 15 Bicarbonate 520 Carbonate 3 Sulfate 960 250 Sulfide 0.21 - 1Nitrite-N 10 1 Nitrate-N 10 10 5 Ammonia-N Total Nitrogen-N 100 2 Phosphate-P COD 170-292 35 Oil and Grease 35 15 5 5 0.1 .05-.2 Al Ag 0.1 As 0.1 0.2 0.15 0.01 В 0.5-15 5 5 2 Ba 154 0.1 Be 0.1 0.004 400 Ca 0.005 Cd 0.01 0.05 0.002 Cl 1050 5000 250 Co 0.05 1 Cr 0.1 1 0.0001 - 0.10.1 Cu 0.2 0.5 0.01 0.37-1.0 1 2 F 1 2 Fe 5 1 0.6-9.4 0.3 0.01 0.004 0.002 Hg 2 K 2.5 Li 60 Mg 0.2 0.05 1.5 1-4.4 0.05 Mn Mo 0.01 920 Na Ni 0.2 2 Pb 5 0.1 2.5 0.4 0.015 Sb 0.006 Se 0.02 0.05 0.005 0.05 0.002 Th V 0.1 0.1 \_ 2 24 5 Zn 0.1 0.4-5

<sup>1.</sup> All units are mg/L unless noted

<sup>2.</sup> Values obtained from NPDES permits used in the oil and gas industry

<sup>3.</sup> USEPA secondary standards selected when available

### Shale Gas Produced Water Reuse

Shale gas is extracted in many areas throughout the United States (e.g. New York, Pennsylvania, Arkansas, Alabama, Texas, and Montana). Shale gas produced water has a variety of potential reuses, including surface discharge and irrigation. Reuse options were determined by location (e.g. latitude, longitude and altitude) and socio-economic needs. Surface water discharge to lakes and rivers after sufficient treatment may be viable in arid regions (e.g. Texas and New Mexico). Irrigation may be a feasible reuse option where suitable water supply is limited because of drought or pollution (Frankenberger and Engberg 1998). Reinjection is commonly used for disposal of SGPW rather than treatment. However, with the increasing need for fresh water sources, treating SGPW for reuse is expected to become more common.

Constituents of concern in SGPW for surface water discharge were identified by comparing the concentrations in SGPW with NPDES surface water discharge permit criteria (Table 11). The COCs are: total suspended solids, total dissolved solids, pH, aluminum, barium, boron, chloride, copper, fluoride, iron, manganese, phosphate, sulfate, and zinc. SGPW COCs for irrigation were determined by comparing the concentrations of constituents of SGPW to USEPA and Food and Agriculture Organization of the United Nations (FAO) irrigation water criteria (Table 12). These COCs are: total suspended solids, total dissolved solids, pH, aluminum, bicarbonate, boron, calcium, chloride, copper, fluoride, iron, lithium, magnesium, manganese, nitrate, phosphate, potassium, sodium, sulfate, and zinc.

### Coal Bed Methane Produced Water Reuse

Beneficial reuse of CBMPW is an attractive option due to: 1) the large quantity of PW generated during coal bed methane extraction; 2) the need for water reuse in coal bed methane producing areas; and 3) the rapid increase in coal bed methane extraction development (Rice and Nuccio 2000). Reuse options may include irrigation, livestock, recreation, aquaculture, industrial cooling and sluicing, wildlife habitat, aquifer recharge, and drinking water (ALL Consulting 2003; Veil et al. 2004). Specific needs may be based on geographic location, climate, water availability, and development. For instance, in rural areas with limited freshwater resources and an adequate growing season, reuse options most likely will include irrigation and livestock rather than industrial reuse.

Based on the locations and climates of the major CBM producing areas, reuse options for CBMPW include irrigation, livestock watering, and aquaculture (Tables 13-15). The selection of multiple guideline sources, including international, federal, and state agencies, promotes adequate determination of reuse guideline levels. Guidelines from Wyoming were selected because the Powder River Basin in Wyoming is responsible for a major portion of CBMPW production (Rice and Nuccio, 2000).

Based on the comparison of CBMPW characterization data to reuse guidelines, the following parameters are considered COCs: conductivity, pH, SAR, bicarbonate, carbonate, sulfate, nitrate, ammonium, Al, Ba, Ca, Cd, Cl, Cu, F, Fe, K, Li, Mg, Mn, Mo, Na, Ni, Pb, Se, and Zn. The performance of the selected treatment method will be maximized so that the most reuse options may be considered.

**Table 11.** NPDES permit criteria and shale gas produced water characteristics for identification of COCs. From USEPA (2004). (All units are mg/L unless noted).

Constituents	Shale Gas Pr	oduced Water	NPDI	ES	COCs
	Min	Max	Min	Max	
Total Suspended Solids	200	15000		45	Yes
Total Dissolved Solids	1724	228971	30	5000	Yes
pH (S.U.)	1.21	8.36	>6	>9	Yes
Temperature °C	25			>40	No
Alkalinity (as CaCO <sub>3</sub> )	160	188			-
Dissolved Oxygen			6	17	-
Salinity	19	337900			-
Aluminum	0	5290		0.05	Yes
Ammonium-N					-
Arsenic				0.05	-
Barium	0	4370		154	Yes
Bicarbonate	0	927			-
Boron	0	4370		1	Yes
Bromide	0	10600			-
Cadmium				0.0025	-
Calcium	0.65	83950			-
Carbonate					-
Chloride	48.9	212700		5000	Yes
Chromium			0.1	0.2	-
Cobalt					-
Copper	0	15	0.37	1	Yes
Fluoride	0	33		30	Yes
Iodine	0.8	105			-
Iron	0	2838	0.6	9.4	Yes
Lead			0.0056	0.015	-
Lithium	0	611			-
Magnesium	1.1	25340			-
Manganese	0	96.5	0.05	4.4	Yes
Mercury			0.0008	0.004	-
Molybdenum					-
Nickel			0.052	2	-
Nitrate-N	0	2670			-
Phosphate-P	0	5.3		1	Yes
Potassium	0.21	5490			-
Selenium			0.02	0.071	-
Silicon	0.03	5020			-
Sodium	10	204302			-
Strontium	0.03	13100			-
Sulfate	0	3663		3000	Yes
Vanadium					-
Zinc	0	20	0.005	5	Yes

**Table 12.** FAO irrigation criteria and shale gas produced water characteristics for identification of COCs. All units are mg/l unless noted.

Constituents	Shale Gas Pro	oduced Water	FAO Irrigat	ion Criteria	COCs
	Min	Max	Long	Short	
Total Suspended Solids	200	15000	< 50 <sup>a</sup>	>100 <sup>a</sup>	Yes
Total Dissolved Solids	1724	228971	$500^{b}$	2000 <sup>a</sup>	Yes
pH (S.U.)	1.21	8.36	$6.0^{b}$	$> 8.0^{a}$	Yes
Aluminum	0	5290	5 <sup>a</sup>	$20^{\rm b}$	Yes
Bicarbonate	0	4370	$0^{a}$	$10^{a}$	Yes
Boron	0	10600	$0.75^{b}$	$2^{a}$	Yes
Calcium			$0^{a}$	$20^{a}$	Yes
Chloride			$O^a$	$30^{a}$	Yes
Copper	0	33	$0.2^{a}$	5 <sup>b</sup>	Yes
Fluoride	0.8	105	$1^a$	15 <sup>b</sup>	Yes
Iron			5 <sup>a</sup>	$20^{b}$	Yes
Lithium	1.1	25340	$2.5^{a}$	2.5 <sup>b</sup>	Yes
Magnesium	0	96.5	$0^{a}$	5 <sup>a</sup>	Yes
Manganese			$0.2^{a}$	$10^{b}$	Yes
Nitrate-N	0	2670	$O^a$	$10^{a}$	Yes
Phosphate-P	0	5.3	$0^{a}$	$2^{a}$	Yes
Potassium	0.21	5490	$0^{\mathrm{a}}$	$2^{a}$	Yes
Sodium	10	204302	$O^a$	$40^{a}$	Yes
Sulfate	0	3663	$0^{\mathrm{a}}$	$20^{a}$	Yes
Zinc	0	20	$2^{\mathrm{a}}$	$10^{b}$	Yes

a Ayers and Westcot, 1994

### Oil Field Produced Water Reuse

COCs were identified by comparing concentrations in the OFPW with guideline concentrations for two water reuses (irrigation and livestock watering). These reuses were selected based on geographic location, climate, and economic need. If the concentration of any given parameter was greater than the concentration guideline for either irrigation or livestock, the parameter was considered a COC (Table 16).

### Conventional Natural Gas Produced Water Reuse

Surface water discharge (i.e. surface water augmentation) was chosen as a potential reuse option for conventional NGPW based on location (e.g. latitude, longitude and altitude) and socioeconomic needs. The constituents of conventional NGPW were compared to USEPA NPDES surface water discharge criteria (Table 17). The COCs in conventional NGPW are: TSS, TDS, Al, Ba, B, Cd, Cl, Cu, Fe, Pb, Mn, and Ni. Additional COCs, which are not specifically identified because of lack of reported analytical data, may include organic constituents such as oil and grease.

b USEPA, 2006

**Table 13.** Identification of COCs in CBMPW for reuse in irrigation. (All units are mg/L unless noted)

Parameter	Maximum Guideline	Maximum Levels	Mean Levels in	Constituent of
T arameter	Concentration	in CBMPW	CBMPW	Concern**
Conductivity (ms/m)	1700-13000	145000	4419	Yes
pH (S. U.)	4.5-9.0	6.56-9.86	-	Yes
SAR (ratio)	15	271	52.6	Yes
Bicarbonate	520	1100	668	Yes
Carbonate	3	59	24.4	Yes
Sulfate	960	5590	335	Yes
Nitrate-N	10	18.700	1.22	Yes
Ammonium-N	5	59	3.53	Yes
Phosphate-P	2	1.50	0.232	No
Al	5	1.24	0.201	No
As	0.1	0.060	0.003	No
В	0.5-15	2.40	0.151	No
Be	0.1	bdl	bdl	No
Ca	400	5870	53.6	Yes
Cd	0.01	0.010	0.005	Yes
Cl	1050	70100	845	Yes
Co	0.05	0.0007	0.0002	No
Cr	0.1	0.053	0.011	No
Cu	0.2	0.060	0.009	No
F	1	15.2	2.51	Yes
Fe	5	220	2.73	Yes
K	2	186	10.6	Yes
Li	2.5	6.88	0.146	Yes
Mg	60	1830	34.5	Yes
Mn	0.2	5.40	0.059	Yes
Mo	0.01	0.100	0.029	Yes
Na	920	34100	1180	Yes
Ni	0.2	0.203	0.020	Yes
Pb	5	0.585	0.118	No
Se	0.02	0.070	0.007	Yes
V	0.1	0.059	0.013	No
Zn	2	0.590	0.037	No

(Ayers and Westcott, 1994; WDEQ, 2001; USEPA, 2004)

<sup>\*</sup> Below detection limit (bdl)

<sup>\*\*</sup> COCs are identified as constituents in CBMPW samples that have maximum concentrations in excess of the reuse guidelines; not all constituents identified as COCs in this study are of concern in all CBMPWs

**Table 14.** Identification of COCs in CBMPW for reuse in livestock watering (All units are mg/L unless noted)

Parameter	Maximum Guideline Concentration	duideline Maximum Levels in CRMPW		Constituent of Concern	
Conductivity (ms/m)	16000	145000	4420	Yes	
pH (S.U.)	6.5-8.5	6.56-9.87	-	Yes	
Nitrate+Nitrite-N	100	18.7	1.22	No	
Nitrite-N	10	bdl	bdl	No	
Al	5	1.24	0.201	No	
As	0.2	0.060	0.003	No	
В	5	2.40	0.151	No	
Be	0.1	bdl	bdl	No	
Cd	0.05	0.010	0.005	No	
Co	1	0.001	0.000	No	
Cr	1	0.053	0.011	No	
Cu	0.5	0.060	0.009	No	
F	2	15.2	2.52	Yes	
Hg	0.01	0.0004	0.0002	No	
Mn	0.05	5.40	0.059	Yes	
Pb	0.1	0.585	0.118	Yes	
Se	0.05	0.070	0.007	Yes	
V	0.1	0.059	0.013	No	
Zn	24	0.590	0.037	No	

<sup>\*</sup>Below Detection Limit (bdl)

(Ayers and Westcot, 1994; WDEQ, 2001)

**Table 15.** Identification of COCs in CBMPW for reuse in aquaculture (All units are mg/L unless noted)

Parameter*	Maximum Guideline Concentration	Maximum Levels in CBMPW	Mean Levels in CBMPW	Constituent of Concern
pH (S.U.)	6.5-9.5	6.56-9.87	-	Yes
Al	0.1	1.24	0.201	Yes
As	0.15	0.060	0.003	No
Ba	5	190	7.04	Yes
Cd	0.002	0.010	0.005	Yes
Cu	0.01	0.060	0.009	Yes
Fe	1	220	2.74	Yes
Mn	1.5	5.40	0.059	Yes
Pb	2.5	0.585	0.118	No
Se	0.005	0.070	0.007	Yes
Zn	0.1	0.590	0.037	Yes

(WDEQ, 2001)

**Table 16.** Comparison of oilfield produced water composition (maximum concentration) with water quality guidelines for irrigation and livestock watering. Constituents of concern identified by OFPW concentration exceeding guideline concentration. – indicates guideline concentration not listed in searchable database.

		Irriga	tion	Lives	tock
	OFPW Sample	Guideline	Constituent	Guideline	Constituent
Parameter	Concentration (mg/L)	Concentration	of Concern	Concentration	of Concern
Alkalinity as CaCO	3 380	=	-	-	-
Barium	7.4	=	-	-	-
Bicarbonate	976	610	YES	-	-
Calcium	300	400	YES	=	-
Carbonate	14.6	3	-	=	-
Chloride	48	1050	NO	-	-
Iron (III)	171	-	-	-	-
Iron (soluble)	0.5	0.2	YES	-	-
Hardness as CaCO <sub>3</sub>	20	-	-	-	-
Magnesium	8.7	60	NO	-	-
Manganese	8.1	0.2	YES	0.05	YES
Nickel	9.5	0.2	YES	-	-
Nitrate	2	10	NO	-	-
Oil and grease	92	35	YES	35	YES
Dissolved oxygen	0.0002	-	-	-	-
Potassium	42.6	2	YES	-	-
Silicon	13.2	=	-	-	-
Sodium	339	920	NO	=	-
Strontium	2.2	=	-	-	-
Sulfate	3	960	NO	-	-
TDS	1370	500	YES	-	-
Tin	1.3	-	-	-	-
Zinc	17.4	2	YES	24	NO

**Table 17.** Identification of COCs in conventional NGPW for surface water discharge. All units are mg/l unless noted.

	NPDES				
Constituent	Natural Gas PW		(surface water)		COCs
	Min	Max	Min	Max	_
Total Suspended Solids	8	5484		45	Yes
Total Dissolved Solids	2600	360000	30	5000	Yes
pН	3.1	7	>6	>9	No
Temperature °C				>40	No
Aluminum	bdl	83		0.05	Yes
Barium	bdl	1740		154	Yes
Boron	bdl	56		1	Yes
Cadmium	bdl	1.21		0.0025	Yes
Chloride	1400	190000		5000	Yes
Chromium	bdl	0.03	0.1	0.2	No
Copper	bdl	5	0.37	1	Yes
Iron	bdl	1100	0.6	9.4	Yes
Lead	< 0.2	10.2	0.0056	0.015	Yes
Manganese	0.045	6.5	0.05	4.4	Yes
Nickel	bdl	9.2	0.052	2	Yes
Sulfate	<1.0	47		3000	No
Zinc	bdl	5	0.005	5	No

<sup>\*</sup> Below Detection Limit (bdl)

# Tight Gas Sand Produced Water Reuse

TGSPW, which is extracted in many states (e.g. New York, Ohio, West Virginia, Wyoming, and Colorado), has a variety of potential reuses, including irrigation and livestock. Reuse options were determined by location (e.g. latitude, longitude and altitude) and socio-economic needs. TGSPW as livestock water allows for a cost-effective alternative to municipal water. Irrigation may be a feasible reuse option where suitable water supply is limited because of drought or pollution (Frankenberger and Engberg 1998). Based on comparison of the constituents of TGSPW with USEPA FAO irrigation criteria (Table 18) the following were identified as COCs: TDS, pH, oil and grease, bicarbonate, Ca, Cl, Fe, K, Mg, Na, Pb, and sulfate. The constituents of TGSPW were also compared to criteria for livestock and wildlife (Table 19), and the following were identified as COCs: pH, oil and grease, Cu, and Mn.

External reuse options were chosen for these PWs because of their stringent requirements. Internal reuse options would be an alternative to selected external reuse options (e.g. surface water discharge) since internal reuse criteria are less stringent.

**Table 18.** FAO irrigation criteria and tight gas sand produced water characteristics for identification of COCs. All units are mg/l unless noted.

		Sand Produced Water	FAO Irrigation Criteria			
Constituents	Min	Max	Min	Max	COCs	
Total Suspended Solids		85.7	< 50 <sup>a</sup>	>100 <sup>a</sup>	No	
Total Dissolved Solids	1588	343300	$500^{b}$	$2000^{a}$	Yes	
pH (S.U.)	5	8.6	$6.0^{b}$	$> 8.0^{a}$	Yes	
Conductivity		24.4	1700	13000	No	
Oil and Grease		42		35	Yes	
Ammonium-N		2.735	$0^{a}$	5 <sup>a</sup>	No	
Arsenic		0.17	$0.1^{a}$	2	No	
Bicarbonate	854	1147	$0^{a}$	$10^{a}$	Yes	
Cadmium		0.037	$0.01^{a}$	$0.05^{b}$	No	
Calcium	3	74185	$0^{a}$	$20^{a}$	Yes	
Chloride	52	216000	$0^{a}$	$30^{a}$	Yes	
Chromium		0.265	$0.1^{a}$	$1^{b}$	No	
Copper		0.539	$0.2^{a}$	5 <sup>b</sup>	No	
Iron		0.015	5 <sup>a</sup>	$20^{b}$	Yes	
Lead		0.488	5 <sup>a</sup>	$10^{b}$	Yes	
Magnesium	2	8750	$0^{a}$	5 <sup>a</sup>	Yes	
Manganese		0.525	$0.2^{a}$	$10^{b}$	No	
Nickel		0.123	$0.2^{a}$	$2^{b}$	No	
Potassium	5	2500	$0^{a}$	$2^{a}$	Yes	
Sodium	648	6273	$0^a$	$40^{a}$	Yes	
Sulfate	12	48	$0^a$	$20^{a}$	Yes	
Zinc		0.076	$2^{a}$	$10^{b}$	Yes	

**Table 19.** Livestock and Wildlife criteria and tight gas sand produced water characteristics for identification of COCs. All units are mg/l unless noted.

	Tight Gas Pr	oduced Water		tock and ildlife	
Constituents	Min	Max	Min	Max	COCs
pН	5	8.6	6.5	8.5	Yes
Conductivity		24.4		16000	No
Oil and Grease		42		35	Yes
Arsenic		0.17		0.2	No
Cadmium		0.037		0.05	No
Chromium		0.265		1	No
Copper		0.539		0.5	Yes
Lead		0.488		5	No
Manganese		0.525		0.05	Yes
Zinc		0.076		2	No

# Design and Construct Pilot-Scale Constructed Wetland Treatment Systems (Task 4)

The objective of Task 4 is to design and construct pilot-scale constructed wetland treatment systems (CWTSs) targeting specific COCs in various produced water types to meet reuse criteria. Prior to designing specific treatment systems, produced waters were characterized (Task 2), and these characteristics were compared to specific reuse criteria (Task 3).

Wetlands offer natural environments within which unique reactions can occur (Rodgers and Castle 2008). CWTSs can be designed in a way that allows these specific reactions to take place for treatment (i.e. transfers and transformations) of targeted constituents. The benefits of CWTSs can include: low cost of operation, low maintenance, effective treatment, solar energy driven, increased effectiveness over time, tolerance of deviations in flow rate and contaminant load, and treatment of multiple COCs simultaneously and more effectively than chemical or physical treatment processes (Bhamidimarri et al. 1999; Sundaravadivel and Vigneswaran 2001; Rodgers and Castle 2008; Dorman et al. 2010). In conjunction with other treatment methods (e.g. oilwater separators, reverse osmosis and granular activated carbon (GAC) systems), hybrid CWTSs can provide effective water treatment, provided that COCs are successfully targeted through operative pathways (Murray Gulde et al. 2005a; Johnson et al. 2008b). CWTSs can be specifically designed based on targeted constituents and/or treatment pathways to transfer or transform those constituents (Hawkins et al. 1997; Gillespie et al. 1999, 2000).

CWTSs offer a potential option to conventional treatment techniques for mitigation of risks from produced water. With regard to CWTSs, there are essentially two types of produced water (PW): fresh PW and PW with elevated salinity. A CWTS designed to receive fresh PW can be designed based on COCs in a specific PW; however, in higher salinity PWs, hybrid CWTSs (i.e. CWTS with reverse osmosis) are often needed to pre-treat the PW prior to entry into a CWTS. Using COCs identified through the initial produced water characterization, CWTSs were specifically designed to transfer and transform constituents in produced water in order to decrease concentrations of targeted COCs. Pilot-scale CWTSs, while sufficiently small to enable control and manipulation of macrofeatures (e.g. hydroperiod, hydrosoil, and vegetation) and allow replication (Hawkins et al. 1997), are sufficiently large to predict performance of a full scale CWTS in terms of COC removal rates and extents (Rodgers and Castle 2008; Dorman et al. 2010).

The design of a CWTS for a specific water (e.g. produced water) includes consideration of the demographic, geologic, and meteorological characteristics of the area from which a PW originates. Depending on these conditions, a CWTS can be designed in one of two ways: subsurface flow (SSF) or free-water surface (FWS) (Figure 1). SSF CWTSs can be further subdivided into vertical flow and horizontal flow designs. Both vertical and horizontal flow SSF CWTSs are designed in a manner that maintains water levels below the top of the aggregate (e.g. rock, gravel); this allows for reduced exposure to elements at the surface (e.g. humans, wildlife, sunlight, wind, etc.) (Halverson 2004). FWS CWTSs have been used for treatment of many waters, including energy derived waters, containing many different COCs (Knight et al. 1999; Gillespie et al. 2000; Eggert et al. 2008; Johnson et al. 2008b); FWS systems are generally larger than SSF systems, but usually cost less to construct and operate/maintain (Halverson 2004). Both SSF and FWS typically exhibit lower capital costs, as well as operational/maintenance costs, than conventional water treatment systems (Halverson 2004; Mooney and Murray Gulde 2008).

Pilot scale CWTSs offer the ability to experimentally test hypotheses under a variety of conditions prior to investing in full scale CWTSs (Rodgers and Castle, 2008). Factors such as treatment rates and extents may be evaluated to determine critical full scale design parameters (e.g. loading rate, wetland area, and hydraulic retention time). In addition, the use of pilot scale CWTSs may reduce full scale performance uncertainties.

Since PWs may contain high levels of dissolved solids, low molecular weight organics, oil and grease, metals, metalloids, and ammonia, four pilot-scale CWTSs were designed and constructed. The pilot-scale CWTSs include: (1) a post-reverse osmosis (RO) PW treating system, (2) a metalloid (e.g. Se, As) and low molecular weight organic (LMWO) treating system, (3) a divalent metal (e.g. Zn, Cu, Cd, Ni) and LMWO treating system, (4) and an oil & grease and metal (e.g. Ni, Zn, Fe, Mn) treating system. Specific treatment pathways for each system were identified by reviewing peer-reviewed publications including Eh-pH diagrams (Brookins 1988), previous CWTS studies, and reduction-oxidation reactions. Each system was then specifically designed and constructed to promote the conditions required that enable the treatment pathways. Specific treatment pathways include nitrification-denitrification, reduction-oxidation, biodegradation, precipitation, and sorption.

# Pilot-Scale CWTS for Simulated Post-RO Produced Water

In waters containing elevated levels of dissolved chlorides, which may adversely impact wetland biota, an external treatment source may be required. Reverse osmosis offers the ability to remove many dissolved solids from produced waters (Veil et al, 2004). However, constituents may pass through the RO membranes. In San Ardo, California, RO is used to treat produced water containing dissolved solids and boron (Myers, pers. comm., 2008). The effluent of the treated water still contains levels of ammonia approaching 20 mg/L (Myers, pers. comm.). Other studies have found ammonia to be a major concern in post-RO treated water (Liske, 2005). For these reasons, ammonia is an important COC in post-RO treated produced water; therefore, a pilot scale CWTS was designed and constructed to treat ammonia in simulated post-RO water.

To assess the removal of ammonia by CWTSs, a FWS pilot scale system was designed and constructed to target the transformation of ammonia to nitrogenous gas by a three step process: (1) nitrification of ammonia to nitrite, (2) nitrification of nitrate to nitrate, (3) and reduction of nitrate to nitrogen gas. Because nitrification and denitrification involve different geochemical conditions (Table 20), special design considerations were made to allow microbial transformations to operate sequentially through the system.

Eight 265 L Rubbermaid<sup>®</sup> containers comprising two series of four containers, or cells (control and experimental), were filled with approximately 15 cm of fluvial sediment collected near Eighteen Mile Creek located near Clemson University. Connections between cells were built using poly-vinyl chloride (PVC) pipes with adjustable overflow levels to control water depth and allow gravity flow from each cell. Each cell was planted with approximately 20 broadleaf cattails (*Typha latifolia*) collected from a nearby aquaculture pond. The first cell from each of the two series is connected to a 5678 L polypropylene carboy holding tank containing simulated post-RO PW (20 mg/L ammonia) by a Fluid Metering, Inc. <sup>®</sup> (FMI<sup>®</sup>) piston pump. The pump delivers 90

mL/minute to one series for a hydraulic retention time of 24 hours per cell (control) and 45 mL/minute to the other series for a hydraulic retention time (HRT) of 48 hours per cell (experimental). Aeration is supplied to the first cell of the experimental series by a submerged, slotted PVC pipe connected to an air pump. Organic carbon is supplied to the second cell of the experimental series using an FMI® pump delivering 0.88 mL/minute of a 20 mg/mL solution of sucrose. A schematic of the design is shown in Figure 2.

In the pilot scale CWTS, the first process in ammonia removal is oxidation of ammonia to nitrite by autotrophic nitrifying bacteria, predominately *Nitrosomonas* (Watson et al. 1981), as shown in equation 1.

$$NH_3 + O_2 > NO_2^- + 3H^+ + 2e^-$$
 Eqn. 1 (Gambrell and Patrick, 1978)

Once oxidized to nitrite, further oxidization by autotrophic nitrifying bacteria, predominantly *Nitrobacter* (Watson et al. 1981), results in the formation of nitrate as (equation 2).

$$NO_2^- + H_2O > NO_3^{-2} + 2H^+ + 2e^-$$
 Eqn. 2 (Gambrell and Patrick, 1978)

The final process in ammonia removal is the reduction of nitrate by heterotrophic facultative anaerobes (equation 3).

$$6NO_3^{-2} + 5CH_3OH > 5CO_2 + 3N_2 + 7H_2O + 6OH^2$$
 Eqn. 3 (Knowles, 1981)

For nitrification, a high level of dissolved oxygen (>2.0 mg/L) is required in addition to a near neutral pH and sufficient levels of alkalinity (>100 mg/L) (Odell et al. 1996; USEPA 2002b). To help meet the high dissolved oxygen demand, broadleaf cattails (*T. latifolia*) were selected to increase soil aeration and consequently increase dissolved oxygen levels in the water column. In addition, a FWS design was selected to enhance diffusion of oxygen into the water by maximizing the surface area of the water/atmosphere interface. To ensure proper pH throughout the nitrification process, crushed oyster shells (50 g per cell) were added to the hydrosoil. The ground oyster shells (98% CaCO<sub>3</sub> by weight) act as both a pH buffer and a source of alkalinity for nitrification; nitrification requires 8.64 mg/L of bicarbonate per mg/L of ammonia removed (Gujer and Jenkins 1974). To further test the ability of amendments to enhance nitrification, an external source of aeration is applied to the first cell of the experimental series.

For denitrification, a low level of dissolved oxygen (<2.0 mg/L) is required in addition to a circumneutral pH. There must also be available dissolved organic carbon as an energy and electron source for the heterotrophic facultative anaerobes. To meet these criteria, the system is designed to allow detritus to serve as a cumulative source of dissolved organic material throughout each of the four cells in each series. This accumulation of dissolved organic carbon decreases the dissolved oxygen level through heterotrophic decomposition and reduces the oxidation/reduction potential (Eh) by adding reduced organic carbon to the hydrosoil. As with the nitrification steps, ground oyster shells were added to the denitrifying cells to maintain neutral pH and alkalinity. To further test the ability of amendments to enhance denitrification, an external source of carbon (sucrose) is connected to the second cell of the experimental series.

To address nutrient demands presented by the macrophytes and microbes, a time-release, nitrogen free fertilizer (Osmocote®) is added to the hydrosoil. Selection of an appropriate fertilizer is crucial as many commercially sold fertilizers may contain additional ammonia or nitrate.

Pilot-Scale CWTS for PW Containing Divalent Metals and Low Molecular Weight Organics

The pilot-scale CWTS is specifically designed to target or enhance pathways of removal (i.e. transfers or transformations) of divalent metals (Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, and Zn<sup>+2</sup>) and low molecular weight organics (LMWO) (e.g. BTEX) in simulated PW. Divalent metals may be removed under specific operating conditions (Table 21) through biogeochemical processes including: precipitation through dissimilatory sulfate reduction, organic complexation with detritus, hydrolysis, and sorption to biotic and abiotic exchange sites (Hawkins et al. 1997; Ford 1998; Murray-Gulde et al. 2005a; Rodgers and Castle 2008). Organic constituents may be removed under specific operating conditions (Table 21) through processes including: biodegradation, sorption, photolysis, and volatilization (Hawkins et al. 1997; Murray-Gulde et al. 2003; Johnson et al. 2008b; Rodgers and Castle 2008). The primary targeted pathway for removal of divalent metals (e.g. Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, and Zn<sup>+2</sup>) is precipitation through dissimilatory sulfate reduction. In a reducing environment (-100 to -350 mV), with adequate concentrations of sulfate and divalent metals, complexes may form between the divalent metals and sulfide (e.g. ZnS) through dissimilatory sulfate reduction (Allen et al. 1993). Hydrosoil redox ranging from -50 to -250 mV in reducing wetland reactors and circumneutral aqueous pH (6-9 S.U.) are favorable for dissimilatory sulfate reduction, which provides reduced sulfur for precipitating available cationic metals such as mercury, copper, lead, zinc, cadmium, and nickel (Brookins 1988; Morse 1993; Carbonell et al. 1999; Rodgers and Castle 2008). Metal-sulfide minerals are stable in aqueous systems and precipitate into the hydrosoil, significantly limiting metal bioavailability (Morse 1993). The pathway targeted for removal of LMWOs is biodegradation by microbes in an anaerobic environment (Lovley 1997). Findings that benzene can be rapidly degraded under sulfate-reducing conditions have enhanced the potential benefit of anaerobic metabolism to BTEX remediation (Edwards et al. 1992; Lovley et al. 1995; Lovley 1997).

In this pilot-scale system, a 3780 L polypropylene carboy retention basin is used for formulation and storage of simulated fresh PW. Simulated PW is homogenized using a ½ hp mixing pump. Simulated PW from the retention basin is pumped using FMI® pumps into the first of the wetland cells. The wetland cells consist of two replicated series, each containing four wetland cells. Each cell is comprised of a 378 L Rubbermaid® container that is 123 cm long by 64 cm wide by 61 cm deep. The cells are connected by PVC pipe fittings approximately 6 cm below the top of each Rubbermaid® container to allow gravity flow from each cell (Figure 3). Vegetation in the first wetland cell is giant bulrush (*Schoenoplectus californicus* C.A. Meyer). *Schoenoplectus californicus* was selected based on its ability to maintain reducing hydrosoil necessary for dissimilatory sulfate reduction (Josselyn et al. 1990; Sinicrope et al. 1992; Hawkins et al. 1997; Gillespie et al. 1999, 2000; Murray-Gulde et al. 2005b) and provide a source of organic carbon (detritus) over time for an energy source for sulfate reducing microbes (Moshiri 1993; Huddleston et al. 2000; Murray-Gulde et al. 2005b). *Schoenoplectus californicus* may also provide sorption sites for LMWOs (Brix 1994). The selected hydrosoil for the first two cells was river sediment (Eighteen Mile Creek, South Carolina) added to a depth of 30 cm to provide an

adequate root zone for *S. californicus*. Amendments to the top 10 cm of the hydrosoil included 3% hay (by volume) as an organic matter and energy source and 1.5% pelletized gypsum (CaSO4·2H<sub>2</sub>O) as a sulfur source for dissimilatory sulfate reduction (Vymazal et al. 1998). The vegetation for the final two cells in the series is narrowleaf cattail (*Typha angustifolia* L.). These final two cells contain porous quartz sand hydrosoil with no organic carbon amendments in order to facilitate aerobic conditions in order to increase the dissolved oxygen content for discharge (EPA 2005). Each cell contained approximately 140 L of surface water. By maintaining a flow rate of 97 mL/min, the nominal HRT of the pilot-scale CWTS is 24 hours per cell or 96 hours for the four wetland cells in series. The pilot-scale CWTS is constructed outdoors; an outdoor pilot-scale CWTS will provide a representation of environmental conditions that may affect full-scale CWTSs.

## Pilot-Scale CWTS for PW Containing Metalloids and Low Molecular Weight Organics

The pilot-scale experiment was designed based on results of Tasks 2 and 3, literature review regarding treatment pathways, and results from previous experiments (Eggert et al. 2008). The targeted pathways for removal of COCs in the pilot-scale CWTS may be contingent on maintenance of specific operating conditions (Table 22); therefore, regular monitoring of conditions is conducted. Removal of the metalloid elements As and Se may be achieved through precipitation; As may be coprecipitated with Fe (Edwards 1994; Hering et al. 1996), and Se may be precipitated through microbial reduction (Maiers et al. 1988; Lortie et al. 1992; Tomei et al. 1995; Garbisu et al. 1996; Ike et al. 2000; Zhang and Frankenberger 2005; Siddique et al. 2007; Zhang et al. 2008). Low molecular weight organic constituents may be removed through processes including biodegradation, sorption, photolysis, and volatilization (Hawkins et al. 1997; Johnson et al. 2008b; Rodgers and Castle 2008). The targeted removal pathway for As is coprecipitation with Fe and the targeted removal pathway for Se is microbial reduction in anaerobic conditions using microbial assemblages capable of dissimilatory selenium reduction. This process can transform aqueous As and Se into insoluble species that are unavailable for biological uptake, thus decreasing pilot-scale CWTS effluent concentrations of As and Se. The targeted pathway for removal of LMWOs is anaerobic microbial biodegradation.

A literature review provided evidence regarding an efficient pathway for Se treatment involving addition of an organic carbon source to enhance removal of soluble Se (selenate and selenite) from water through microbial reduction (Zawislanski et al. 2001b; Zhang and Frankenberger 2005) in anaerobic aquatic environments (Maiers et al. 1988; Siddique et al. 2007). This was incorporated into design of the pilot-scale CWTS by evaluating the efficiency of two different organic carbon/energy source amendments to enhance microbial abundance and activity: sucrose and a dried yeast product/nutrient additive (Aqua-Smart<sup>TM</sup>, Diamond V Corporation, Cedar Rapids, IA). Therefore, two experimental treatment series will be evaluated (e.g. sucrose and dried yeast product) and compared to one untreated control series.

The pilot-scale CWTS uses a 5678 L polypropylene carboy retention basin to house simulated PW and twelve 378 L Rubbermaid® containers (123 cm long by 64 cm wide by 61 cm deep) placed into three series of four cells using a free-water surface (FWS) design (Figure 4). The cells are connected with PVC pipe fittings approximately 6 cm below the top of each Rubbermaid® container to allow gravity flow from each cell. The untreated control series is

designated series C; the dried yeast culture treatment series is designated series Y; the sucrose treatment series is designated series S. Each cell is filled to a depth of approximately 30 cm with river sand from 18-mile Creek in Clemson, SC, and subsequently planted with *T. latifolia* harvested from an aquaculture pond on the Clemson University campus. The cells are planted at a density of approximately 25-30 plants per cell. Three piston pumps (FMI® QG400) deliver 128 mL simulated PW/minute to achieve a nominal 24-hr HRT per cell, or 96-hr per series. To maintain a circumneutral pH (6.5 – 8) and increase alkalinity in the pilot-scale CWTS, 1000 g of ground oyster shells (98% CaCO<sub>3</sub> by weight) were added to each treatment cell. To maintain reducing conditions, 100 g of zero-valent iron (Fe<sup>0</sup>) were added to each treatment cell, as well as 12 g of 19-6-12 Osmocote® fertilizer to increase the nitrogen, phosphorus, and potassium concentrations to provide essential nutrients for the microbes and plants.

## Pilot-Scale CWTS for PW Containing Oil & Grease and Divalent Metals

A CWTS must be designed in a strategic way to facilitate biogeochemical reactions that will target specified COCs. Based on comparison of characteristics of the produced waters studied in Task 2 to water quality guidelines (Task 3), the following were identified as COCs in waters such as OFPW containing oil and grease: oil and grease, Fe, Mn, Ni, and Zn. The following biogeochemical pathways were identified for treating these COCs: dissimilatory sulfate reduction (metals), oxidation (oil and grease, metals), and sorption (oil and grease, metals) (Table 23). As oil and grease in the produced water enters the system for treatment, the oil and grease are expected to sorb to available sites (e.g. hydrosoil and plant roots). The upstream cells are expected to enter a reductive state in response to the oil and grease, thus enabling dissimilatory sulfate reduction to produce sulfides that may combine with cations (e.g. Zn and Ni) to precipitate metal-sulfide minerals. The pilot-scale CWTS is designed for oil and grease content to decrease as the water moves through the system, resulting in oxidizing conditions in downstream cells. Remaining metals (e.g. Mn and Fe) will be transferred to the solid phase under oxidizing conditions.

A pilot-scale CWTS was constructed within a greenhouse owned by Clemson University. The pilot-scale system uses a 3785 L polypropylene carboy retention basin and three series of cells (Figure 5). Each series comprises four 378 L wetland cells (Rubbermaid® containers). Surface area of each wetland cell is 0.66 m<sup>2</sup>, and axial dimensions are 61 cm in height, 123 cm in length, and 64 cm in width at its widest point. Water residing in the retention basin is pumped continuously through 0.64 cm diameter plastic tubing to each of the three series using FMI® piston pumps. For each series, four cells are arranged at decreasing elevations to achieve gravity flow through the system. Water is transferred between the cells through 2.5 cm diameter PVC pipes. Two series incorporate a SSF design, and the third series incorporates a FWS design. The purpose of using two SSF series is for replication. The reasons for using a FWS series are: (1) for comparison of data with previous pilot-scale studies in which FWS cells were used and (2) for comparison of treatment effectiveness with the SSF series. In both the SSF and FWS series, the specific types of hydrosoil and plants selected promote oxidizing conditions (Armstrong et al. 1999; Kanagy et al. 2008a). Wetland cells for SSF series consist of a two layer hydrosoil of pea gravel and medium-sized gravel, and are planted with *Phragmites australis* to provide sorption sites and support oxidation. Wetland cells for FWS series consist of a hydrosoil of coarse, well sorted, quartz sand and are planted with Typha latifolia to promote oxidizing conditions in the

hydrosoil. The wetland cells were amended with Osmocote<sup>®</sup> time-released fertilizer to provide nutrients required for plant growth, which are not found in sufficient quantities in the water supply or newly emplaced hydrosoil. Hydraulic retention time in each series is controlled by manipulating inflow rate.

Table 20. Nitrification and Denitrification Operating Conditions for Simulated Post-RO Produced Water Containing

Ammonia in a Pilot-Scale Constructed Wetland Treatment System

Constituent	Pathway	Inflow Conc.	pH (S.U.)	Temp (°C)	Redox (mV)	Dissolved Oxygen (mg/L)	Alkalinity (mg CaCO3/L)	Organic Carbon
Ammonia	Nitrification	20 mg/L	$6.6 - 9.7^5$	8-30 <sup>7</sup>	$(+100) - (+350)^2$	>2.01	>100 <sup>3</sup>	None
Nitrate	Denitrification	None	7.0-8.5 <sup>1</sup>	>25 <sup>6</sup>	(-50) - (+50) <sup>2</sup>	<0.5 <sup>1</sup>	>100 <sup>3</sup>	Organic matter <sup>1,4</sup>

<sup>(1)</sup> Gambrell and Patrick 1978; (2) Gerardi 2007; (3) Gujer and Jenkins 1974; (4) Knowles, R. 1981; (5) Odell et al. 1996; (6) Pierzynski et al. 2005; (7) USEPA 2002a

**Table 21.** Operating Conditions for Simulated PW Containing Divalent Metals and LMWOs in a Pilot-Scale Constructed Wetland Treatment System

Constituents	Concentration		рН	
	(mg/L)	Pathway	(S.U.)	Redox
Cadmium	6.0	Sulfide formation; precipitation through	6-9	Reducing $-100$ to $-250 \text{ mV}^1$
		Dissimilatory Sulfate reduction		
Copper	4.0	Sulfide formation; precipitation through	6-9	Reducing -100 to -250 $\text{mV}^1$
		Dissimilatory Sulfate reduction		
Nickel	6.0	Sulfide formation; precipitation through	6-9	Reducing -100 to -250 $\text{mV}^1$
		Dissimilatory Sulfate reduction		
Zinc	6.0	Sulfide formation; precipitation through	7-9	Reducing -100 to -250 $\text{mV}^1$
		Dissimilatory Sulfate reduction		-
LMWOs	25.0	Biodegradation <sup>2</sup> ; Sorption;		Reducing
		Volatilization		_

<sup>(1)</sup> Brookins 1988; (2) Junfeng et al. 2007

**Table 22.** Operating Conditions for Simulated Produced Water Containing Metalloids and Low Molecular Weight Organics in a Pilot-Scale Constructed Wetland Treatment System

Constituent	Conc.	Pathway	pH (S.U.)	Redox	DO (mg	Alkalinity	Hardness
	(mg/L)			(mV)	$O_2/L)$	(mg	(mg
						CaCO <sub>3</sub> /L)	CaCO <sub>3</sub> /L)
Selenium	0.050	Microbial reduction <sup>5</sup>	$6.5 - 9.5^{1}$	$< +50 \text{ mV}^2$	< 3.0	> 50	> 50
Arsenic	0.020	Coprecipitation <sup>6</sup>	6.5 - 8	$< -200 \text{ mV}^2$	< 2.0	> 50	> 50
LMWOs	25.0	Biodegradation <sup>3</sup> ,	6.5 - 8	Reducing <sup>4</sup>	> 5.0	> 50	> 50
		Sorption					

<sup>(1)</sup> Lortie et al. 1992; (2) Brookins 1988; (3) Junfeng et al. 2007; (4) Lovley 1997; (5) Zhang et al. 2008; (6) Edwards 1994

Table 23. Operating conditions for simulated PW containing divalent metals and oil and grease

Constituent	Pathway	Inflow conc.	pH (S.U.)	Redox (mV)
Oil and Grease	Oxidation, Sorption	25-100 mg/L	$6.5 - 8.0^{1}$	> -501
Zn, Ni	Sulfate Reduction, Sorption	Zn: 5 mg/L Ni: 0.37 mg/L	$6.0-9.0^1$	< -150 <sup>1</sup>
Fe	Oxidation	0.4 mg/L	$6.5 - 8.0^{1}$	> -501
Mn	Oxidation, Sorption	0.5 mg/L	$6.5 - 8.0^{1}$	> -501

(1) Brookins, 1988

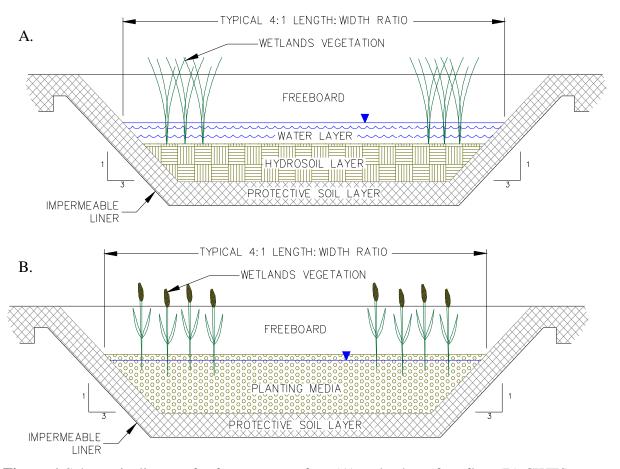


Figure 1 Schematic diagram for free-water surface (A) and sub-surface flow (B) CWTSs (Rodgers and Castle, 2008)

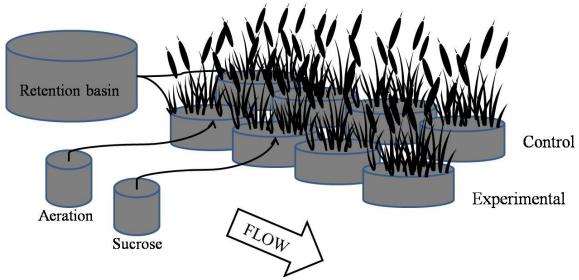
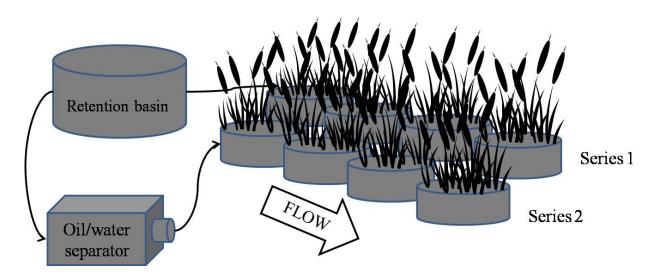
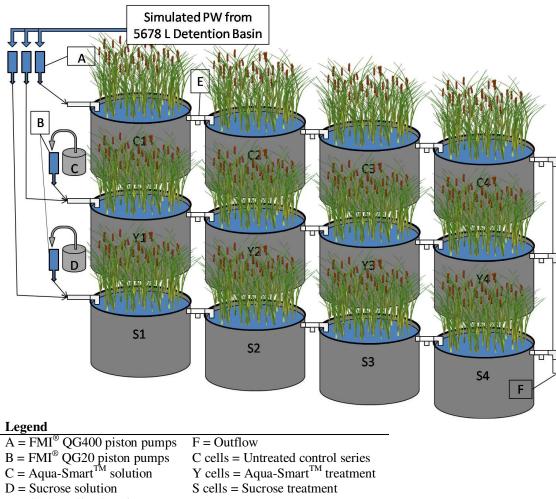


Figure 2 Schematic diagram of the pilot-scale CWTS for simulated post-RO PW

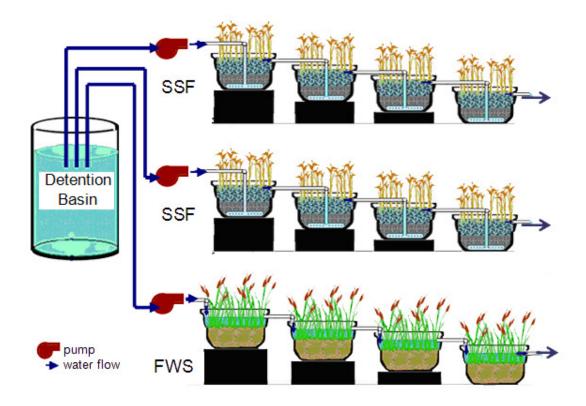


**Figure 3** Schematic diagram of the pilot-scale CWTS for simulated PW containing divalent metals and LMWOs



E = C1 Sampling location

Figure 4 Schematic diagram of the pilot-scale CWTS for simulated PW containing metalloids and LMWOs



**Figure 5** Schematic of the pilot-scale constructed wetland system constructed for simulated PW containing oil and grease. Wetland cells in subsurface flow (SSF) series are planted with *Phragmites australis*; free water surface (FWS) series is planted with *Typha latifolia*.

# Measure Treatment Performance of Pilot-Scale Constructed Wetland Systems (Task 5)

The objective of Task 5 is to measure performance of the pilot-scale systems designed and constructed during Task 4. Performance for each system is defined as the rate and extent of removal of targeted COCs. To determine the rate and extent of removal, strategic sampling and analyses for COC concentrations from sampling ports located after each cell was required. In addition, explanatory water chemistry parameters were monitored along with performance to determine whether targeted conditions for treatment pathways identified in Task 4 were met. These explanatory water chemistry parameters are in many cases directly related to the performance of the systems and are therefore referred to as explanatory parameters.

During a sampling period, water samples were collected from the influent of the systems and effluent of each cell along the treatment path. These samples were collected on a time interval equal to the designed HRT to follow the same volume of water through the length of the system. For example, with a 24 hour HRT, the first sampling day of a sampling period consisted of sampling the influent. The effluent of the first cell is sampled 24 hours later. This 24 hour sampling interval was followed until sampling of the final effluent is completed. The interval sampling method is essential for calculating accurate rate coefficients.

All samples collected from the pilot scale CWTSs were brought to the laboratory at Clemson University for immediate analysis. Performance measurements are dependent upon the COC being treated and are discussed within the individual performance sections for each system. Explanatory parameters measured for each system included pH, conductivity, alkalinity, hardness, DO, temperature, and redox potential. Table 24 lists the methods used for explanatory parameter analysis.

**Table 24.** Analytical methods for explanatory parameters in the pilot scale CWTSs

Parameter	Method	Detection Limit
pН	Direct Instrumentation: Orion Model 420A	0.01 S.U.
Conductivity	Direct Instrumentation: YSI 30	0.1 μS/cm
Alkalinity	Standard Methods: 2320 B	2 mg/L as CaCO3
Hardness	Standard Methods: 2340 B	2 mg/L as CaCO3
DO	Direct Instrumentation: YSI Model 52	0.1 mg/L
Temperature	Direct Instrumentation: YSI Model 52	0.5°C
Redox Potential	Modified standard method 2580B: GDT-11 Multi-meter	10 mV

Following completion of a sampling period, the extent of performance, or COC concentrations in each system's effluents, was compared with use criteria to determine whether treatment goals were being met.

#### Pilot-Scale CWTS for Simulated Post-RO Produced Water

Performance of the pilot scale CWTS for treating post-RO produced water was measured by determining the concentrations of both ammonia and nitrate. Although ammonia is initially the only COC present in the simulated post-RO produced water, nitrification leads to production of nitrate which may then become a COC. The treatment goals of this system were selected to make the simulated post-RO produced water suitable for irrigation. Therefore, effluent concentrations of ammonia must be below 5 mg/L and concentrations of nitrate must be below 10 mg/L. These guidelines are based on the FAO guidelines for typical irrigation waters (Ayers and Westcott, 1994). In addition, although there is no regulation for ammonia concentration for domestic use of waters in the United States, the European Union recommends a concentration of no more than 0.5 mg/L ammonia and no more than 50 mg/L nitrate (European Union, 1998). Although the system is designed to treat post-RO produced water for crop irrigation, domestic use could also be considered.

For performance measurements, samples were collected following the sampling interval outlined above (24h HRT for the control system and 48h HRT for the experimental system) from both the control (Ctrl) and experimental systems (Exp). Laboratory analysis of both ammonia and nitrate were performed using standard methods (Table 25). Results for six sampling periods are shown in Tables 26-31.

**Table 25.** Analytical methods for determining performance parameters in pilot scale CWTS for treating post-RO produced water

Parameters	Method	Method Detection Limit	Initial Concentration	Target Concentration
Ammonia	Standard Method:45000-NH3 D (Ammonia ISE)	0.1 mg/L	20 mg/L	5 mg/L
Nitrate	Colorimetric Method (HACK-modified from Standard Methods:4500-N03 C)	1 mg/L		10 mg/L

**Table 26.** Ammonia and nitrate analysis and explanatory parameters for Sampling Period 1

	Performance 1	Parameters			Expla	anatory Pa	rameters		
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	20.9	<1	11.0	355	8.14	nd*	27	28	18
Ctrl 1	18.9	<1	2.79	424	6.50	nd	26	44	36
Ctrl 2	17.6	2	3.01	430	6.05	nd	28	50	56
Ctrl 3	17.6	3	1.54	422	6.13	nd	27	50	60
Ctrl 4	17.6	4	1.32	434	6.17	nd	28	52	66
Exp 1	16.8	<1	1.76	609	6.82	nd	26	76	178
Exp 2	10.1	2	1.53	645	7.22	nd	28	180	194
Exp 3	3.8	<1	0.35	592	6.92	nd	27	166	172
Exp 4	2.3	7	0.24	575	6.92	nd	28	164	184

<sup>\*</sup> nd- not determined

**Table 27.** Ammonia and nitrate analysis and explanatory parameters for Sampling Period 2

	Performance	Parameters			E	xplanatory	Parameter	S	
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Lab Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	21.2	<1	6.01	249	6.01	nd*	25	16	14
Ctrl 1	14.4	<1	4.12	425	6.62	-194.9	26	20	16
Ctrl 2	12.9	<1	4.21	441	5.84	-133	21	20	26
Ctrl 3	13.7	<1	3.99	441	5.49	100.4	22	10	34
Ctrl 4	16.7	<1	5.08	426	5.42	111.6	21	12	32
Exp 1	7.8	1	2.42	549	6.45	-112.2	26	60	128
Exp 2	7.4	2	3.17	670	6.28	-46.2	22	110	124
Exp 3	8.0	4	2.66	720	6.43	-36.3	20	192	280
Exp 4	7.8	4	2.33	760	6.51	68.4	20	212	304

<sup>\*</sup> nd – not determined

Table 28. Ammonia and nitrate analysis and explanatory parameters for Sampling Period 3

	Performance l	Parameters			E	xplanatory	Parameter	S	
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	20.3	<1	6.91	237	7.00	nd*	14	14	16
Ctrl 1	18.2	<1	3.20	236	6.08	-156	12	8	20
Ctrl 2	17.2	<1	2.87	234	5.58	24	11	8	22
Ctrl 3	16.4	<1	2.45	233	4.80	61	11	6	24
Ctrl 4	14.3	<1	3.80	235	5.22	-27	11	10	32
Exp 1	15.9	<1	4.57	247	6.17	-263	12	8	34
Exp 2	10.3	<1	1.29	292	6.08	-260	13	50	92
Exp 3	10.3	<1	1.90	256	6.20	-190	11	56	108
Exp 4	7.4	<1	1.90	309	6.41	-17	11	96	140

<sup>\*</sup> nd – not determined

Table 29. Ammonia and nitrate analysis and explanatory parameters for Sampling Period 4

	Performance l	Parameters			E	xplanatory	Parameter	S	
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	22.3	<1.0	6.95	347	6.93	nd*	19	16	18
Ctrl 1	18.3	<1.0	4.85	332	5.01	18	16	10	28
Ctrl 2	16.5	<1.0	4.01	349	4.73	-202	16	6	26
Ctrl 3	14.2	<1.0	5.64	333	5.01	-51	16	8	28
Ctrl 4	12.6	<1.0	2.99	364	5.13	47	16	6	34
Exp 1	14.6	4	6.43	362	5.56	-291	16	10	74
Exp 2	1.4	9	2.43	379	5.62	-277	16	68	174
Exp 3	0.2	6	2.60	390	6.12	-143	15	72	140
Exp 4	< 0.1	4	1.94	422	6.09	-33	15	106	186

<sup>\*</sup> nd – not determined

Table 30. Ammonia and nitrate analysis and explanatory parameters for Sampling Period 5

	Performance 1	Parameters			E	xplanatory	Parameter	S	
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (μS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	23.2	<1.0	7.03	413	7.76	nd*	25	12	14
Ctrl 1	20.3	<1.0	1.5.0	419	5.01	-24	25	10	16
Ctrl 2	18.9	3	2.19	404	5.01	-200	24	12	18
Ctrl 3	17.6	8	2.58	400	4.94	-31	25	10	24
Ctrl 4	15.0	5	3.63	387	4.94	47	26	6	36
Exp 1	9.8	2	4.91	444	5.20	-301	26	20	22
Exp 2	1.6	7	0.69	508	5.94	-256	25	84	170
Exp 3	< 0.1	1	3.22	539	6.61	-107	26	150	208
Exp 4	< 0.1	3	4.27	567	6.18	29	27	140	204

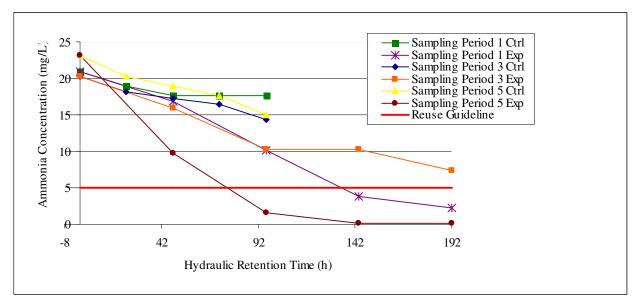
<sup>\*</sup> nd – not determined

Table 31. Ammonia	and nitrate and	alveis and	explanatory:	narameters fo	r Sampling	Period 6
Table 31. Allillollia	and muaic and	u voio anu v	CADIAHAIOIV	Darameters 10	ı Sambiniz	I CHOU O

	Performance 1	Parameters			E	xplanatory	Parameter	S	
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	25.7	<1.0	7.56	352	6.86	nd*	29	22	16
Ctrl 1	19.2	<1.0	4.07	348	5.00	28	28	10	24
Ctrl 2	17.2	<1.0	2.65	356	5.46	-165	28	18	42
Ctrl 3	15.6	2	2.31	367	5.33	42	28	14	50
Ctrl 4	13.2	<1.0	3.68	387	5.21	67	28	16	56
Exp 1	4.4	<1.0	5.71	373	5.31	-254	28	10	100
Exp 2	2.6	6	2.22	476	6.44	-310	28	98	170
Exp 3	< 0.1	4	3.21	626	6.20	-159	28	104	210
Exp 4	< 0.1	1	4.09	789	6.34	41	29	106	226

<sup>\*</sup> nd – not determined

For each of the sampling periods, the control system did not meet the targeted performance goal for ammonia. During sampling periods 2 and 3, the experimental system did not meet the targeted performance goal for ammonia; however, during sampling periods 1, 4, 5, and 6 the targeted performance goal was met (Figure 6). Sampling periods 2, 4, and 6 were excluded from the figure due to similar ammonia concentrations



**Figure 6.** Ammonia removal during 3 sampling periods.

Sampling periods 2 and 3 were performed during cold winter months when freezing conditions negatively affected nitrification. This was demonstrated by simultaneously sampling the outdoor systems (Ctrl and Exp) during sampling period 3 and a pair of indoor control systems (Ictrl 1 and 2). These indoor systems were constructed with the same design parameters as the outdoor control system (24h HRT, cattails, identical water simulation, and fluvial hydrosoil) to establish similar environmental conditions excluding water and air temperatures. The steady water

temperatures were obtained by placing the indoor control systems inside a climate controlled greenhouse. Results (Table 32) indicate that the outdoor control system was not able to obtain a level of performance comparable to that of the indoor control systems. Therefore, water and air temperatures are important considerations for design of a CWTS treating post-RO produced water.

Table 32. Ammonia and nitrate analysis and explanatory parameters for control systems located in a climate controlled

greenhouse during Sampling Period 2

	Performance I	Parameters							
Sample	[Ammonia-N] (mg/L)	[Nitrate-N] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	21.9	<1	5.95	241	7.73	Nd*	15	18	22
Ictrl 1	10.7	<1	3.68	332	5.79	-187	15	34	98
Ictrl 2	12.0	<1	3.08	364	5.47	-66	16	26	82

nd - not determined

Nitrate concentrations were below the use guideline concentrations for all samples during all sampling periods.

Pilot-Scale CWTS for PW Containing Divalent Metals and Low Molecular Weight Organics

An outdoor pilot-scale CWTS was designed and constructed to remove Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs. Composition of the simulated water is similar to that of fresh shale gas produced water (McIntosh et al. 2002). Performance of this system is defined as the rate and extent of removal of COCs. Performance was measured by an Inductively Coupled Plasma—Atomic Emission Spectrometer (ICP-AES) for Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, and Zn<sup>+2</sup> concentrations (USEPA 1994a) and total LMWOs were measured using a spectrofluorometric technique modified from Zhou et al. (2009).

Targeted inflow concentrations of constituents in the simulated PW were 4 mg/L Cu<sup>+2</sup>, 6 mg/L Cd<sup>+2</sup>, 6 mg/L Ni<sup>+2</sup>, 6 mg/L Zn<sup>+2</sup>, and 25 mg/L LMWOs. For sampling period 1, the initial performance (3.6 mg/L Cd<sup>+2</sup>, 1.98 mg/L Cu<sup>+2</sup>, 2.74 mg/L Ni<sup>+2</sup>, and 3.9 mg/L of Zn<sup>+2</sup>) of the pilot-scale CWTS (Table 33 and Figures 7 and 8) was improved with the addition of calcium carbonate to the simulated PW during sampling period 2 (Table 34 and Figures 9 and 10) to provide buffering capacity (i.e. alkalinity) to the simulated water. Table 35 illustrates the differences in percent removal before and after the addition of calcium carbonate to the simulated PW. The improvement in performance with the addition of calcium carbonate may be due to maturity of the system, calcium carbonate providing buffering capacity for the simulated fresh PW to maintain a pH above 6, and temperature increases (~15°C to 24°C; Tables 34, 36-38 and Figures 11-16), which may promote microbial activity. Microbial processes, such as dissimilatory sulfate reduction (for Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>) and biodegradation by heterotrophic aerobic microbes (for LMWOs) are the targeted removal pathways, and both may be temperature dependent (Abdollahi and Nedwell 1979; Halda-Alija and Johnston 1999). From sampling periods 2 to 5 for series 1, copper concentrations increased from 0.04 mg/L to 0.08 mg/L; nickel concentrations decreased from 1.87 mg/L to 0.41 mg/L; zinc concentrations decreased from 1.52 mg/L to 0.08 mg/L; and cadmium concentrations decreased from 0.82 mg/L to 0.2 mg/L. Copper

was the only element to increase from sampling period 2 to 5 for series 1. From sampling periods 2 to 5 for series 2, copper concentrations decreased from 0.05 mg/L to 0.02 mg/L; nickel concentrations from 1.60 mg/L to 0.04 mg/L; zinc concentrations from 1.51 mg/L to 0.02 mg/L; and cadmium concentrations from 0.79 mg/L to 0.09 mg/L.

LMWOs concentrations were non-detect (< 1mg/L) after the first wetland cell. The goal of 1 mg LMWOs/L in the outflow water has been achieved consistently by series 1 and 2. The treatment goals to achieve specific water use criteria (i.e. irrigation and livestock watering) were accomplished for each of the four divalent metals and for LMWOs.

**Table 33.** Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs concentrations and explanatory parameters for Sampling Period 1

		Perf	ormance P				Expla	anatory F	Paramete	rs		
	Copper	Nickel	Zinc	Cadmium	[LMWO]	DO	Cond.	рН	Redox	Temp	Alkalinity (mg/L as	Hardness (mg/L as
Sample	(mg/L)	(mg/L)	(mg/L)	(mg/L)	(mg/L)	(mg/L)	(μS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	CaCO <sub>3</sub> )
Inflow	4	6.2	6.1	5.9	bdl	10.56	1732	6.75	nd	11.2	14	152
T1-1	3.4	5.7	5.4	6.15	0.35	9.06	1660	6.6	-233	10.6	10	408
T1-2	3.1	4.9	4.8	5.61	bdl	7.69	1655	6.76	-175	8.3	12	420
T1-3	2.4	2.9	4.1	3.9	bdl	11.76	1638	6.84	-140	9.8	14	580
T1-4	1.98	2.74	3.9	3.6	bdl	9.01	1723	6.7	139	10.3	16	472
T2-1	4.12	6.2	5.85	4.8	0.65	7.51	1732	6.5	-162	11.9	12	308
T2-2	2.55	6.1	5.7	4.2	bdl	6.16	1790	6.42	-206	12.4	14	268
T2-3	2.37	5.9	5.1	3.9	bdl	9.64	1576	6.62	-213	8.9	14	368
T2-4	2.43	5.8	4.5	3.75	bdl	10.2	1675	6.59	-17.9	10.1	12	284

nd- not determined

bdl- below detection limit

**Table 34.** Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs concentrations and explanatory parameters for Sampling Period 2

		Perf	ormance Pa				Expl	lanatory I	Parameter	·s		
Sample	Copper (mg/L)	Nickel (mg/L)	Zinc (mg/L)	Cadmium (mg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	3.53	6.56	5.79	5.35	bdl	7.30	1838	6.91	nd	18.9	16	352
T1-1	0.21	1.99	1.49	1.46	bdl	7.81	1620	6.72	-233	16.3	29	308
T1-2	0.12	2.32	1.86	1.36	bdl	4.64	1560	6.60	-215	16.3	30	360
T1-3	0.15	2.45	2.06	1.51	bdl	9.39	1412	6.79	-100	15.3	40	312
T1-4	0.04	1.87	1.52	0.82	bdl	8.23	1078	6.52	122	16.3	36	290
T2-1	0.26	1.69	1.42	0.99	bdl	8.12	1642	6.61	-241	14.0	32	254
T2-2	0.12	2.49	2.40	1.58	bdl	7.59	1556	6.42	-222	17.5	34	236
T2-3	0.13	2.43	2.39	1.43	bdl	6.79	1101	6.49	-57	16.2	40	126
T2-4	0.05	1.60	1.51	0.79	bdl	6.79	799	6.48	118	15.9	40	276

nd-not determined

bdl- below detection limit

**Table 35**. Percent removal of Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, and Zn<sup>+2</sup> before and after addition of calcium carbonate

Before CaCO <sub>3</sub>	Copper	Cadmium	Nickel	Zinc	After CaCO <sub>3</sub>	Copper	Cadmium	Nickel	Zinc
Series 1	50.5%	55.8%	36.1%	38.9%	Series 1	98.7%	85.4%	75.4%	73.7%
Series 2	39.2%	6.5%	26.2%	36.4%	Series 2	98.7%	84.7%	71.2%	73.7%

 $\textbf{Table 36.} \ \text{Cd}^{+2}, \ \text{Cu}^{+2}, \ \text{Ni}^{+2}, \ \text{Zn}^{+2}, \ \text{and LMWOs concentrations and explanatory parameters for Sampling Period 3}$ 

		Per	formance P	arameters				Exp	lanatory F	Parameter	s	
Sample	Copper (mg/L)	Nickel (mg/L)	Zinc (mg/L)	Cadmium (mg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.49	4.26	4.57	4.37	5.41	3.55	410	6.81	nd	24.2	32	542
T1-1	0.87	2.24	1.43	1.62	1.22	2.91	430	6.44	-197.7	23.5	20	286
T1-2	0.73	0.68	0.58	0.34	bdl	2.25	594	6.69	-118	23.9	42	236
T1-3	0.63	0.58	0.46	0.28	bdl	3.15	526	6.71	-202	23.7	38	178
T1-4	0.54	0.61	1.22	0.25	bdl	2.53	596	6.67	-72	22.9	42	144
T2-1	1.11	1.38	1.21	1.43	1.89	4.87	428	6.59	-175.2	23.4	46	254
T2-2	1.41	0.89	0.38	0.25	bdl	2.51	448	6.63	-164.2	23.8	52	174
T2-3	0.89	0.38	0.29	0.05	bdl	1.27	434	6.63	-210	23.6	50	152
T2-4	0.06	0.09	0.09	0.01	bdl	1.59	473	6.71	-124.9	23.7	36	122

nd-not determined

bdl– below detection limit

**Table 37.** Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs concentrations and explanatory parameters for Sampling Period 4

		Per	formance l	Parameters				Expl	lanatory F	arameter	s	
Sample	Copper (mg/L)	Nickel (mg/L)	Zinc (mg/L)	Cadmium (mg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	3.51	4.82	3.51	4.45	5.23	3.21	1691	7.04	nd	26.7	30	576
T1-1	1.16	3.63	2.48	2.28	0.17	2.09	1620	6.5	-245.6	24.6	20	296
T1-2	1.94	2.17	1.19	0.12	bdl	1.91	1388	6.52	-251.7	23.9	44	224
T1-3	1.1	1.28	0.52	bdl	bdl	1.91	1386	6.6	-168	23.5	42	156
T1-4	1.49	1.31	0.44	2.09	bdl	1.92	1152	6.75	-25	23.5	44	184
T2-1	0.88	0.63	0.39	0.75	1.14	2.47	1264	6.54	-258	23.2	44	216
T2-2	0.92	0.59	0.41	bdl	bdl	2.05	1100	6.5	-237	22.8	58	160
T2-3	0.053	0.12	0.12	0.07	bdl	1.95	862	6.45	-144	23.4	52	136
T2-4	0.03	0.01	0.08	bdl	bdl	2.55	741	6.39	-22	23.3	34	100

nd-not determined

bdl– below detection limit

**Table 38.** Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs concentrations and explanatory parameters for Sampling Period 5

		Per	formance F	Parameters				Exp	lanatory P	arameters	S	
Sample	Copper (mg/L)	Nickel (mg/L)	Zinc (mg/L)	Cadmium (mg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	2.839	2.88	3.81	1.98	6.59	2.93	2014	7.02	nd	34.2	40	208
T1-1	1.958	1.51	1.96	0.65	1.611	2.64	1537	6.29	-222.7	25.1	32	164
T1-2	0.296	1.58	0.30	0.34	bdl	1.41	1047	6.47	-203	25.3	64	196
T1-3	0.1513	0.52	0.15	0.31	bdl	2.17	965	6.45	-342	25.1	38	136
T1-4	0.084	0.41	0.08	0.02	bdl	1.56	946	6.43	-289	24.7	26	100
T2-1	0.85	1.07	0.85	0.64	0.103	1.36	1295	6.65	-169	24.5	82	152
T2-2	0.45	0.63	0.45	0.50	bdl	1.3	1179	6.46	-177	24.5	80	212
T2-3	0.26	0.11	0.26	0.18	bdl	1.54	727	6.48	-12	24.4	80	140
T2-4	0.02	0.04	0.02	0.09	bdl	2.15	756	6.51	13	24.6	44	104

nd - not determined

bdl– below detection limit

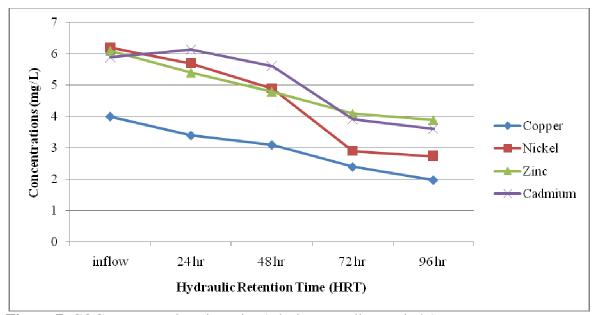


Figure 7. COC concentrations in series 1 during sampling period 1.

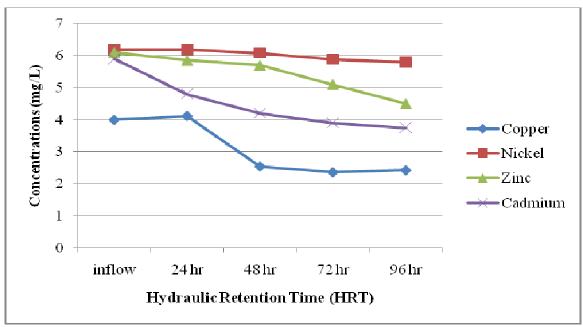
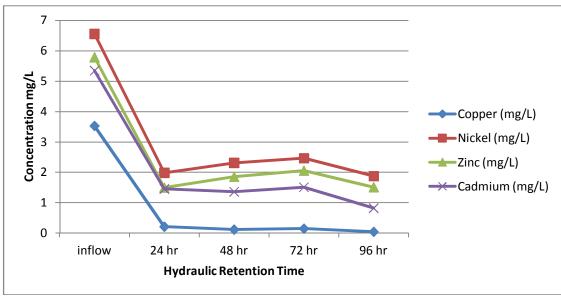
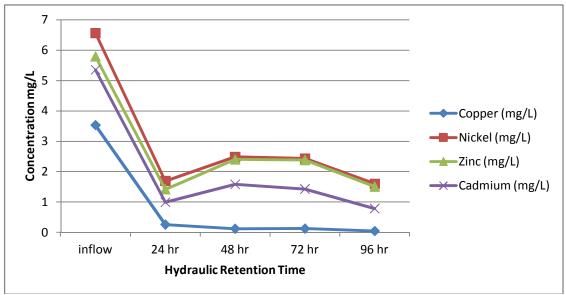


Figure 8. COC concentrations in series 2 during sampling period 1.

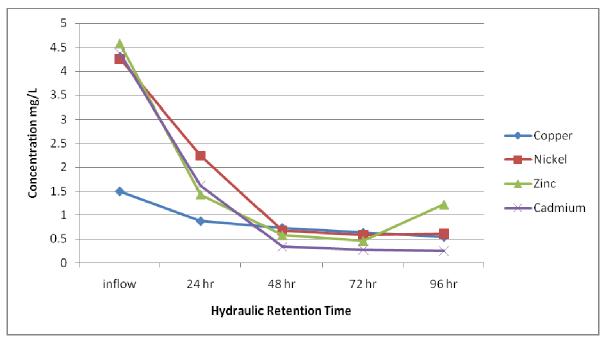


**Figure 9.** COC concentrations in series 1 after the addition of calcium carbonate salt to simulated fresh PW during sampling period 2.

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**Figure 10.** COC concentrations in series 2 after the addition of calcium carbonate salt to simulated fresh PW during sampling period 2.



**Figure 11.** COC concentrations in series 1 during sampling period 3.

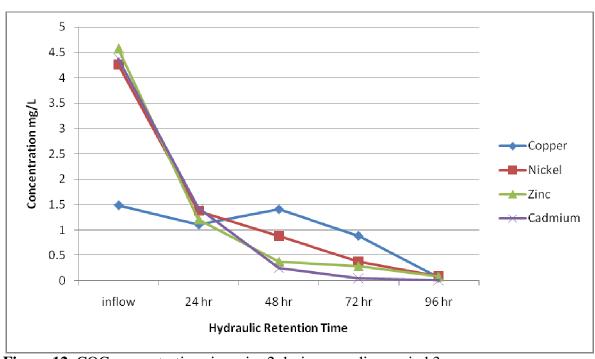


Figure 12. COC concentrations in series 2 during sampling period 3.

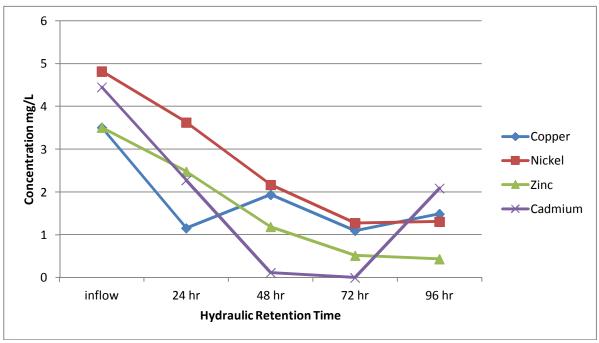


Figure 13. COC concentrations in series 1 during sampling period 4.

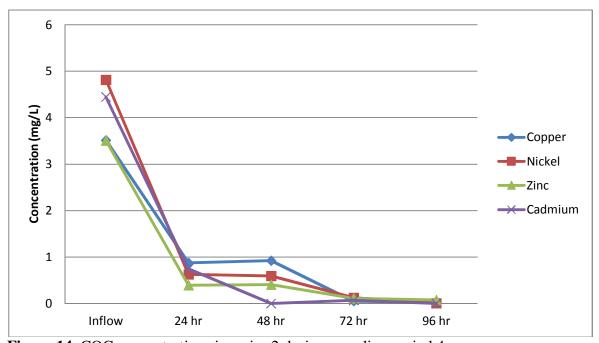


Figure 14. COC concentrations in series 2 during sampling period 4.

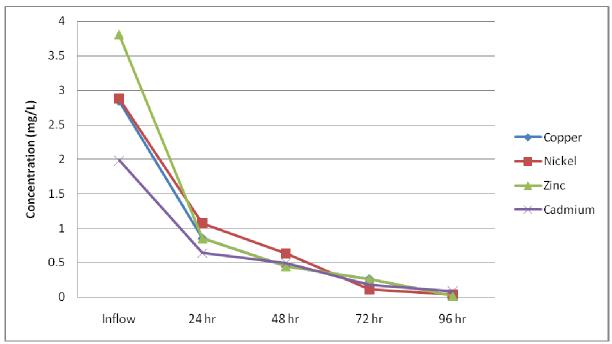


Figure 15. COC concentrations in series 1 during sampling period 5.

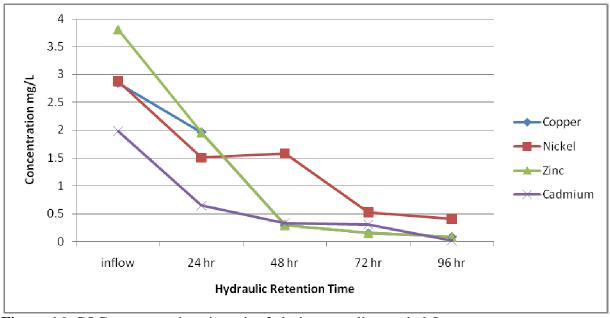


Figure 16. COC concentrations in series 2 during sampling period 5.

## Pilot-Scale CWTS for PW Containing Metalloids and Low Molecular Weight Organics

Simulated PW containing metalloids (e.g. Se, As) and LMWOs was used for the pilot-scale experiment; simulating this water increases repeatability, allows easier manipulation of constituents and their respective concentrations, and is more cost effective than shipment of large volumes of PW for experimentation. After initial construction of the pilot-scale CWTS, a period of maturation was allowed, permitting the plants time to become established and initiate new growth. To accomplish acclimation, simulated PW was loaded into the pilot-scale CWTS at increasing COC concentrations for one month prior to addition of organic carbon sources; sucrose and nutrient (i.e. AquaSmart<sup>TM</sup>) amendments were subsequently loaded into the pilot-scale CWTS inflow for two weeks.

The primary objectives for this part of the project were to remove Se, As, and LMWOs from simulated PW. To measure performance (i.e. removal of COCs), simulated PW samples were collected from the pilot-scale CWTS inflow and outflow from each cell. Samples were analyzed for total Se, total As, and LMWOs, as well as general water chemistry parameters (i.e. explanatory parameters). Total Se was measured using atomic absorption spectrometry (hydride generation: Standard Method 3114 C; APHA 2005); total As was measured using graphite furnace atomic absorption spectrometry (modified EPA method 200.9). Total LMWOs were measured using a spectrofluorometric method modified from Zhou et al. (2009). Initial COC concentrations of Se, As and LMWOs in the simulated PW were  $50 \,\mu\text{g/L}$ ,  $20 \,\mu\text{g/L}$ , and  $25 \,\text{mg/L}$ , respectively; targeted outflow COC concentrations were  $5 \,\mu\text{g}$  Se/L,  $5 \,\mu\text{g}$  As/L, and 1 mg LMWOs/L.

After the maturation and acclimation period, the goal of  $5 \mu g$  Se/L in the outflow water was achieved and maintained in the pilot-scale CWTS with the AquaSmart<sup>TM</sup> amendment (Figures 17-22). The sucrose amendment also achieved the goal of  $5 \mu g$  Se/L, but with less consistency than the AquaSmart<sup>TM</sup> amendment. The untreated control series achieved no significant Se treatment. Arsenic treatment was accomplished, but less successfully than Se; As treatment is under further investigation. The goal of  $5 \mu g$  As/L in the outflow water was achieved by both the untreated control series and the AquaSmart<sup>TM</sup> series; however, removal was not achieved consistently (Figures 23-28). The goal of 1 mg LMWOs/L in the outflow water was achieved consistently by all treatments, as well as the untreated control (Tables 39-44). This goal was achieved in the detention basin, prior to entering the pilot-scale CWTS. This implies that LMWOs brought to the surface in water produced from a well may readily volatilize when exposed to lower pressure than present in underground formations.

Table 39. Se, As, LMWO analysis and explanatory parameters for Sampling Period 1

	Perfor	rmance Pa	arameters			Expl	anatory P	arameter	S	
Sample	[Se] (µg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	44.6	21.8	bdl	8.2	2812	7.35	nm	25.8	40	138
Control 1	43.4	13.3	bdl	8.1	2838	7.00	-203.9	25.1	24	134
Control 2	42.8	12.1	bdl	8.2	2856	6.90	-181.0	24.3	24	134
Control 3	44.3	7.9	bdl	8.3	2868	6.80	+10.4	24.1	22	132
Control 4	45.7	5.6	bdl	8.1	2880	6.73	-115.4	24.0	24	136
AquaSmart <sup>TM</sup> 1	3.0	15.2	bdl	1.4	2845	6.56	-126.8	25.2	38	140
$AquaSmart^{TM}$ 2	1.3	16.5	bdl	2.5	2871	6.67	-69.9	24.0	34	144
AquaSmart <sup>TM</sup> 3	bdl	12.0	bdl	6.3	2890	6.95	-45	23.2	74	148
AquaSmart <sup>TM</sup> 4	bdl	3.3	bdl	7.6	2930	7.05	+54.2	23.0	70	148
Sucrose 1	18.3	19.2	bdl	5.1	2819	6.41	-234.7	24.9	38	138
Sucrose 2	15.9	16.3	bdl	2.6	2802	6.38	-97.6	24.0	46	136
Sucrose 3	9.9	15.0	bdl	4.8	2804	6.60	-94.4	23.3	50	136
Sucrose 4	6.1	13.4	bdl	5.2	2812	6.63	-44.3	23.2	64	146

nm - not measured

bdl- below detection limit

**Table 40.** Se, As, LMWO analysis and explanatory parameters for Sampling Period 2

	Perfo	rmance Pa	arameters			Expl	anatory P	arameter	S	
Sample	[Se] (µg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	49.7	14.0	bdl	8.4	2612	7.26	nm	13.2	42	140
Control 1	45.1	10.6	bdl	8.2	2630	7.06	-208.6	15.1	26	138
Control 2	46.3	7.5	bdl	8.1	2643	6.86	-188.2	15.9	26	136
Control 3	43.7	6.4	bdl	8.2	2666	6.88	+9.22	16.5	24	130
Control 4	44.6	5.0	bdl	8.2	2701	6.80	-125.2	16.9	28	132
AquaSmart $^{TM}$ 1	8.8	9.5	bdl	1.2	2642	6.49	-135.2	14.9	40	142
$AquaSmart^{TM}\ 2$	2.8	6.9	bdl	2.3	2665	6.64	-84.6	15.7	38	142
AquaSmart <sup>TM</sup> 3	1.1	7.8	bdl	5.7	2684	6.94	-51.3	16.7	76	152
AquaSmart $^{TM}$ 4	bdl	6.3	bdl	7.4	2713	7.10	+46.5	16.5	74	154
Sucrose 1	38.2	8.2	bdl	4.3	2680	6.34	-241.2	15.2	42	140
Sucrose 2	24.2	5.2	bdl	2.2	2695	6.54	-101.3	15.8	48	140
Sucrose 3	13.8	5.1	bdl	4.5	2721	6.72	-91.2	16.3	50	138
Sucrose 4	8.1	5.5	bdl	5.4	2732	6.74	-49.2	16.9	66	152

nm – not measured

bdl – below detection limit

Table 41. Se, As, LMWO analysis and explanatory parameters for Sampling Period 3

	Perfo	rmance Pa	Parameters Explanatory Parameters  Alkalinity Har								
Sample	[Se] (µg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )	
Inflow	46.5	18.4	bdl	8.5	1727	7.15	nm	11.5	40	136	
Control 1	44.9	16.5	bdl	8.3	1865	6.89	-194.9	13.0	28	132	
Control 2	41.9	13.1	bdl	8.0	2086	6.71	-133.0	14.8	24	132	
Control 3	39.1	11.9	bdl	8.2	2145	6.59	+100.4	15.0	26	128	
Control 4	37.9	10.9	bdl	8.1	2128	6.62	+111.6	14.6	26	128	
AquaSmart <sup>TM</sup> 1	7.9	19.0	bdl	1.1	1849	6.43	-112.2	13.5	42	140	
$AquaSmart^{TM}$ 2	2.2	15.3	bdl	2.1	1908	6.54	-46.2	14.7	36	140	
AquaSmart <sup>TM</sup> 3	bdl	15.6	bdl	5.2	2055	6.87	-36.3	13.7	74	148	
AquaSmart <sup>TM</sup> 4	bdl	14.7	bdl	7.7	2020	6.90	+68.4	13.5	76	152	
Sucrose 1	36.2	16.7	bdl	3.9	1805	6.21	-221.7	13.0	40	138	
Sucrose 2	26.5	17.1	bdl	2.0	1867	6.43	-63.5	13.7	46	136	
Sucrose 3	13.8	17.2	bdl	4.3	2055	6.75	-46.3	14.2	48	140	
Sucrose 4	7.3	18.3	bdl	5.1	2020	6.73	-16.3	13.6	58	148	

nm – not measured

bdl – below detection limit

Table 42. Se, As, LMWO analysis and explanatory parameters for Sampling Period 4

	Perfo	rmance Pa	arameters	•							
Sample	[Se] (µg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )	
Inflow	42.7	18.4	bdl	8.4	2103	7.26	nm	14.6	44	144	
Control 1	38.3	13.5	bdl	8.1	2154	6.94	-168.5	15.4	34	136	
Control 2	45.7	10.4	bdl	8.2	2209	6.76	-115.2	16.8	28	134	
Control 3	39.7	8	bdl	8.0	2231	6.62	+64.2	17.2	30	130	
Control 4	39.7	6	bdl	8.1	2281	6.54	+80.5	17.8	28	126	
$AquaSmart^{TM}$ 1	27.3	15.1	bdl	0.9	2145	6.52	-145.6	15.2	44	146	
$AquaSmart^{TM}\ 2$	4.5	8.1	bdl	1.6	2201	6.59	-64.2	16.9	38	144	
AquaSmart <sup>TM</sup> 3	bdl	8.6	bdl	4.6	2263	6.90	-29.5	17.6	72	146	
AquaSmart $^{TM}$ 4	bdl	9	bdl	7.4	2298	6.87	+58.6	17.9	78	150	
Sucrose 1	29.8	18.4	bdl	1.7	2142	6.42	-240.3	15.6	42	140	
Sucrose 2	3.0	11.6	bdl	1.6	2211	6.51	-52.3	16.4	44	142	
Sucrose 3	6.5	8.2	bdl	4.1	2245	6.65	-50.1	17.5	46	144	
Sucrose 4	bdl	6.3	bdl	4.4	2284	6.64	-25.9	18.0	60	150	

nm – not measured

bdl – below detection limit

 Table 43. Se, As, LMWO analysis and explanatory parameters for Sampling Period 5

	Perfo	rmance Pa	arameters	Explanatory Parameters						
Sample	[Se] (µg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	76.0	18.3	bdl	8.2	2210	7.52	nm	15.9	44	146
Control 1	52.7	14.1	bdl	8.0	2264	6.89	-171.2	16.2	32	132
Control 2	55.2	8.7	bdl	8.0	2300	6.74	-119.4	16.7	32	136
Control 3	56.7	7.1	bdl	7.8	2323	6.64	+52.3	17.0	32	134
Control 4	58.2	4.3	bdl	7.9	2384	6.41	+65.3	17.5	30	128
AquaSmart $^{TM}$ 1	11.2	17.9	bdl	1.1	2225	6.58	-154.8	16.3	46	152
$AquaSmart^{TM}\ 2$	bdl	9.8	bdl	1.7	2284	6.69	-71.6	16.9	36	148
AquaSmart <sup>TM</sup> 3	bdl	8.7	bdl	5.2	2317	6.87	-35.2	17.4	68	154
AquaSmart $^{TM}$ 4	bdl	9.7	bdl	7.5	2345	6.96	+54.3	17.6	76	156
Sucrose 1	6.3	7.1	bdl	1.9	2231	6.25	-234.5	16.5	40	142
Sucrose 2	2.5	5.4	bdl	2.0	2256	6.49	-64.2	16.7	46	144
Sucrose 3	2.2	5.4	bdl	4.0	2289	6.54	-42.1	17.0	42	138
Sucrose 4	5.8	3.5	bdl	4.3	2321	6.59	-34.8	17.5	54	146

nm – not measured

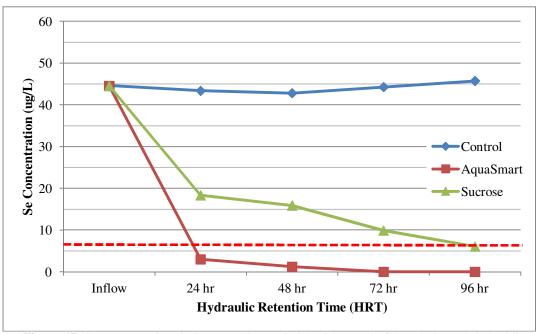
bdl – below detection limit

**Table 44.** Se, As, LMWO analysis and explanatory parameters for Sampling Period 6

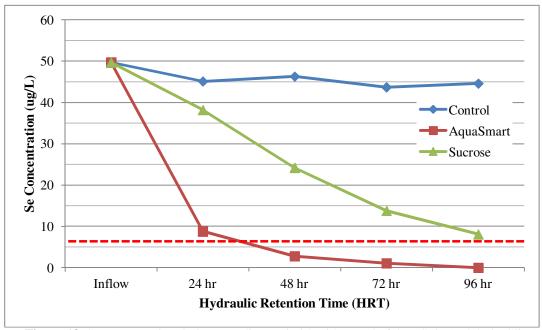
	Performance Parameters			Explanatory Parameters						
Sample	[Se] (μg/L)	[As] (µg/L)	[LMWO] (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L CaCO <sub>3</sub> )	Hardness (mg/L CaCO <sub>3</sub> )
Inflow	50.8	17.2	bdl	8.6	2525	7.4	nm	16.3	42	144
Control 1	51.6	22.6	bdl	8.4	2528	6.91	-170.3	16.8	34	134
Control 2	61.4	4.4	bdl	8.9	2500	6.93	-120.4	17.1	32	134
Control 3	63.6	4.0	bdl	8.4	2523	6.96	+51.8	17.4	34	136
Control 4	56.4	2.4	bdl	7.8	2489	6.92	+65.4	17.8	32	130
$AquaSmart^{TM}$ 1	6.3	12.7	bdl	3.5	2345	6.60	-157.5	16.7	48	152
$AquaSmart^{TM}$ 2	5.0	15.1	bdl	4.8	2360	6.82	-72.6	17.0	38	150
AquaSmart <sup>TM</sup> 3	1.9	10.1	bdl	6.1	2230	6.82	-38.7	17.5	70	156
AquaSmart $^{TM}$ 4	bdl	6.9	bdl	6.8	2290	6.79	+52.6	17.8	78	158
Sucrose 1	22.2	13.5	bdl	5.6	2330	6.32	-245.1	16.3	42	144
Sucrose 2	11.7	10.5	bdl	2.4	2280	6.51	-68.2	16.7	48	146
Sucrose 3	9.2	9.5	bdl	5.0	2260	6.68	-45.6	17.2	44	140
Sucrose 4	2.9	9.8	bdl	4.9	2265	6.71	-35.6	17.7	56	148

nm – not measured

bdl – below detection limit



**Figure 17.** Se concentration during sampling period 1 with a goal of 5  $\mu$ g Se/L (red dashed line)



**Figure 18.** Se concentration during sampling period 2 with a goal of 5  $\mu$ g Se/L (red dashed line)

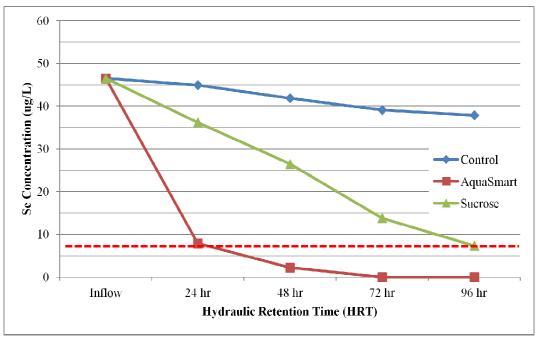
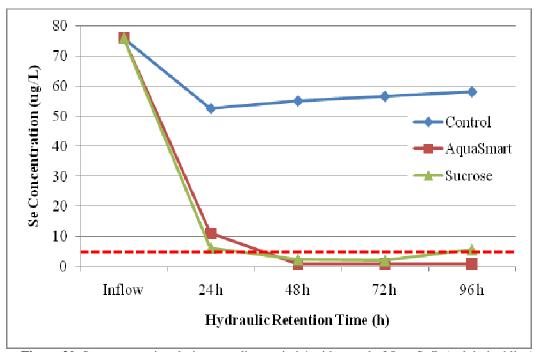


Figure 19. Se concentration during sampling period 3 with a goal of 5 μg Se/L (red dashed line)



**Figure 20.** Se concentration during sampling period 4 with a goal of 5  $\mu$ g Se/L (red dashed line)

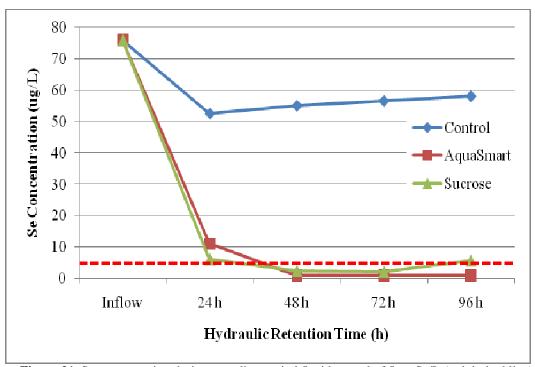


Figure 21. Se concentration during sampling period 5 with a goal of 5 μg Se/L (red dashed line)

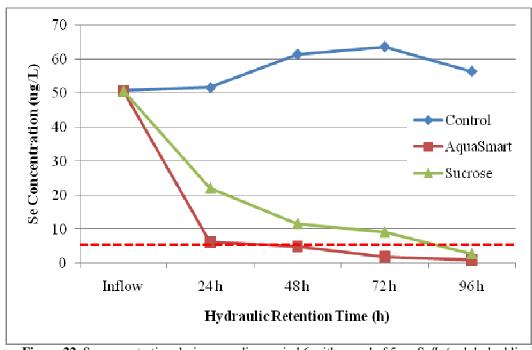


Figure 22. Se concentration during sampling period 6 with a goal of 5  $\mu$ g Se/L (red dashed line)

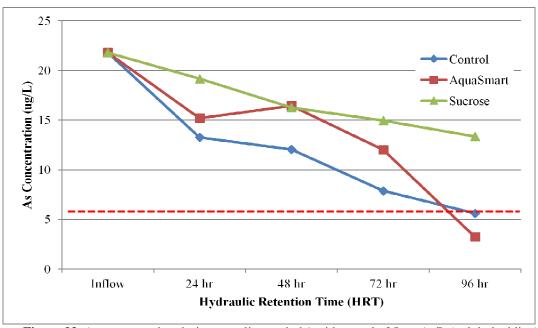
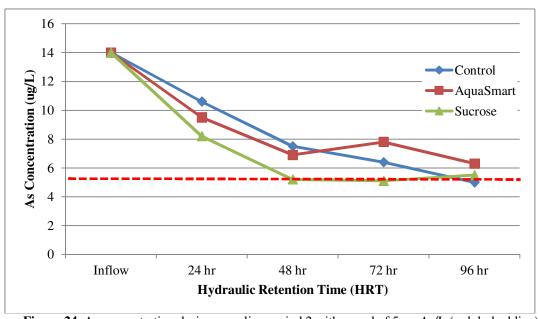
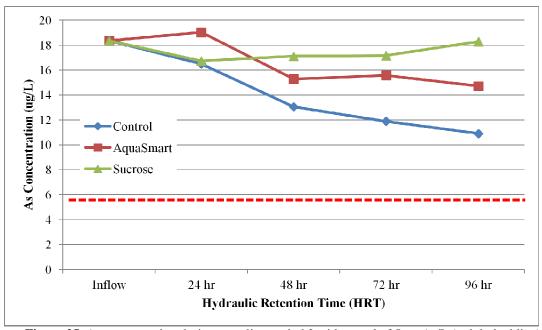


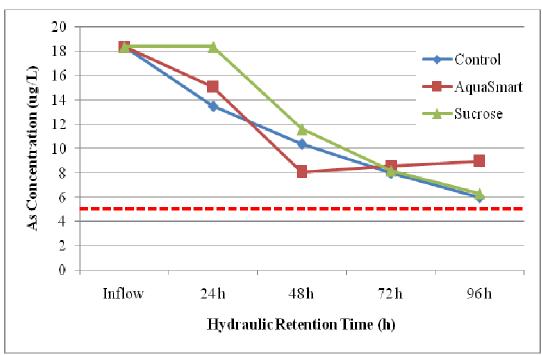
Figure 23. As concentration during sampling period 1 with a goal of 5  $\mu$ g As/L (red dashed line)



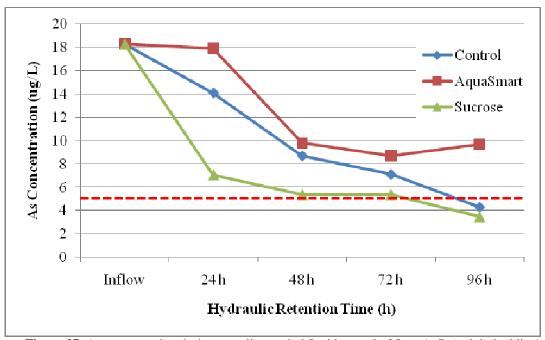
**Figure 24.** As concentration during sampling period 2 with a goal of 5  $\mu$ g As/L (red dashed line)



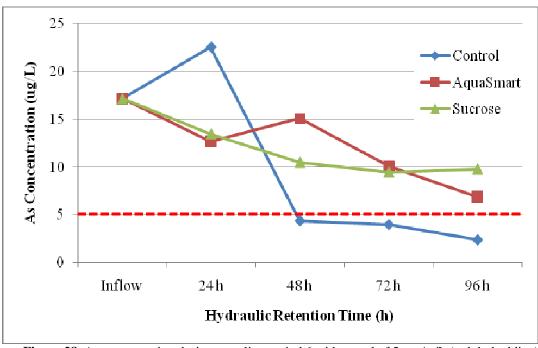
**Figure 25.** As concentration during sampling period 3 with a goal of 5  $\mu$ g As/L (red dashed line)



**Figure 26.** As concentration during sampling period 4 with a goal of 5  $\mu$ g As/L (red dashed line)



**Figure 27.** As concentration during sampling period 5 with a goal of 5  $\mu$ g As/L (red dashed line)



**Figure 28.** As concentration during sampling period 6 with a goal of 5  $\mu$ g As/L (red dashed line)

#### Pilot-Scale CWTS for PW Containing Oil & Grease and Divalent Metals

To determine treatment performance of the pilot-scale CWTSs, water samples were collected from five sampling locations in each wetland series. Sampling locations included inflow to the CWTS and after each cell A, B, C, and D (final effluent). Samples were collected following a sampling interval reflecting the HRT (24h). Samples collected were analyzed for total concentrations of metals (Fe, Mn, Ni, and Zn), oil and grease (O&G) concentration, and other water quality (explanatory) parameters. Total metal concentrations were determined by inductively coupled plasma-atomic emissions spectrometry (ICP-AES, EPA Method 200.7). Quality assurance and control measures included standard additions. Oil and grease concentration was determined by extraction with n-hexane using a StepSaver apparatus manufactured by Environmental Express (modification of EPA Method 1664A). Explanatory parameters were measured using Standard Methods (APHA, 1998) and included measurements of temperature, pH, conductivity, alkalinity, hardness, and dissolved oxygen. Oxidation-reduction (redox) potential of wetland hydrosoil was measured using milli-volt meters, an Accumet® calomel reference electrode, and in situ platinum-tipped electrodes (Faulkner et al., 1989).

Explanatory parameters including temperature, pH, conductivity, alkalinity, hardness, and DO for sampling period 1-6 (Tables 45-50) were measured in the CWTSs because of their influence on wetland system pathways and constituent behavior. The same amount of NaHCO<sub>3</sub>, MgSO<sub>4</sub>•7H<sub>2</sub>O, KNO<sub>3</sub>, and CaCO<sub>3</sub> was added to the simulated PW over the study period. These chemical sources were added to simulate ionic composition, hardness, and pH of PW. The pilot-scale CWTSs were located in a greenhouse during the experiments, the water temperature ranged from approximately 19-27°C over the course of the study.

Redox potential was measured in each cell. Redox conditions affect chemical and microbial processes, and can greatly influence the biological availability of major and trace nutrients (Kadlec and Wallace, 2009). Measured redox potential in cell A of the SSF1 system indicates oxidizing conditions (246 mV). Cells B and C of the SSF1 system had variable redox values (-51 and -14 mV), and cell D indicates oxidizing conditions (98 mV). For the FWS series redox potential in cell A indicates oxidizing conditions (82 mV), cells B and C range from slightly reducing (-55 mV) to slightly oxidizing (25 mV), and cell D is oxidizing (80 mV). Dissolved oxygen concentrations for sampling period 1 decreased from 7.99 mg/L measured in the influent to (5.89, 6.5, & 7.38 mg/L) measured after the first cell of each respective system (Table 45).

For sampling periods 4-6 water temperature ranged from approximately 26-28°C over the course of the study. For sampling period 4, redox potential measured in cell A of the SSF1 system indicated oxidizing conditions (210 mV). Redox potential ranged from -28 to -101 mV in cells B and C of the SSF1 system, and redox was 189 mV in cell D. For the FWS series oxidizing conditions (99 mV) were measured in cell A, reducing (-101 mV) to slightly oxidizing (49 mV) conditions in cells B and C, and oxidizing conditions (102 mV) in cell D. For each respective series dissolved oxygen concentrations for sampling period 4 decreased from 7.72 mg/L in the influent to 6.23, 7.1, and 6.89 mg/L in the effluent of cell A (Table 48). Explanatory parameters for sampling periods 5 and 6 exhibited trends similar to those observed in sampling period 4.

The concentration of oil and grease (O&G) decreased to below guideline concentrations for irrigation and livestock watering after treatment in the pilot-scale CWTSs (HRT = 4 days) for the subsurface flow series and free-water surface series operating at 50 mg/L (Table 47) and 100 mg/L O&G target influent (Tables 45,46 and 48-50). At 50 mg/L influent O&G concentration, a 24-hr HRT was sufficient to meet guidelines for irrigation and livestock watering.

The concentrations of iron, manganese, nickel, and zinc in simulated PW decreased to below irrigation and livestock watering guidelines after treatment in the pilot-scale CWTSs (HRT 4-days). Removal of iron, manganese, nickel, and zinc was greater in subsurface systems compared to that of the free water system.

**Table 45.** Fe, Mn, Ni, Zn, O&G concentrations and explanatory parameters for Sampling Period 1

		Pe	rformanc	e Parame	ters			E	xplanator	y Param	eters	
											Alkalinity	_
	[Fe]	[Mn]	[Ni]	[Zn]	[Oil & Grease]	DO	Cond.	pН	Redox	Temp	(mg/L	Hardness
Sample	(μg/L)	(µg/L)	(µg/L)	(mg/L)	(mg/L)	(mg/L)	(µS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	(mg/L CaCO <sub>3</sub> )
Inflow	547.0	480.7	346.7	6.48	97.6	7.99	199	7.16	nd	24.2	38	45
FWS A	182.5	151.7	308.4	4.35	11.5	5.89	201	6.62	82	26.9	40	36
FWS B	183.2	129.7	267.0	2.23	7.6	4.58	204	6.79	-55	26.5	45	38
FWS C	84.4	34.5	273.6	1.65	4.8	4.61	207	6.81	25	27.1	42	37
FWS D	58.8	16.1	266.7	1.04	3.2	7.65	210	6.92	80	27.4	48	41
SSF1 A	46.4	6.0	255.3	0.566	25.4	6.5	200	6.88	246	26.6	35	21
SSF1 B	25.0	2.9	221.9	0.153	13.8	7.1	204	6.79	-51	26.8	39	25
SSF1 C	15.2	2.0	152.3	0.051	8.6	7.13	209	6.84	-14	26.8	31	28
SSF1 D	16.7	2.9	116.7	0.023	4.2	7.38	215	6.94	98	26.3	47	37
SSF2 A	40.6	6.8	249.6	0.454	30.6	6.21	213	6.83	212	24.1	39	28
SSF2 B	18.7	3.2	193.5	0.144	18.9	6.15	227	6.89	-65	25.3	45	31
SSF2 C	16.5	2.9	154.2	0.060	9.2	7.1	222	6.95	15	25.7	35	35
SSF2 D	14.5	2.4	117.9	0.029	3.8	6.89	229	6.78	73	24.9	41	41

**Table 46.** Fe, Mn, Ni, Zn, O&G concentrations and explanatory parameters for Sampling Period 2

	Performance Parameters							Ex	kplanator	y Paramet	ers	
	[Fe]	[Mn]	[Ni]	[Zn]	[Oil & Grease]	DO	Cond.	рН	Redox	Temp	Alkalinity (mg/L	Hardness (mg/L
Sample	$(\mu g/L)$	$(\mu g/L)$	$(\mu g/L)$	(mg/L)	(mg/L)	(mg/L)	(µS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	CaCO <sub>3</sub> )
Inflow	296.5	409.6	312.8	4.74	94.2	7.91	214	6.87	nd	24.4	45	42
FWS A	157.3	183.6	243.4	3.54	13.4	5.55	216	6.45	32	25.9	41	29
FWS B	81.0	113.0	246.0	3.63	10.1	5.82	211	6.52	-79	26.4	43	35
FWS C	70.8	53.3	219.5	3.16	6.9	6.13	210	6.66	-101	26.6	47	33
FWS D	71.7	32.4	180.7	2.56	2.3	6.01	217	6.59	15	25.9	44	38
SSF1 A	18.5	3.4	221.2	2.64	37.2	6.31	210	7.01	121	24.1	39	28
SSF1 B	16.8	2.7	190.6	2.05	19.1	6.14	209	6.95	-112	25.9	48	33
SSF1 C	13.0	3.3	135.6	1.15	12.9	6.41	216	6.83	-130	26.1	45	38
SSF1 D	10.9	3.0	109.0	0.791	5.9	6.87	213	6.91	-54	25.1	51	41
SSF2 A	19.5	3.6	207.1	2.60	12.9	6.5	217	6.42	98	23.9	26	31
SSF2 B	14.6	2.9	175.7	1.97	11.3	6.34	223	6.66	-82	24.1	28	29
SSF2 C	10.8	2.7	147.0	1.48	8.2	6.79	221	6.49	-23	23.5	37	37
SSF2 D	13.3	2.7	112.1	0.955	4.7	7.07	226	6.78	11	23.1	35	41

 Table 47. O&G concentrations and explanatory parameters for Sampling Period 3

	Performance Parameters	Explanatory Parameters								
							Alkalinity	Hardness		
		DO	Cond.	pН	Redox	Temp	(mg/L	(mg/L		
Sample	[Oil & Grease] (mg/L)	(mg/L)	(µS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	CaCO <sub>3</sub> )		
Inflow	52.3	8.17	173	6.1	nd	20.1	29	44		
FWS A	9.9	7.56	176	6.19	-120	20.9	31	42		
FWS B	4.3	7.39	174	6.04	-23	20.4	27	30		
FWS C	3.6	7.73	170	6.21	31	20.3	33	37		
FWS D	2.2	8.02	178	6.13	40	19.9	39	48		
SSF1 A	12.1	6.77	176	6.03	-72	19.7	22	41		
SSF1 B	10.3	6.97	172	6.15	-99	20.4	25	29		
SSF1 C	7.6	6.63	180	6.17	-31	20.7	27	33		
SSF1 D	2.1	7.23	178	6.21	41	20	31	30		
SSF2 A	12.6	6.03	198	6.09	-69	19.8	45	32		
SSF2 B	9.8	5.99	201	6.11	-201	20.1	39	37		
SSF2 C	4.5	6.19	195	6.23	-132	21.2	53	34		
SSF2 D	2.9	6.31	208	6.2	-29	20.6	47	35		

Table 48. Fe, Mn, Ni, Zn, O&G concentrations and explanatory parameters for Sampling Period 4

		Pe	rformanc	e Parame	ters			E	xplanator	y Param	eters	
					_						Alkalinity	
	[Fe]	[Mn]	[Ni]	[Zn]	[Oil & Grease]	DO	Cond.	pН	Redox	Temp	(mg/L	Hardness
Sample	(µg/L)	(µg/L)	(µg/L)	(mg/L)	(mg/L)	(mg/L)	(µS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	(mg/L CaCO <sub>3</sub> )
Inflow	286.7	510.2	322.2	5.23	101.2	7.72	221	7.21	nd	27.1	42	40
FWS A	179.6	193.2	299.2	4.89	29.1	6.23	215	6.88	99	26.5	49	41
FWS B	163.2	152.1	278.2	3.67	17.1	4.98	214	6.67	-101	26.9	42	48
FWS C	97.8	59.7	261.9	2.01	8.9	4.99	203	6.72	49	27.6	47	39
FWS D	64.4	31.1	255.2	1.01	7.1	7.78	212	6.85	102	27.9	46	43
SSF1 A	52.2	9.2	270.1	0.633	28.2	7.1	208	6.91	210	27.1	39	32
SSF1 B	29.7	5.6	211.1	0.545	19.1	7.21	209	6.85	-28	27.2	41	29
SSF1 C	18.2	6.1	189.1	0.314	11.1	7.78	213	6.92	-101	26.9	48	27
SSF1 D	15.2	4.3	139.2	0.289	5.6	7.67	217	6.99	189	27.1	39	34
SSF2 A	38.7	5.1	263.3	0.685	33.2	6.89	223	6.93	267	26.9	34	26
SSF2 B	21.4	3.9	201.2	0.233	15.2	6.76	229	7.11	-103	26.1	31	36
SSF2 C	19.7	2.8	170.4	0.123	10.9	7.98	231	6.92	65	26.6	39	39
SSF2 D	12.1	3.1	138.2	0.111	4.2	6.57	227	6.89	99	25.9	44	31

**Table 49.** Fe, Mn, Ni, Zn, O&G concentrations and explanatory parameters for Sampling Period 5

		Pe	rformanc	e Parame	ters			E	kplanator	y Paramet	ers	
0 1	[Fe]	[Mn]	[Ni]	[Zn]	[Oil & Grease]	DO	Cond.	рН	Redox	Temp	Alkalinity (mg/L	Hardness (mg/L
Sample	(µg/L)	(µg/L)	(μg/L)	(mg/L)	(mg/L)	(mg/L)	(μS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	CaCO <sub>3</sub> )
Inflow	320.4	463.2	367.9	6.23	95.9	7.44	209	6.81	nd	26.4	41	37
FWS A	190.5	224.1	262.2	4.52	15.2	6.01	201	6.55	-56	26.3	48	35
FWS B	178.9	109.3	239.3	3.09	9.5	6.11	205	6.51	-160	26.9	41	39
FWS C	104.3	81.4	199.5	2.58	5.4	6.19	209	6.76	-23	27.1	39	32
FWS D	73.2	50.2	191.3	1.97	4.2	5.99	213	6.81	67	27.3	43	37
SSF1 A	98.2	9.7	245.6	1.21	31.1	6.22	207	6.99	48	27.2	37	29
SSF1 B	31.5	8.9	209.8	0.735	22.8	6.29	211	6.89	-100	26.9	41	36
SSF1 C	17.7	9.3	142.2	0.747	18.1	6.19	213	7.01	45	26.6	44	35
SSF1 D	12.2	4.7	129.2	0.587	9.9	6.73	219	7.03	10	27.5	48	39
SSF2 A	29.9	7.8	227.2	1.5	21.1	5.87	212	6.58	101	26.2	29	33
SSF2 B	23.2	7.9	174.9	0.897	19.4	6.15	217	6.71	-120	26.5	31	26
SSF2 C	11.1	3.9	153.2	0.777	15.3	6.66	225	6.88	-89	26.5	28	31
SSF2 D	10.4	2.1	135.1	0.619	7.2	6.92	221	6.52	49	26.8	31	42

**Table 50.** O&G concentrations and explanatory parameters for Sampling Period 6

	Performance Parameters	Explanatory Parameters								
							Alkalinity	Hardness		
		DO	Cond.	pН	Redox	Temp	(mg/L	(mg/L		
Sample	[Oil & Grease] (mg/L)	(mg/L)	(µS/cm)	(S.U.)	(mV)	(°C)	CaCO <sub>3</sub> )	CaCO <sub>3</sub> )		
Inflow	99.7	8.09	189	6.5	nd	26.7	47	49		
FWS A	16.6	6.47	187	6.37	-87	26.2	39	40		
FWS B	12.4	6.41	181	6.29	25	27.1	42	39		
FWS C	7.8	6.81	179	6.41	109	27.5	43	45		
FWS D	2.9	6.99	183	6.39	150	27.2	43	42		
SSF1 A	15.2	6.03	186	6.6	67	26.8	45	50		
SSF1 B	11.4	6.25	181	6.57	-102	26.8	39	49		
SSF1 C	6.9	5.89	192	6.61	-15	26.2	41	48		
SSF1 D	3.1	6.32	194	6.68	54	27.1	42	46		
SSF2 A	18.9	6.77	193	6.52	35	26.9	40	42		
SSF2 B	13.9	6.55	188	6.5	-233	26.5	37	38		
SSF2 C	9.4	6.31	181	6.48	-65	26.8	39	36		
SSF2 D	5.3	6.69	191	6.41	13	27.2	44	40		

### Assess Pilot-Scale Performance (Task 6)

The objective of Task 6 is to assess performance of the pilot-scale CWTSs by (1) calculating removal rate coefficients, removal efficiencies, and removal extents and (2) comparing removal extents with selected reuse criteria.

Removal rate coefficients were calculated to determine the efficiency and effectiveness of each system's design in terms of removal processes and pathways and to allow for determination of critical design parameters including HRT. Removal rate coefficients (*k*) were calculated assuming first order rate kinetics (Equation 4).

Removal rate coefficient 
$$(k) = \frac{-\ln([C]/[C_o])}{t}$$

Eqn. 4

where,  $[C_0]$  is the initial inflow concentration (mg/L), [C] is the system outflow concentration (mg/L), and t is the time (hours) and corresponds to HRT of the system. Negative values for removal rates occur when outflow concentrations are greater than inflow concentrations. Removal efficiencies, defined as the percent decrease in outflow concentration relative to inflow concentration were calculated (Equation 5).

Removal efficiency (%) = 
$$\frac{[C_o] - [C]}{[C_o]} x 100$$

Eqn. 5

where,  $[C_0]$  is the initial inflow concentration (mg/L) and [C] is the outflow concentration (mg/L). Removal extent is defined as the outflow concentration, and can be compared with selected reuse criteria to determine the success of the system performance.

Pilot-Scale CWTS for Simulated Post-RO Produced Water

Removal rates, efficiencies, and extents were calculated for ammonia for all sampling periods using Equations 4 and 5 (Table 51). Removal rate efficiencies were greater in the experimental system during all sampling periods indicating that ammonia removal by the targeted removal pathways including nitrification can be enhanced when the correct operating conditions are applied (Table 20). Removal extents suggest that the concentration ammonia in simulated post-RO produced can be reduced to meet irrigation and livestock watering criteria.

**Table 51.** Ammonia removal extents, efficiencies, and rate coefficients for all sampling periods.

Sampling Period	Parameter Parameter	Conc. (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
	Influent	20.9		
1	Control Extent	17.6	15.8	0.043
	Experimental Extent	2.3	89.0	0.276
	Influent	21.2		
2	Control Extent	16.7	21.2	0.060
	Experimental Extent	7.8	63.2	0.125
	Influent	20.3		
3	Control Extent	14.3	29.6	0.088
	Experimental Extent	7.4	63.5	0.126
	Influent	22.3		
4	Control Extent	12.6	43.5	0.071
	Experimental Extent	< 0.1	>99.6	1.35
	Influent	23.2		
5	Control Extent	15	35.3	0.073
	Experimental Extent	< 0.1	>99.6	1.36*
	Influent	25.7		
6	Control Extent	13.2	48.6	0.111
	Experimental Extent	< 0.1	>99.6	1.39*

<sup>\*</sup>Rate coefficient calculated for 6 days of treatment; concentrations in cells 3 and 4 were identical.

Pilot-Scale CWTS for PW Containing Divalent Metals and Low Molecular Weight Organics

Removal rates, efficiencies, and extents of Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs in the pilot-scale system were calculated to determine the efficiency and effectiveness of the system design in terms of removal processes and pathways for each cell using equations 4 and 5 (Table 52). Removal rate coefficients were calculated assuming first order kinetics; however, upon examination, not all data fit first order (i.e. Figures 9-10).

In sampling period 1, both series had calculated removal efficiencies less than 56% for COCs removed from simulated fresh PW. These removal efficiencies may be attributed to lack of maturity of the system. For sampling periods 2 through 5, majority of the removal efficiency values were 80% or greater. The increase from 56% to 80% removal efficiency for the series may be a result of maturity and acclimation to the simulated fresh PW in addition to increased microbial activity (i.e. dissimilatory sulfate reducers and heterotrophic aerobic microbes). Removal extent and beneficial use criteria and guidelines were compared to determine if the effluent from the CWTS met or exceeded the use criteria (Table 53). For 70% of the sampling periods, copper concentrations met or were below all of the beneficial use guidelines and criteria. Cadmium concentrations met the beneficial uses for two sampling periods, while 80% of the nickel concentrations were below the surface water discharge criteria and 30% met the irrigation

guidelines. Every zinc concentration met the livestock watering criteria, and irrigation guidelines and surface discharge criteria were met for all sampling periods except for sampling period 1. Concentrations of LMWOs were below all of the beneficial use guidelines and criteria for every sampling period.

Pilot-Scale CWTS for PW Containing Metalloids and Low Molecular Weight Organics

Removal rate coefficients, removal efficiencies, and removal extents of selenium and arsenic in the pilot-scale system were calculated to determine the efficiency and effectiveness of the system design in terms of removal processes and pathways for each cell using equations 4 and 5 (Table 54).

The goal of 5  $\mu$ g Se/L in the outflow water was achieved and maintained in the pilot-scale CWTS with the AquaSmart<sup>TM</sup> amendment. The sucrose amendment also achieved the goal of 5  $\mu$ g Se/L, but with less consistency than the AquaSmart<sup>TM</sup> amendment. The untreated control series did not meet the 5  $\mu$ g Se/L goal during any sampling periods. The goal of 5  $\mu$ g As/L in the outflow water was achieved by both the untreated control series and the AquaSmart<sup>TM</sup> series.

Pilot-Scale CWTS for PW Containing Oil & Grease and Divalent Metals

The concentrations of iron, manganese, nickel, and zinc in simulated OFPW decreased to below irrigation and livestock watering guidelines by treatment in the pilot-scale CWTSs (HRT 4-days) designed for removal of oil and grease (0&G). Removal efficiencies (%) and removal rate coefficients (k) for iron, manganese, nickel, and zinc were greater in the subsurface flow series than in the free-water surface series (Table 55). Removal efficiencies and removal rate coefficients (k) were greater for iron, manganese, and zinc than for nickel (Table 55).

The concentration of O&G in simulated OFPW decreased to below irrigation and livestock watering guidelines by treatment in the pilot-scale CWTSs (HRT 4-days) in the subsurface flow series and free-water surface series operating at 50 and 100mg/L O&G target influent (Table 56). There was no discernable difference in O&G removal, measured through removal rate coefficients (*k*) and removal efficiencies (%), between the subsurface flow and free-water surface series. Removal was greatest in the first cell of each series, with oil and grease concentrations reduced by fifty percent (from influent concentrations) in effluent samples collected from the first cell of each system. Effluent samples collected from the final cell of each series increased through the duration of the experiment; however, these respective concentrations remained below irrigation and livestock watering guidelines.

**Table 52.** Removal Efficiency Calculations for Cd<sup>+2</sup>, Cu<sup>+2</sup>, Ni<sup>+2</sup>, Zn<sup>+2</sup>, and LMWOs in the Pilot-scale CWTS

		Copper			Cadmiun	1		Nickel			Zinc			LMWO	s
	Rate		Outflow	Rate	Rem-	Outflow	Rate	Rem-	Outflow	Rate	Rem-	Outflow	Rate		Outflow
Sampling 1	Coef I	Removal (%)	Conc. (mg/L)	Coef (d <sup>-1</sup> )	oval (%)	Conc. (mg/L)	Coef (d <sup>-1</sup> )	oval (%)	Conc. (mg/L)	Coef (d <sup>-1</sup> )	oval (%)	Conc. (mg/L)	Coef (d <sup>-1</sup> )	Removal (%)	Conc. (mg/L)
Series 1	0.007	50.5%	1.98	0.005	38.9%	3.6	0.009	55.8%	2.74	0.004	32.8%	4.1	0.082	99.9%	bdl
Series 2	0.005	39.3%	2.43	0.005	36.4%	3.75	0.0007	6.5%	5.8	0.003	26.2%	4.5	0.082	99.9%	bdl
Sampling 2															
Series 1	0.047	98.9%	0.04	0.019	84.7%	0.82	0.013	71.5%	1.87	0.014	73.7%	1.52	0.082	99.9%	bdl
Series 2	0.044	98.6%	0.05	0.02	85.2%	0.79	0.015	75.6%	1.6	0.014	73.9%	1.51	0.082	99.9%	bdl
Sampling 3															
Series 1	0.011	63.8%	0.54	0.029	94.3%	0.25	0.02	85.7%	0.61	0.014	73.3%	1.22	0.082	99.9%	bdl
Series 2	0.033	95.9%	0.06	0.04	97.9%	0.09	0.04	97.9%	0.09	0.064	99.8%	0.01	0.082	99.9%	bdl
Sampling 4															
Series 1	0.009	57.5%	1.49	0.008	53.0%	2.09	0.014	72.8%	1.31	0.022	87.5%	0.44	0.082	99.9%	bdl
Series 2	0.049	99.1%	0.03	0.076	99.9%	< 0.003	0.044	98.5%	0.07	0.039	97.7%	0.08	0.082	99.9%	bdl
Sampling 5															
Series 1	0.037	97.2%	0.08	0.048	98.9%	0.02	0.02	85.8%	0.41	0.04	97.9%	0.08	0.082	99.9%	bdl
Series 2	0.052	99.3%	0.02	0.032	95.5%	0.09	0.045	98.6%	0.04	0.055	99.5%	0.02	0.082	99.9%	bdl

bdl - below detection limit

**Table 53.** Comparisons Between Removal Extent (mg/L) and Beneficial Use Guidelines and Criteria (mg/L).

		Copper	Nickel	Zinc	Cadmium	LMWOs
Sampling 1						
	Series 1	1.98	2.74	3.9	3.6	<1.0
	Series 2	2.43	5.8	4.5	3.75	<1.0
Sampling 2						
	Series 1	0.04	1.87	1.52	0.82	<1.0
	Series 2	0.05	1.6	1.51	0.79	<1.0
Sampling 3						
	Series 1	0.54	0.61	1.22	0.25	<1.0
	Series 2	0.06	0.09	0.01	0.09	<1.0
Sampling 4						
	Series 1	1.49	1.31	0.44	2.09	<1.0
	Series 2	0.03	0.07	0.08	< 0.003	<1.0
Sampling 5						
	Series 1	0.084	0.41	0.08	0.02	<1.0
	Series 2	0.02	0.04	0.02	0.09	<1.0
Irrigation (	Guidelines	0.2	0.2	2	0.01	35
Livestock Guidelines Surface Discharge		0.5	-	24	0.05	35
Crite		0.37-1.0	2	0.4-5	-	15

**Table 54.** Selenium and arsenic removal extents, efficiencies, and rate coefficients for all sampling periods.

Table 54. Scienta	m and arsenic removal ext	ciits, ciiicici	Selenium	c cocificien	101 411 54	Arsenic	ous.
			Selemum	D (		Arsenic	
		Conc.	Removal	Rate	Conc.	Removal	Rate Coef.
Sampling Period	Parameters	$(\mu g/L)$	(%)	Coef. (d <sup>-1</sup> )	$(\mu g/L)$	(%)	$(d^{-1})$
	Influent	44.6			21.8		
1	Control Extent	45.7	-2.5	-0.006	5.6	74.3	0.340
1	AquaSmart <sup>TM</sup> Extent	nd	$97.8^{1}$	1.27	3.3	84.9	0.472
	Sucrose Extent	6.1	86.3	0.497	13.4	38.5	0.122
	Influent	49.7			14.0		
2	Control Extent	44.6	10.3	0.027	5.0	64.3	0.257
2	AquaSmart <sup>TM</sup> Extent	nd	98.0	0.977	6.3	55.0	0.200
	Sucrose Extent	8.1	83.7	0.454	5.5	60.7	0.234
	Influent	46.5			18.4		
3	Control Extent	37.9	18.5	0.051	10.9	40.8	0.131
3	AquaSmart <sup>TM</sup> Extent	1.0	$97.8^{1}$	1.28	7.7	58.2	0.218
	Sucrose Extent	7.3	84.3	0.463	5.1	72.3	0.321
	Influent	42.7			18.4		
4	Control Extent	39.7	7.0	0.018	6.0	67.4	0.280
4	AquaSmart <sup>TM</sup> Extent	1.0	$97.7^{1}$	1.25	9.0	51.1	0.179
	Sucrose Extent	1.0	97.7	0.939	6.3	65.8	0.268
	Influent	76.0			18.3		
5	Control Extent	58.2	23.4	0.067	4.3	76.5	0.362
J	AquaSmart <sup>TM</sup> Extent	1.0	$98.7^{2}$	2.17	9.7	47.0	0.159
	Sucrose Extent	5.8	92.4	0.643	3.5	80.9	0.414
	Influent	50.8			17.2		
6	Control Extent	56.4	-11.0	-0.026	2.4	86.0	0.492
υ	AquaSmart <sup>TM</sup> Extent	1.0	98.0	0.982	6.9	59.9	0.228
	Sucrose Extent	2.9	94.3	0.716	9.8	43.0	0.141

<sup>&</sup>lt;sup>1</sup> Rate coefficient calculated for 3 days of treatment since concentrations in cells 3 and 4 were non-detectable. The rate coefficient was calculated for the HRT during which removal was measured.

<sup>&</sup>lt;sup>2</sup> Rate coefficient calculated for 2 days of treatment since concentrations in cells 2, 3, and 4 were non-detectable. The rate coefficient was calculated for the HRT during which removal was measured.

**Table 55.** Metal concentrations measured in CWTS influent, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiences and removal rate coefficients were calculated using measured concentrations in the effluent (4-day HRT).

			Iron			Manganes	se		Nickel			Zinc	
	•	Conc. (µg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )	Conc. (µg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )	Conc. (µg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )	Conc. (µg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
ıt	Influent	547.0			480.7			346.7			6480		
Treatment Period 1	FWS Effluent	58.8	89.3	0.523	16.1	96.7	0.827	266.7	23.1	0.064	1040	84.0	0.463
reatme Period	SSF1 Effluent	16.7	96.9	0.809	2.9	99.4	1.13	116.7	66.3	0.269	23	99.6	1.37
Tr P	SSF2 Effluent	14.5	97.3	0.816	2.4	99.5	1.15	117.9	66.0	0.269	29	99.6	1.28
5 nt	Influent	296.5			409.6			312.8			4740		
	FWS Effluent	71.7	75.8	0.364	32.4	92.1	0.631	180.7	42.2	0.120	2560	46.0	0.135
reatme Period	SSF1 Effluent	10.9	96.3	0.696	3.0	99.3	0.986	109.0	65.2	0.260	791	83.3	0.441
Tr P	SSF2 Effluent	13.3	95.5	0.680	2.7	99.3	1.03	112.1	64.2	0.240	955	79.9	0.377
t t	Influent	286.7			510.2			322.2			5230		
Treatment Period 4	FWS Effluent	64.4	77.5	0.359	31.1	93.9	0.677	255.2	20.8	0.060	1010	80.7	0.418
reatmo Period	SSF1 Effluent	15.2	94.7	0.693	4.3	99.2	0.996	139.2	56.8	0.204	289	94.5	0.649
T A	SSF2 Effluent	12.1	95.8	0.701	3.1	99.4	1.08	138.2	57.1	0.213	111	97.9	0.942
o tr	Influent	320.4			463.2			367.9			6230		
me od 5	FWS Effluent	73.2	77.2	0.356	50.2	89.2	0.546	191.3	48.0	0.158	1970	68.4	0.286
Treatment Period 5	SSF1 Effluent	12.2	96.2	0.825	4.7	99.0	0.922	129.2	64.9	0.264	587	90.6	0.521
Ti P	SSF2 Effluent	10.4	96.8	0.785	2.1	99.5	1.15	135.1	63.3	0.240	619	90.1	0.528

**Table 56.** Total O & G concentrations measured in CWTS influent, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the effluent (4-day HRT).

			Oil and Grease	
	•	Conc. (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
ti _	Influent	97.6		
me	FWS Effluent	3.2	96.7	0.771
Treatment Period 1	SSF1 Effluent	4.2	95.7	0.738
T	SSF2 Effluent	3.8	96.1	0.769
nt 7	Influent	94.2		
Treatment Period 2	FWS Effluent	2.3	97.6	0.809
reatme Period	SSF1 Effluent	5.9	93.7	0.660
T H	SSF2 Effluent	4.7	95.0	0.645
nt 3	Influent	52.3		
Treatment Period 3	FWS Effluent	2.2	95.8	0.735
reatm Period	SSF1 Effluent	2.1	96.0	0.690
T	SSF2 Effluent	2.9	94.5	0.681
t t	Influent	101.2		
Treatment Period 4	FWS Effluent	7.1	93.0	0.650
reat Peri	SSF1 Effluent	5.6	94.5	0.672
T T	SSF2 Effluent	4.2	95.8	0.748
s nt	Influent	95.9		
me od .	FWS Effluent	4.2	95.6	0.729
Treatment Period 5	SSF1 Effluent	9.9	89.7	0.508
T	SSF2 Effluent	7.2	92.5	0.550
ot S	Influent	99.7		
me od (	FWS Effluent	2.9	97.1	0.783
Freatment Period 6	SSF1 Effluent	3.1	96.9	0.773
T.	SSF2 Effluent	5.3	94.7	0.657

# <u>Design and Construct Demonstration Constructed Wetland Treatment System (Task 7)</u>

Specific experiments in support of designing a demonstration system were performed to gain a better understanding of conditions affecting treatment pathways and hybrid design strategies that can be incorporated into constructed wetland treatment. These experiments provide insight into potential design strategies that may be used for the construction of demonstration and full-scale systems. The experiments, which are discussed below, are:

- 1. Addition of zeolite for sorption of ammonia from produced water;
- 2. Effects of biogeochemical conditions and processes on vertical distribution of metals in hydrosoil;
- 3. Effects of water depth on conditions and treatment performance;
- 4. Effects of seasonal variation on treatment performance;
- 5. Addition of a nutrient amendment for removal of selenium from produced water;
- 6. Addition of an oil/water separator for removal of oil and grease; and
- 7. Evapotranspiration experiments and tracer tests.

# Sorption Experiments

Sorptive materials may be used to remove dissolved ammonia from water. Transfer from the aqueous phase to binding sites may enhance performance in constructed wetland treatment systems (CWTSs) by concentrating ammonia where nitrifying bacteria (*Nitrosomonas* spp. and *Nitrobacter* spp.) may be active. The zeolite clinoptilolite was selected as a sorptive medium for use in CWTSs based on its affinity for ammonia and high surface area for growth and attachment of periphytic biofilms. The objective of this research was to determine the effects of clinoptilolite on ammonia treatment performance in pilot-scale CWTSs.

A serial batch sorption experiment was performed to determine the ammonia ion exchange capacity of clinoptilolite. Data from the serial batch sorption experiment were used to predict ammonia removal in response to clinoptilolite added (1000 and 500 g) to a pair of pilot-scale CWTSs containing sandy sediment. After 50 days, nitrifying bacterial activity reactivity tests (n-BARTs) were used to detect the presence of nitrifying bacteria in the added clinoptilolite and sandy sediment.

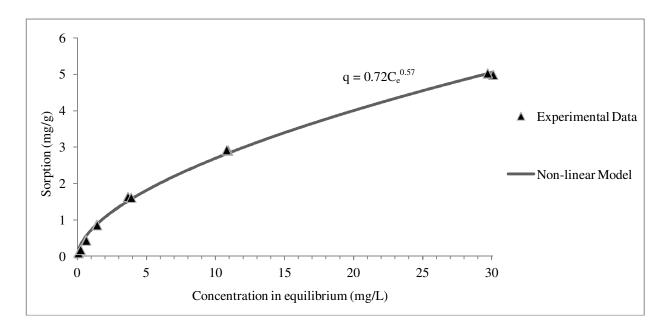
Results indicate that clinoptilolite has an affinity for ammonia described by the Freundlich equation Q=0.72C<sub>e</sub><sup>0.57</sup> for equilibrium ammonia-N concentrations from 0.07 to 30.1 mg/L (Table 57 and Figure 29). Addition of 1000g clinoptilolite to a pilot-scale CWTS operating on a 2-day hydraulic retention time (HRT) and containing *Schoenoplectus californicus* produced a measurable decrease in the outflow concentration of ammonia-N when compared with a system containing no clinoptilolite (2.0 vs. 5.6 mg/L) during a 10-day sampling period (Figure 30). Addition of 500g clinoptilolite to a CWTS operating on 4-day HRT and containing *Typha latifolia* produced no discernable difference in the outflow concentration of ammonia compared with a system containing no clinoptilolite (Figure 31). n-BARTs indicated similar levels of activity in both the sediment and clinoptilolite in the *Schoenoplectus* system. No activity was detected in either the sediment or clinoptilolite in the *Typha* system. The difference in performance between the *Schoenoplectus* and *Typha* 

systems was attributed to factors including dissolved oxygen and equilibrium ammonia concentrations. This work demonstrates that clinoptilolite can be effective for increasing ammonia removal and nitrifying activity in pilot-scale CWTSs.

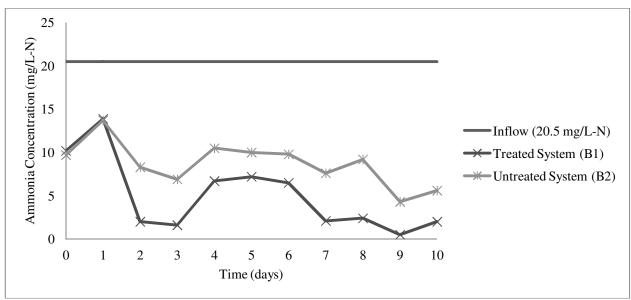
**Table 57.** Resultant equilibrium concentrations (C<sub>e</sub>) and calculated sorption values (q) from sealed 300mL

BOD bottles containing ammonia solution (C<sub>o</sub>) and clinoptilolite (M<sub>c</sub>).

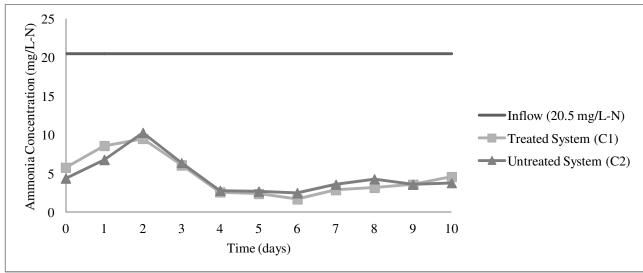
D 441	C ( /I)	C ( T)		( 1 )	T ()	т
Bottle	C <sub>o</sub> (mg/L)	C <sub>e</sub> (mg/L)	$M_{c}(g)$	q (mg/g)	Log C <sub>e</sub>	Log q
1	1	< 0.1	3.00	0.09	-1.00	-1.05
2	1	< 0.1	3.00	0.09	-1.00	-1.05
3	2	0.21	3.00	0.18	-0.68	-0.75
4	2	0.22	3.00	0.18	-0.66	-0.75
5	5	0.59	3.00	0.44	-0.23	-0.36
6	5	0.63	3.00	0.44	-0.20	-0.36
7	10	1.38	3.00	0.86	0.14	-0.06
8	10	1.42	3.00	0.86	0.15	-0.07
9	20	3.65	3.00	1.63	0.56	0.21
10	20	3.89	3.00	1.61	0.59	0.21
14	40	10.9	3.00	2.91	1.04	0.46
15	40	10.8	3.00	2.92	1.03	0.47
16	80	30.1	3.00	4.99	1.48	0.70
17	80	29.7	3.00	5.03	1.47	0.70



**Figure 29.** Non-linearly derived Freundlich sorption isotherms generated from serial batch sorption experiment (Table 57).



**Figure 30.** Comparison of outflows from treated and untreated (B1 and B2) *Schoenoplectus* systems during the 10-day monitoring period.



**Figure 31.** Comparison of outflows from treated and untreated (C1 and C2) *Typha* systems during the 10-day monitoring period.

## Biogeochemical Conditions and Processes

Environmental conditions in hydrosoil (sediment and overlying organic detritus) of a free water surface pilot-scale constructed wetland treatment system (CWTS) were investigated to determine their effects on sequestering Cd, Cu, Ni, and Zn. Redox potential, pH, organic matter content (OM), acid-volatile sulfide (AVS) concentration, simultaneously extracted metal (SEM) concentration, and carbonate content were measured in detritus grab samples and sediment cores extracted from two cells planted with *Schoenoplectus californicus* (cells 1 and 2) and two cells planted with *Typha angustifolia*.

The detritus consisted of an organic-rich layer (i.e. plant roots and plant litter) approximately 16 to 26 cm thick (Figure 32). The detritus-sediment interface was recognized by a distinct boundary at which dark, organic-rich detritus sharply overlaid lighter mineral-rich sediment. OM was 30-85% in the detritus, decreasing gradually downward to 0.65-3% at 15 cm below the top of the sediment (Figure 32). Reducing conditions (-249 to -34 mV) and pH between 5.01 and 7.41 were measured in the detritus and sediment of all four cells (Figure 32). Highest redox potential (-145 to -34 mV) and pH (6.28 to 7.41) occurred in the upper 6 cm of sediment. The greatest AVS concentration (36-160  $\mu$ mole/g) and greatest total SEM concentration (Cd, Cu, Ni, and Zn  $\geq$  5.45  $\mu$ mole/g) occurred in the detritus.

Measured values of redox potential, pH, OM, and AVS concentration in CWTS cells were in favorable ranges to promote sorption and complexation of Cd, Cu, Ni, and Zn with AVS (Table 58). Reducing conditions and acidic to circumneutral pH in the detritus and sediment are interpreted to have resulted from organic matter degradation, nutrient uptake by roots, and microbial processes. Hydrosoil amendments (hay and gypsum) to cells 1 and 2 promoted metal complexation with AVS in the sediment, which resulted in greater mean AVS and  $\sum$ SEM concentrations in sediment of cells 1 and 2 (planted with *S. californicus*) compared to cells 3 and 4 (planted with *T. angustifolia*).

Sequential extractions (after Tessier et al., 1979) and mineralogical analyses using a scanning electron microscopy (EM) equipped with energy dispersion spectroscopy (EDS) were performed on detritus and sediment samples to determine metal mobility and treatment processes in the system. The sequential extraction procedure (SEP) released metals into five operationally defined geochemical fractions: exchangeable; bound to carbonates; bound to Fe-Mn oxides, bound to organic matter or sulfides; and residual.

Metal concentrations in each fraction in the detritus and sediment are shown in Figure 33, and the sum of metal concentrations in each fraction in the detritus and sediment is shown in Figure 34. Metal fractionation and relative (percentage) importance with depth is shown in Figure 35. The fraction of all metals associated with the most mobile fraction (i.e. exchangeable) in each cell was low (< 2%; Figure 34). The high percent of metals associated with the Fe-Mn oxide (i.e. Fe-Mn hydroxide) fraction (22-42%) indicates oxidation and hydrolysis (Figure 34). OM and concentrations of Cd, Cu, Ni, and Zn decreasing with depth in the hydrosoil of each cell are consistent with sorption to organic matter (Figure 32). The high percent of metals bound to sulfides (39-46 %) confirmed by chemical extractions (SEP and AVS-SEM; Figure 32-35) and mineralogical analysis (scanning EM-EDS) indicates precipitation in the form of sulfide minerals. High AVS concentration (5.9-100 µmole/g) indicates that dissimilatory sulfate reduction occurred (Figure 32). 10 to 15% of all metals were associated with the residual fraction (Figure 34), which is unlikely to be released into the surface water from pH or redox potential changes. The metal fractionation results suggest that wetlands should be designed with careful consideration of metal fluxes (i.e. biogeochemical processes) with changes in hydrosoil conditions (i.e. redox potential and pH).

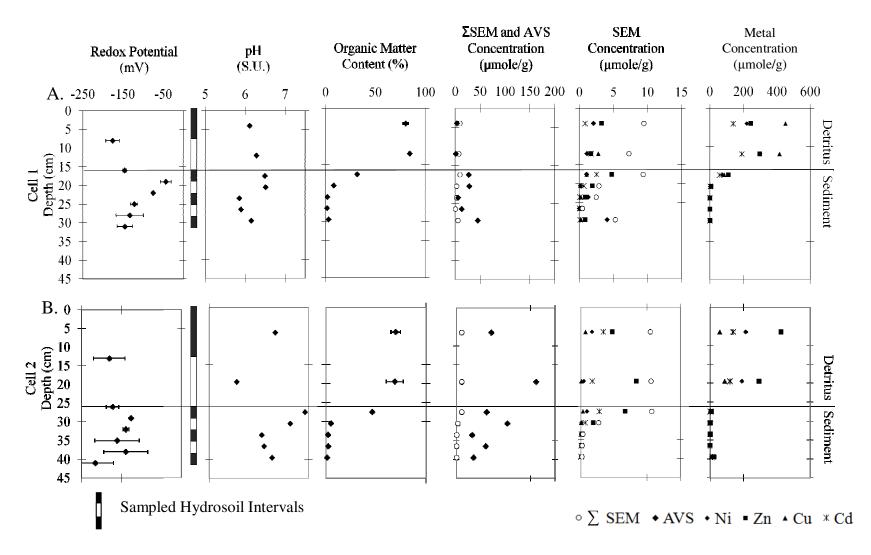


Figure 32 (continued on next page).

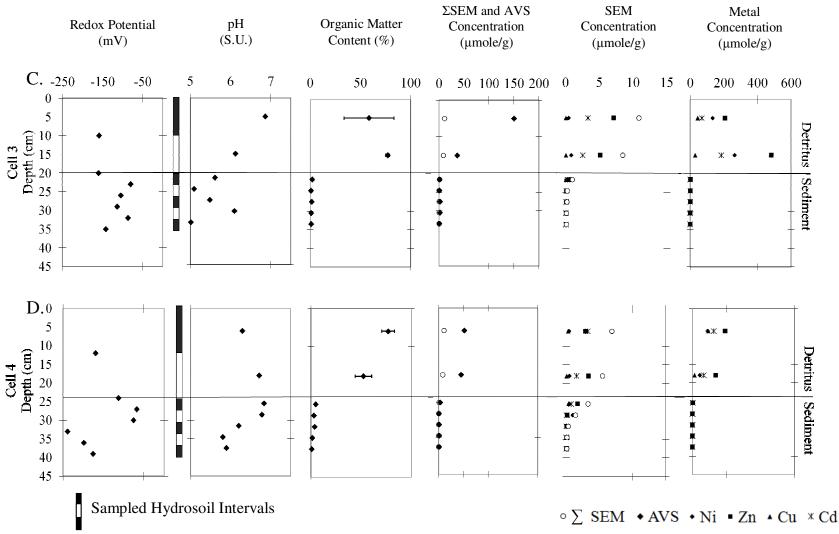
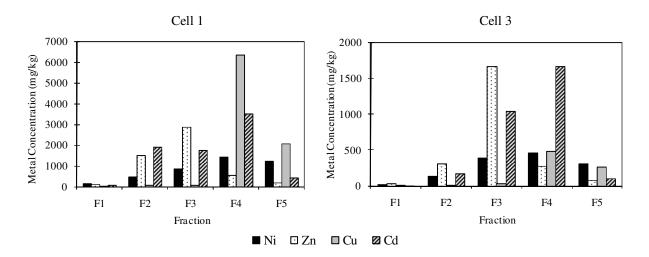
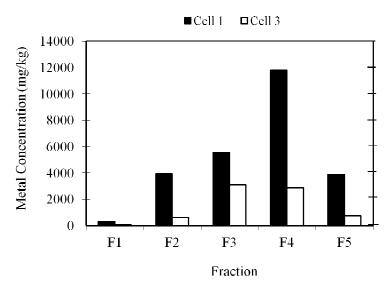


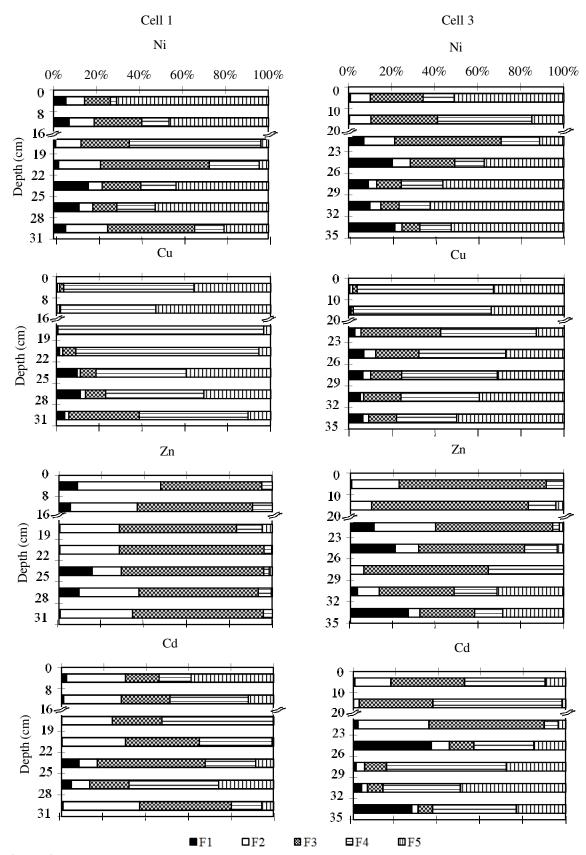
Figure 32 (continued). Vertical (depth) profiles of measured hydrosoil conditions and concentrations of Cd, Cu, Ni, and Zn. A. cell 1, B. cell 2, C. cell 3, and D. cell 4. Redox potential was plotted at the specific depths where measured. Seven depth intervals (2 detritus; 5 sediment) in each cell, represented by the shaded bar, were analyzed for pH, organic matter content, ∑SEM concentration, AVS concentration, SEM concentration of Cd, Cu, Ni, and Zn, and concentration of Cd, Cu, Ni, and Zn (measurement graphed as a point in the middle of each interval). Zero depth represents the surface water-detritus interface, and the line across the graphs represents the detritus-sediment interface.



**Figure 33.** Ni, Cu, Zn, and Cd fractionation in cells 1 and 3 of the pilot-scale CWTS hydrosoil expressed as concentration in mg/kg: F1= exchangeable; F2= bound to carbonates; F3= bound to Fe-Mn oxides; F4= bound to organics or sulfides; F5= bound to the residual fraction (Table 3). Fractionation of metals demonstrates changes in concentrations among fractions which a metal occurs in a cell planted with *S. californicus* (cell 1) and a cell planted with *T. angustifolia* (cell 3).



**Figure 34.** Fractionation of all metals (total Cd, Cu, Ni, and Zn) in cells 1 and 3 of the pilot-scale CWTS expressed as concentration in mg all metals/kg of hydrosoil: F1= exchangeable; F2= bound to carbonates; F3= bound to Fe-Mn oxides; F4= bound to organics or sulfides; F5= bound to the residual fraction (Table 3). Fractionation of all metals demonstrates changes in concentrations among fractions which all metals measured occur in a cell planted with *S. californicus* (cell 1) and a cell planted with *T. angustifolia* (cell 3).



**Figure 35.** Cd, Cu, Ni, and Zn fractionation in cells 1 and 3 of the pilot-scale CWTS expressed as percentages of the total concentrations with depth: F1= exchangeable; F2= bound to carbonates; F3= bound to Fe-Mn oxides; F4= bound to organics or sulfides; F5= bound to the residual fraction.

Table 58. Comparison of values for hydrosoil conditions favorable for biogeochemical processes and measured values of conditions in detritus and sediment

	Favorable Values		Measured Values		Summary
	Sorption	Complexation with AVS	Detritus <sup>c</sup>	Sediment <sup>c</sup>	
Redox Potential Range (mV)	-300 to 700 <sup>a,b</sup>	-300 to 0 <sup>a, b</sup>	-209 to -113	-249 to -34	Redox favorable for sorption and sulfide formation in hydrosoil
pH Range (S.U.)	2 to 10 <sup>a</sup> Greatest sorption at pH ~8, sorption decreases as pH decreases <sup>d</sup>	4 to 9.5 <sup>f</sup> Complexation with AVS favored in basic pH <sup>e</sup>	5.69 to 6.86	5.01 to 7.41	pH favorable for sorption and sulfide formation in hydrosoil
Organic Matter Content (OM; %)	Sorption increases with increasing organic content	> 0.1 % dry weight needed for dissimilatory sulfate reducing bacteria <sup>h</sup>	29.7 to 85.0	0.646 to 46.1	OM favorable for sorption and sulfide formation in hydrosoil
Vegetation	Sorption increases with increasing mass of roots and shoots <sup>i</sup>	Complexation with AVS favored by vegetation with small radial oxygen loss from root zone <sup>i</sup>	16 to 26 cm thick root mass comprising 35-46% of the hydrosoil	Few thick plant roots, extend downward as plant anchors	Vegetation favorable for sorption in detritus
AVS concentration (µmole/g)	N/A	Complexation with AVS increases with increasing AVS concentration	36-160 in cells 2 to 4	5.9-100 in cells 1 and 2	AVS concentration most favorable for sulfide formation in detritus of
			1.2-3.4 in cell 1	0.54-3.0 in cells 3 and 4	cells 2 to 4 and in sediment of cells 1 and 2

N/A Not applicable for treatment process or hydrosoil zone

a. Under Conditions of 25°C and 1 bar (Brookins, 1988)

b. Typical wetland Eh values (Reddy and DeLaune, 2008)

c. Inflow and outflow values from cells 1 and 2; Outflow values from cells 3 and 4

d. Abolinno et al. (2003); Msaky and Calvet (1990); Oyanedel-Craver and Smith (2006)

e. Patterson et al. (1977)

f. Barton and Tomei (1995); Postgate (1984) g. John and Leventhal (1995) h. Rickard (1973) i. Rodgers and Castle (2008)

#### Water Depth

Water depths in free water surface constructed wetland treatment systems (CWTSs) are usually established based on practicalities such as environmental requirements and tolerances of plants, height of water control or containment structures (berms), and periodicity and intensity of rainfall events. The availability of sufficient land at a specific site for the required "footprint" is often another important consideration regarding water depth in constructed wetlands for treatment of contaminated waters. In design of CWTSs, the "footprint" is based on the hydraulic retention time (HRT) necessary to achieve the required treatment performance (Kadlec and Knight, 1996; Rodgers and Castle, 2008). Water depth is a critical factor related to the flow volume, flow rate, and the HRT required for removal of targeted constituents (Kadlec and Knight, 1996). However, the influence of water depth on treatment performance in CWTSs is a relatively unstudied factor (Gillespie et al. 1998; Tanner et al. 1998; Headley et al. 2005; Aguirre et al. 2005).

Some factors impacting performance of free water surface CWTSs that can be influenced by water depth include: 1) net oxygen supply rate (NOSR; Kadlec and Knight, 1996), 2) sediment oxidation-reduction potential (redox; Gambrell et al. 1991), 3) removal pathways and processes for targeted constituents that occur in CWTSs (e.g. metal precipitation and biodegradation; Rodgers and Castle, 2008), 4) plant survival and growth (Brix and Sorrell, 1996; Sorrell et al. 2002), 5) mass loading rates (Tao et al. 2005; Caselles-Osorio and García, 2006), and 6) organic matter content in the sediment (Aguirre et al. 2005). Clearly, these factors can contribute independently and collectively to the treatment performance achieved by a CWTS (Allen et al. 1993; Brix, 1994; Obire and Nwaubeta, 2001; Kadlec and Knight, 1996).

The purpose of this research was to evaluate the influence of a series of water depths on factors affecting treatment performance of pilot-scale CWTSs for renovating simulated FOPW. Specific objectives of this study were to: 1) prepare simulated FOPW, 2) design and construct pilot-scale CWTSs, 3) compare treatment performance data (i.e. rates and extents of removal of Cd, Cu, Ni, Zn, and oil) for the pilot-scale CWTSs with a 36 h HRT at five incremental water depths (i.e. 15, 23, 33, 46, and 56 cm), and 4) measure sediment redox potential and sediment organic matter content in pilot-scale CWTS wetland cells planted with *Schoenoplectus californicus* and wetland cells containing *Typha angustifolia*.

Simulated FOPW was formulated based on constituent concentrations in FOPWs reported in peer-reviewed literature (Veil et al. 2004; Thordsen et al. 2007; Alley et al. 2011) and government documents (USEPA, 2000; USGS, 2002) as well as analyses of PW samples from oilfield sites in the continental United States. An ionic balance was calculated (i.e.  $\Sigma$  cations (meq/L) +  $\Sigma$  anions (meq/L) from analyte concentrations in actual FOPW to formulate simulated FOPW with appropriate salts to accurately achieve ionic composition and strength as well as salinity and hardness. The oil component for the simulated FOPW formulation was conventional light oil (Rotella® T 15W40), a cost-effective and reproducible alternative to using an actual crude oil. General water characteristics (i.e. pH, dissolved oxygen, conductivity, alkalinity, and hardness) were also measured to compare simulated and actual FOPW. Simulated FOPW was composed of Cd, Cu, Ni, Zn, oil, and

salts well-mixed with 3780 L of municipal water in a reservoir (USEPA, 2000; USGS, 2002; Veil et al. 2004; Thordsen et al. 2007; Alley et al. 2011; Table 1). This formulation of FOPW resulted in the following concentrations of constituents targeted for treatment: 6 mg Cd<sup>+2</sup>/L, 4 mg Cu<sup>+2</sup>/L, 6 mg Ni<sup>+2</sup>/L, 6 mg Zn<sup>+2</sup>/L, and 100 mg Rotella® T 15W40/L.

The pilot-scale wetlands were designed and constructed with emphasis on water depth as a treatment variable. Design of the pilot-scale systems was premised on the calculated rate of oxygen diffusion through the water column to the sediment of a wetland cell. The film theory of mass transfer and the NOSR equation from Kadlec and Knight (1996) were used to estimate the rate of oxygen supply to the sediment at different water depths to predict sediment redox conditions that could be achieved. Oxygen diffusivity, defined as the ability of oxygen to diffuse through water at a specific depth, was estimated based on the film theory of mass transfer equation:

$$K = \begin{bmatrix} D \\ d \end{bmatrix} \tag{6}$$

where, K is the reaeration rate, m/d, D is the molecular diffusivity of oxygen in water, m<sup>2</sup>/d, and d is the water depth, m. Transfer of oxygen from the atmosphere to the sediment in the wetland can be estimated using the NOSR equation (Kadlec and Knight, 1996):

$$NOSR = K(C^{sat}_{DO} - C_{DO}) - SOD$$
 (7)

where NOSR is the net oxygen supply rate,  $g/m^2*d$ , K is the reaeration coefficient, m/d (estimated from the film theory of mass transfer equation),  $C_{DO}^{sat}$  is the saturated dissolved oxygen concentration at 1 atm, 8.7 mg  $O_2/L$  ( $g/m^3$ ; APHA, 2005; Huddleston et al. 2008),  $C_{DO}$  is the average dissolved oxygen concentration in the bulk of water, 8.0 mg  $O_2/L$  ( $g/m^3$ ; Huddleston et al. 2008), and SOD is sediment oxygen demand,  $0.1 \pm 0.01$  mg  $O_2/day$  (Huddleston et al. 2008). We assumed that the SOD value from Huddleston et al. (2008) will estimate the SOD for these wetland cells. Huddleston et al. (2008) measured SOD using the same sediment and comparable formulation and loadings of Osmocote® as in this experiment with a water depth of 30 cm. A general equation was formulated relating water depth to HRT for these specific pilot-scale CWTSs:

$$d = \frac{HRT * Q}{\pi r^2}$$
 (8)

where, d is the water depth, cm, HRT is the hydraulic retention time, min, Q is flow rate, ml/min, and,  $\pi r^2$  is the surface area of the wetland, cm<sup>2</sup>. The following equation determines the mass loading rates for each wetland cell and series:

$$ML = C * R \tag{9}$$

where, ML is the mass loading rate, mg/min, C is the concentration of constituents in the inflow water, mg/ml, and R is the flow rate, min/ml.

For experimental purposes, pilot-scale CWTSs were used due to their sufficiently small size to facilitate control and manipulation of macrofeatures (e.g. water depth, hydraulic retention time, vegetation, and hydrosoil; Dorman et al. 2009; Rodgers and Castle, 2008). For this experiment, wetland cells were constructed in 150 L polypropylene barrels that were 34.5 cm in diameter and 75.5 cm deep. The wetland cells were connected by polyvinyl chloride (PVC) pipe fittings and arranged to permit flow of water to each cell in series by gravity. The PVC pipe fittings were positioned at 30 cm from the bottom of the polypropylene barrel approximately 15 cm above the sediment with a standpipe to adjust the water depth internally to allow for changes in the wetland cells during the experiment. Six replicated series were constructed for this pilot-scale experiment with four wetland cells in each series.

For this study, experimental water depths were determined from the film theory of mass transfer and NOSR equations. Values calculated from the film theory of mass transfer equation generated a linear model (y = -0.0001x + 0.0022) containing reaeration rates and the corresponding depths from 0 to 60 cm. Five values related to specific water depths and reaeration rates were used in the NOSR equation (y = -0.0001x - 0.0491) to predict the supply of oxygen to the sediment. Theoretically, these five water depths would produce oxidizing (i.e. 15, 23, 33 cm) or reducing (i.e. 46 and 56 cm) sediment conditions in the wetland cells. Five water depths were grouped according to predicted sediment redox conditions [1)15 and 23 cm water depths, 2) 33 cm water depth, and 3) 46 and 56 cm water depths]. For experimental purposes, two wetland series were used for each water depth group. The standpipes were positioned according to a specific water depth for each series. For the initial experiment, standpipes were positioned for the 15, 33, and 46 cm water depths in the wetland series. For the subsequent experiment, standpipes were changed in length to produce the 23, 33, and 56 cm water depths. During this study, the 33 cm water depth series were not altered to establish commonality between the two experiments. At the initiation of the experiment and between changes in water depths, pilot-scale systems were allowed to acclimate. An acclimation period is the time needed to obtain stability of a system as reflected by sediment redox potential, pH, and dissolved oxygen concentration (OECD, 2002), and the period of acclimation in this case was approximately two to three weeks. Sediment for all wetland cells was from Eighteen Mile Creek (South Carolina) and composed of ~85% sand, ~15% silt, and clay-size particles. Vegetation in the first two wetland cells of each series was giant bulrush (S. californicus). Narrowleaf cattail (T. angustifolia) was planted in the final two wetland cells of each series. Initial density of these plants was 8-10 plants/m<sup>2</sup> in each wetland cell. Approximately three grams of Osmocote® Outdoor & Indoor Smart-Release® Plant Food (19-6-12 formula) were added to each wetland cell every two weeks for the duration of the experiment.

For complex mixtures such as FOPWs, a hybrid CWTS may be necessary to achieve the required performance to meet specific water use criteria. For this experiment, the hybrid system was comprised of an energy-driven component (i.e. oil/water separator) followed by a specifically designed pilot-scale CWTS. For FOPWs containing sufficient oil to merit separation, an oil/water separator may promote additional performance and opportunity to recover cost of operation and maintenance. For this pilot-scale free water surface CWTS,

simulated FOPW was mixed with a ¾ horsepower sump pump in the simulated FOPW reservoir. From the reservoir to the oil/water separator, FMI® piston pumps were calibrated to maintain a 6 h HRT to allow for sufficient flow to achieve the targeted HRT in the pilot-scale systems. After exiting the oil/water separator, simulated FOPW flowed to each series using FMI® pumps. The equation relating water depth to HRT (Eq. 8) was used to calculate flow rates for each series depending on the corresponding water depth. FMI® pumps were calibrated for the pilot-scale systems to the corresponding flow rates to maintain a 36 h HRT for each water depth in the wetland cells.

Aqueous samples were collected from three locations: simulated FOPW reservoir, oil/water separator outflow, and each wetland cell outflow in the pilot-scale CWTSs. Cd, Cu, Ni, Zn, and oil were measured to determine if treatment performance was achieved for surface water discharge and irrigation in the pilot-scale CWTSs (Table 2). Aqueous Cd, Cu, Ni, and Zn samples (50 mL) were collected bi-weekly in Nalgene bottles and prepared for analysis within 48 hours of collection (APHA, 2005). Aqueous samples for oil analysis were collected from each wetland cell monthly using 1 liter glass jars using the APHA (2005) method (5520 A) and a sheen net at the water's surface according to the ASTM (2011) method (D4489-95) and prepared for analysis within four hours of collection (APHA, 2005). Cd, Cu, Ni, Zn, and oil samples were stored at 4°C until analysis at Clemson University (APHA, 2005). Soluble Cd, Cu, Ni, and Zn concentrations were prepared by filtering aqueous samples through a 0.45-µm filter, acidifying to pH < 2 with trace metal grade nitric acid (Thermo Fisher Scientific, Inc.), and analyzing using Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES; Perkin Elmer; USEPA, 1994; APHA, 2005). According to USEPA (1993), soluble metal concentrations may be measured to determine bioavailable constituents in contaminated waters (Allen and Hansen, 1996). Oil samples were prepared by liquid-liquid extraction (Korda et al. 1997). The sample flowed (10 ml/min) into a silica column (Supelclean<sup>TM</sup> ENVI<sup>TM</sup>- 18 SPE cartridge; Li and Lee, 2001). Flow rate was controlled by adjusting the vacuum suction pressure. Oil compounds were eluted from the column using 10 ml chloroform [moderately insoluble (MI)], then 10 ml *n*-hexane [water insoluble (WI)], and 10 ml methanol [water soluble (WS); Siron et al. 1987]. WI, MI, and WS eluates were dried under a nitrogen current to a volume of 2 ml to inject in vials for analysis (Siron et al. 1987). Dried WI, MI, and WS eluates were analyzed initially for spectral identification of compounds for the use of calibration standards to estimate concentrations of those compounds (MDEP, 2009). WI and MI compounds (1,2-Benzofluorenone and 1-Methylcyclopentanol, respectively) were analyzed using gas chromatography (GC; Varian 8510) with a flame ionization detector using an ZB-1 capillary column (50m length, i.d. 0.3mm; Siron et al. 1987; Korda et al. 1997). Temperature was programmed for the GC from 170 to 290°C at 3°C min<sup>-1</sup> with a retention time of 45 minutes and the carrier gas was helium (Siron et al. 1987). The WS compound (Diethylhydroxypentanol) was analyzed using high pressure liquid chromatography (HPLC; Dionex UltiMate 3000; Siron et al. 1987) with an Acclaim Polar Advantage II column (4.6x250mm, 5µm, 120Å). Temperature was programmed on the HPLC from 30-45°C at 5°C min<sup>-1</sup> with a retention time of 30 minutes (Siron et al. 1987). The following gradient elution was applied—solvent A: acetonitrile; solvent B: water at time 0: A = 40%, B = 60%; time 5 min: A = 40%, B = 60%; time 40 min: A = 100%, B = 60%; time 40 min: A = 100%, B = 60%; time 40 min: A = 100%, B = 60%; time 40 min: A = 100%, B = 100%0% (Moret et al. 1997).

Water pH, temperature, conductivity, dissolved oxygen, alkalinity, and hardness were measured bi-weekly from each wetland cell (APHA, 2005). Temperature and water pH were measured with an Orion model GDT-11 field probe, and dissolved oxygen and conductivity were measured using a YSI meter model 85 from the inflow and each wetland cell (APHA, 2005). Sediment redox potential was measured bi-weekly in each wetland cell. Sediment redox potential was measured with a platinum electrode and an Ag/AgCl reference system (accuracy: ±10 mV) at 2.5 cm depth in the middle of each wetland cell (Faulkner et al. 1989).

Sediment redox potential and organic matter content are parameters that may be influenced by mass loading rates and plant detritus decomposition which can potentially affect treatment performance of the system. Wetland cells were compared within and between each water depth group to discern differences in the sediment redox potential and organic matter content. Sediment redox potential was measured as described previously, and organic matter content in the sediment was measured at the initiation and termination of this experiment from each wetland cell. Organic matter content was measured from the top 2.5 cm of sediment using loss-on-ignition method (USEPA, 2002d).

Data collected for actual and simulated FOPW for Cd, Cu, Ni, Zn, oil, and salts were analyzed for normal distribution. Significant differences in the Cd, Cu, Ni, Zn, oil, and salt concentrations for the actual and simulated FOPWs were determined by ANOVA and Tukey's procedure ( $\alpha = 0.05$ ). Treatment performance data were analyzed for normal distribution. Significant differences in mean outflow concentrations of the five water depth wetland cells of the pilot-scale systems were determined by ANOVA and Tukey's procedure ( $\alpha = 0.05$ ). Sediment redox potential and organic matter content for the different water depths and wetland cells containing *S. californicus* and *T. angustifolia* were analyzed for normal distribution. Significant differences in sediment redox potential and organic matter content for the different water depths and wetland cells containing *S. californicus* and *T. angustifolia* were determined by ANOVA and Tukey's procedure ( $\alpha = 0.05$ ). Statistical analyses were conducted using Statistical Analytical System (SAS) 9.2 (SAS Institute, 2002).

Statistical comparisons using ANOVA and Tukey's procedure were conducted on the different water depths for treatment performance. There is a significant difference ( $\alpha$ =0.05) between the water depths for concentrations of Cd, Cu, Ni, Zn, and oil fractions. For the 15, 23, and 33 cm water depths, the Cd, Ni, and oil fraction concentrations were not significantly different. For Zn concentrations, the 15 and 23 cm and the 23 and 33 cm water depths were not significantly different from each other. For Cu concentrations in the wetlands, the 15 cm water depth was significantly different from the other depths, while the 23 and 33 cm water depths and the 33, 46, and 56 cm water depths were not significantly different from each other. Cd, Cu, Ni, Zn, and oil concentrations were not significantly different ( $\alpha$ =0.05) for the 46 and 56 cm water depths. The following Cd, Cu, Ni, Zn, and oil outflow concentrations are presented as two different categories: wetland cells with oxidizing conditions in the sediment (i.e. 15, 23, and 33 cm) and wetland cells with reducing conditions in the sediment (i.e. 46 and 56 cm).

Average inflow concentrations ( $\pm$  standard deviation) of the simulated FOPW were 3.99 ( $\pm$ 1.23) mg Cd/L, 3.31 ( $\pm$ 2.49) mg Cu/L, 3.74 ( $\pm$ 0.82) mg Ni/L, and 3.77 ( $\pm$ 1.04) mg Zn/L. The average inflow oil concentration and after the oil/water separator were 43.73 ( $\pm$ 1.59) mg/L and 14.15 ( $\pm$ 1.09) mg/L (Table 59).

The average soluble concentrations exiting the 46 cm water depth constructed wetland were  $0.16~(\pm 0.15)~\text{mg}$  Cd/L,  $0.04~(\pm 0.03)~\text{mg}$  Cu/L,  $0.07~(\pm 0.03)~\text{mg}$  Ni/L, and  $0.15~(\pm 0.11)~\text{mg}$  Zn/L (Figures 35-38; sediment redox potential (SRP): -65 to -168.9 mV). At a water depth of 56 cm, the average soluble concentrations exiting the constructed wetland were  $0.19~(\pm 0.27)~\text{mg}$  Cd/L,  $0.06~(\pm 0.08)~\text{mg}$  Cu/L,  $0.09~(\pm 0.06)~\text{mg}$  Ni/L, and  $0.18~(\pm 0.22)~\text{mg}$  Zn/L (SRP: -132.8 to -212 mV; Figures 36-39). The average Cd, Cu, and Ni concentrations had a slight increase as the water depth increased from 46 to 56 cm. Significant differences ( $\alpha$ =0.05) were observed for the Cd, Cu, Ni, and Zn concentrations in the inflow water and 46 and 56 cm water depth wetland cell outflows. Removal of Cd, Cu, Ni, and Zn in the wetland cells may be attributed to the reducing environment conducive for metal precipitation by sulfides through dissimilatory sulfate reduction.

The average oil concentrations from the initial to the final cell for the 46 cm water depth wetland cells were 13.89 (±0.59) mg/L, 13.73 (±0.61) mg/L, 13.59 (±0.52) mg/L, and 13.48 (±0.49) mg/L. The average oil concentrations after each 56 cm water depth wetland cell were 13.67 (±0.85) mg/L, 13.49 (±0.76) mg/L, 13.39 (±0.71) mg/L, and 13.35 (±0.69) mg/L. We expected more removal in the oxidizing wetland cells (i.e. 15, 23, and 33 cm water depth wetland cells) than in the reducing wetland cells (i.e. 46 and 56 cm water depth wetland cells) which had a slower degradation rate (Figure 40; Table 59).

The average soluble concentrations exiting the 15 cm water depth constructed wetland were 3.38 (±0.99) mg Cd/L, 1.78 (±0.81) mg Cu/L, 3.54 (±0.47) mg Ni/L, and 3.55 (±0.57) mg Zn/L (SRP: 162 to 250.1 mV). The average soluble concentrations exiting the 23 cm water depth constructed wetland were 3.94 (±1.25) mg Cd/L, 2.23 (±2.23) mg Cu/L, 3.02 (±1.31) mg Ni/L, and 2.74 (±0.84) mg Zn/L (SRP: 115.8 to 249.5mV). The average soluble concentrations exiting the 33 cm water depth constructed wetland were 3.01 (±0.97) mg Cd/L, 1.29 (±1.78) mg Cu/L, 2.74 (±0.83) mg Ni/L, and 2.44 (±0.73) mg Zn/L (SRP: 15.3 to 138.4 mV). We observed decreases in the divalent metal concentrations as the depth increased from 15 to 33 cm (Figures 36-39; Table 59). Divalent metal outflow concentrations were greater in wetland cells having 15 and 23 cm water depths than in cells having 46 and 56 cm water depths. The 46 and 56 cm water depth wetland cells provided reducing conditions conducive for dissimilatory sulfate reduction. Precipitation of divalent metals by acid-volatile sulfide binding requires reducing conditions in the sediment in order to provide the appropriate environment for dissimilatory sulfate reducing microbial assemblages.

The average oil concentrations from the initial to the final cell for the 15 cm water depth wetland cells were  $10.26~(\pm~0.16)~\text{mg/L}$ ,  $7.52~(\pm~0.35)~\text{mg/L}$ ,  $3.12~(\pm~0.66)~\text{mg/L}$ , and  $0.96~(\pm~0.07)~\text{mg/L}$  (SRP: 162~to~250.1~mV). The average oil concentrations for the 23 cm water depth wetland cells were  $10.29~(\pm~0.51)~\text{mg/L}$ ,  $6.92~(\pm~1.65)~\text{mg/L}$ ,  $3.07~(\pm~0.47)~\text{mg/L}$ , and

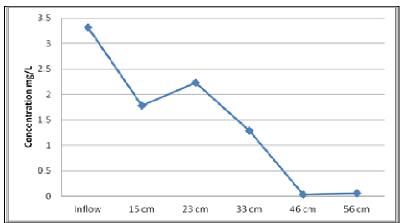
1.08 (±0.14) mg/L. The average oil concentrations for the 33 cm water depth wetland cells were 10.92 (±0.59) mg/L, 8.33 (±0.27) mg/L, 5.31 (±0.30) mg/L, and 2.52 (±0.09) mg/L. We observed a greater decrease in oil concentrations in the 15 and 23 cm water depths wetland cells than in the 33 cm water depth wetland cell. This may be attributed to a lower redox potential in the sediment (i.e. less oxygen to the sediment) of the 33 cm than the 15 and 23 cm water depth wetland cells. As water depth increases, the concentration of oil removed decreases (Figure 40; Table 59). We observed lower oil concentrations in the 15 and 23 cm water depths than in the 46 and 56 cm water depths wetland cells (Figure 40). It is generally concluded that oil degrades more quickly under aerobic conditions than under anaerobic conditions (Zobell, 1964; Hughes and McKenzie, 1975; Ward and Brock, 1978; Delaune et al. 1980).

Water depth influences sediment redox in a wetland, which may alter treatment performance of a system. The 15 and 23 cm water depth wetland cells were not efficient for removing Cd, Cu, Ni, and Zn concentrations from the simulated FOPW. However, the 15 and 23 cm water depths wetland cells were more effective for removing oil components than the 46 and 56 cm water depth wetland cells since aerobic degradation occurs at a faster rate than anaerobic degradation. The 46 and 56 cm water depth wetland cells were more effective at removing Cd, Cu, Ni, and Zn from simulated FOPW than the 15, 23, and 33 cm water depth wetland cells. The 33 cm water depth redox potential (i.e. 15.3 to 138.4 mV) was inefficient for providing conditions conducive for divalent metal precipitation and degradation of oil and, as a result, was ineffective at removing these constituents from simulated FOPW. Differences in the water depth used for Phase II of this project was 40.6 cm with a HRT of 24 h, while the water depth experiment wetland cells contained water depths from 15 to 56 cm in depth and a HRT of 36 h. In comparison, the wetland cells having a 40.6 cm water depth removed more divalent metals than the 46 and 56 cm wetland cells. This may be attributed to the wetland cell size and maturity of the system.

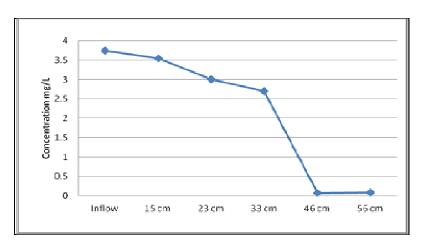
Removal rate coefficients were calculated using Equation 9 for the 5 water depths investigated. For the 15, 23, and 33 cm water depths, oxidizing conditions (+15.3 to +250.1 mV) were established along with greater removal rates for oil (0.01-0.056 day-1) compared with the 46 and 56 cm water depths (~0.001 day-1). Reducing environments (-65 to -212 mV) were generated in 46 and 56 cm water depths and supported greater removal rates for metals (0.01-0.185 day-1) than the 15, 23, 33, cm water depths (no change-0.077 day-1), which may be attributed to precipitation by sulfides from dissimilatory sulfate reduction.



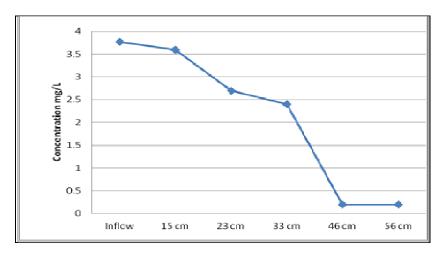
**Figure 36.** Average cadmium concentrations in the inflow and outflow for each water depth in the constructed wetland.



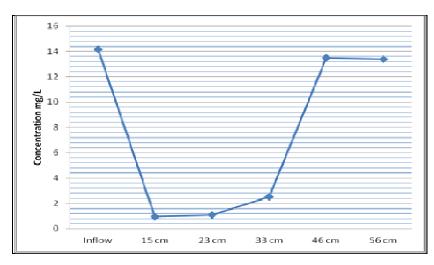
**Figure 37.** Average copper concentrations in the inflow and outflow for each water depth in the constructed wetland.



**Figure 38.** Average nickel concentrations in the inflow and outflow for each water depth in the constructed wetland.



**Figure 39.** Average zinc concentrations in the inflow and outflow for each water depth in the constructed wetland.



**Figure 40.** Average oil concentrations in the inflow and outflow for each water depth in the constructed wetland.

**Table 59.** Average constituent concentrations in the simulated FOPW at various water depths.

simulated 1 of w at various water depths.						
	Oil	Cd	Cu	Ni	Zn	
Inflow	14.15	3.99	3.31	3.74	3.77	
15 cm	0.96	3.38	1.78	3.54	3.55	
23 cm	1.08	3.94	2.23	3.02	2.74	
33 cm	2.52	3.01	1.29	2.74	2.44	
46 cm	13.48	0.16	0.04	0.07	0.15	
56 cm	13.35	0.19	0.06	0.09	0.18	

### Seasonal Variation

Among aquatic systems, constructed wetlands display the greatest seasonal variability of physical and chemical characteristics (e.g. macrophytes, sedimentation rates, flow volume, type and concentration of constituents) in response to terrestrial and climatic events (Kalff, 2002; Rai, 2008). Seasonal variability may have direct or indirect effects on biological processes that may be temperature dependent (Abdollahi and Nedwell, 1979; Halda-Alija and Johnston, 1999). Biological processes targeted for constituent removal in pilot-scale CWTSs include biodegradation of oil by heterotrophic aerobic microbial assemblages and precipitation of Cd, Cu, Ni, and Zn by sulfides through dissimilatory sulfate reduction.

Responses of microbial populations to changes in temperature are crucial knowledge for successful design and implementation of a full-scale CWTS. Investigations were conducted concerning the effect of seasonal variability on performance of a hybrid pilot-scale CWTS for the removal of Cd, Cu, Ni, Zn, and oil. For this experiment, the hybrid system was comprised of an energy-driven component (i.e. oil/water separator) followed

by a specifically designed pilot-scale CWTS (Figure 41). For FOPWs containing sufficient oil to merit separation, an oil/water separator may promote additional performance and opportunity to recover cost of operation and maintenance. The objective of this research was to measure the seasonal performance of a hybrid pilot-scale constructed wetland system for treatment of Cd, Cu, Ni, Zn, and oil in simulated FOPW.

Simulated FOPW was formulated based on constituent concentrations in FOPWs reported in peer-reviewed literature (Veil et al., 2004; Thordsen et al., 2007; Alley et al., 2011) and government documents (USEPA, 2000; USGS, 2002) as well as analyses of PW samples provided from oilfield sites in the continental United States. For simulated FOPW, an ionic balance was calculated (i.e.  $\Sigma$  cations (meg/L) +  $\Sigma$  anions (meg/L) from analyte concentrations in actual FOPW to formulate this water with appropriate salts to accurately achieve ionic composition and strength as well as salinity and hardness. The oil component for the simulated FOPW formulation was conventional light oil (Rotella® T 15W40), a cost-effective and reproducible alternative to using an actual crude oil. General water characteristics (i.e. pH, dissolved oxygen, conductivity, alkalinity, and hardness) were also measured to compare simulated FOPW and actual FOPW. Simulated FOPW was composed of Cd, Cu, Ni, Zn, oil, and salts well-mixed with 3780 L of municipal water in a reservoir (USEPA, 2000; USGS, 2002; Veil et al., 2004; Thordsen et al., 2007; Alley et al., 2011). This formulation of FOPW resulted in the following concentrations of targeted constituents required for treatment: 6 mg Cd<sup>+2</sup>/L, 4 mg Cu<sup>+2</sup>/L, 6 mg Ni<sup>+2</sup>/L, 6 mg Zn<sup>+2</sup>/L, and 100 mg Rotella® T 15W40/L.

For this pilot-scale free water surface CWTS, simulated FOPW was mixed with a <sup>3</sup>/<sub>4</sub> horsepower sump pump in the simulated FOPW reservoir. From the reservoir to the oil/water separator, FMI® piston pumps were calibrated to maintain a 6 h HRT to allow for sufficient flow to achieve the targeted HRT in the pilot-scale systems. The nominal HRT of the pilot-scale CWTS was 24 hours per cell or 96 hours for the series.

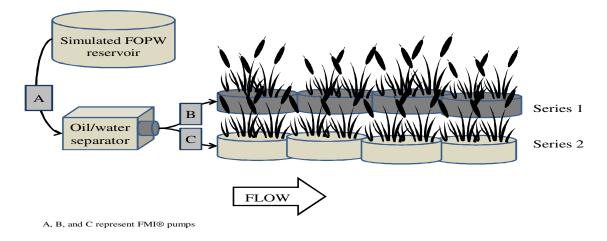


Figure 41. Schematic of the hybrid pilot-scale constructed wetland treatment systems

Simulated FOPW samples were collected and analyzed at Clemson University from May 2010 through May 2011. Aqueous samples were collected from three locations: simulated FOPW reservoir, oil/water separator outflow, and each wetland cell outflow in the pilotscale CWTSs. Cd, Cu, Ni, Zn, and oil were targeted constituents measured to determine whether treatment performance was achieved for surface water discharge and irrigation in the pilot-scale CWTSs (Table 60). Aqueous Cd, Cu, Ni, and Zn samples (50 mL) were collected bi-weekly in Nalgene bottles and prepared for analysis within 48 hours of collection (APHA, 2005). Aqueous samples for oil analysis were collected from each wetland cell monthly using 1 liter glass jars using the APHA (2005) method (5520 A) and a sheen net at the water's surface according to the ASTM (2011) method (D4489-95) and prepared for analysis within four hours of collection (APHA, 2005). These samples were stored at 4°C until analysis at Clemson University (APHA, 2005). Acid-soluble Cd, Cu, Ni, and Zn concentrations were prepared by acidifying to pH < 2 with trace metal grade nitric acid, filtering aqueous samples through a 0.45-um filter (Thermo Fisher Scientific, Inc.), and analyzing using Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES; Perkin Elmer; USEPA, 1994b; APHA, 2005). Oil samples were prepared by liquid-liquid extraction (Korda et al. 1997). The sample flowed (10 ml/min) into a silica column (Supelclean<sup>TM</sup> ENVI<sup>TM</sup>- 18 SPE cartridge; Li and Lee, 2001). The flow rate was controlled by adjusting the vacuum suction pressure. Oil compounds were eluted from the column using 10 ml chloroform [moderately insoluble (MI)], then 10 ml n-hexane [water insoluble (WI)], and 10 ml methanol [water soluble (WS); Siron et al. 1987]. The WI, MI, and WS eluates were dried under a nitrogen current to a volume of 2 ml to inject in vials for analysis (Siron et al. 1987). Dried WI, MI, and WS eluates were analyzed initially for spectral identification of compounds for the use of calibration standards to estimate concentrations of those compounds (MDEP, 2009). WI and MI compounds (1,2-Benzofluorenone and 1-Methylcyclopentanol, respectively) were analyzed using gas chromatography (GC; Varian 8510) with a flame ionization detector using an ZB-1 capillary column (50m length, i.d. 0.3mm; Siron et al. 1987; Korda et al. 1997). Temperature was programmed for the GC from 170 to 290°C at 3°C min<sup>-1</sup> with a retention time of 45 minutes and the carrier gas was helium (Siron et al. 1987). The WS compound (Diethylhydroxypentanol) was analyzed using high pressure liquid chromatography (HPLC; Dionex UltiMate 3000; Siron et al. 1987) with an Acclaim Polar Advantage II column (4.6x250mm, 5µm, 120Å). Temperature was programmed on the HPLC from 30-45°C at 5°C min<sup>-1</sup> with a retention time of 30 minutes (Siron et al. 1987). The following gradient elution was applied—solvent A: acetonitrile; solvent B: water at time 0: A = 40%, B = 60%; time 5 min: A = 40%, B = 60%; time 40 min: A = 100%, B = 0% (Moret et al. 1997).

Water pH, temperature, conductivity, dissolved oxygen, alkalinity, and hardness were measured and recorded bi-weekly from each wetland cell (APHA, 2005). Temperature and water pH were measured with an Orion model GDT-11 field probe, and dissolved oxygen and conductivity was measured using a YSI meter model 85 from the inflow and each wetland cell (APHA, 2005). Sediment redox potential was measured and recorded bi-weekly in each wetland cell. Sediment redox potential was measured with a platinum electrode and an Ag/AgCl reference system (accuracy: ±10 mV) at a 2.5 cm depth in the

near middle of each wetland cell (Faulkner et al. 1989). Performance data were analyzed for normal distribution. Significant differences in mean outflow concentrations between the two wetland cell series were determined by ANOVA and Tukey's procedure ( $\alpha = 0.05$ ).

There were no observed differences ( $\alpha$ =0.05) in the performance of the two wetland series for oil, Cd, Cu, Ni, and Zn. Average (±s.d.) oil concentration exiting Series 1 was 1.32 (±0.28) mg oil compounds/L. Average (±s.d.) oil concentration exiting Series 2 was 1.3 (±0.29) mg oil compounds/L. Both wetland series remediated approximately 82.0-91.4% of the oil compounds measured in the simulated FOPW samples. Approximately 98.3-99.9% of the divalent metal concentrations in the simulated FOPW were renovated by the pilot-scale system. Average (±s.d.) Cd, Cu, Ni, and Zn concentrations exiting Series 1 were  $0.003 (\pm 0.002)$  mg Cd/L,  $0.004 (\pm 0.002)$  mg Cu/L,  $0.063 (\pm 0.028)$  mg Ni/L, and 0.051(±0.041) mg Zn/L (Figure 42). For Series 1, Cu concentrations measured in July, September, and October were non-detect and were recorded for calculation purposes as the detection limit (0.003). Average (±s.d.) Cd, Cu, Ni, and Zn concentrations exiting Series 2 were  $0.003 (\pm 0.002)$  mg Cd/L,  $0.004 (\pm 0.002)$  mg Cu/L,  $0.053 (\pm 0.034)$  mg Ni/L, and 0.052 (±0.038) mg Zn/L (Figure 43). Differences in removal efficiencies for Cd, Cu, Ni, and Zn may be attributed to the Ksp values for sulfide complexation (i.e. Ksp: Cu 10<sup>-37</sup>, Cu 10<sup>-32</sup>, and ~Ni and Zn 10<sup>-25</sup>) or preferential uptake by S. californicus or T. angustifolia (Kamal et al. 2004). The temperature ranged from 2.7 to 30.2°C, and average temperatures for each month are illustrated in Figure 44. No observed significant differences ( $\alpha$ =0.05) in performance of either series were affected by changes in temperature in the pilot-scale system. Performance data for wetland series 1 and 2 were compared to FAO irrigation guidelines and USEPA water quality criteria (Table 60). During the experiment, Cd, Cu, Ni, Zn, and oil outflow concentrations met FAO irrigation guidelines, however, Cd and oil compound concentrations did not continuously met the USEPA water quality criteria. Although chemical-specific data met the irrigation and surface water discharge criteria, toxicity data were needed to determine if any synergist or additive effects occurred between the constituents in the simulated FOPW.

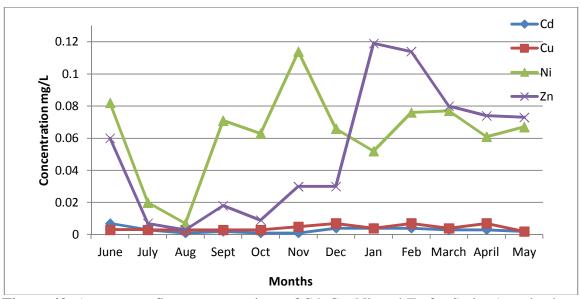


Figure 42. Average outflow concentrations of Cd, Cu, Ni, and Zn for Series 1 wetland

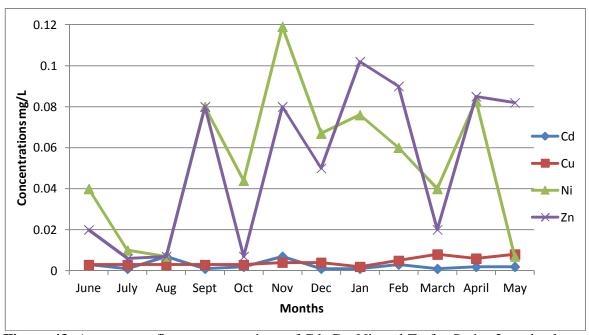


Figure 43. Average outflow concentrations of Cd, Cu, Ni, and Zn for Series 2 wetland

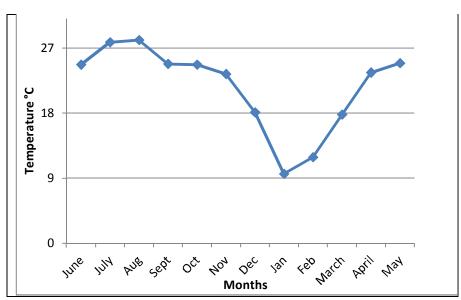


Figure 44. Average temperatures for each month during the pilot-scale CWTS experiment

**Table 60.** Comparisons between FAO Irrigation Guidelines and USEPA Water Quality Criteria and Outflow Concentrations (mg/L) from Constructed Wetland Series 1 and 2.

	Cadmium	Copper	Nickel	Zinc	Oil
Inflow	3.99	3.31	3.74	3.77	42.78
Series 1	0.003	0.004	0.063	0.051	1.32
Series 2	0.003	0.004	0.053	0.052	1.3
Irrigation Guidelines	0.01	0.2	0.2	2	35
Surface Discharge	0.002	0.009	0.47	0.12	0.1

Toxicity experiments using fish and invertebrate species and crop seedlings (i.e. corn, cabbage, and soybean) were conducted to discern if this simulated FOPW was suitable for use (i.e. surface water discharge and irrigation) after treatment.

Toxicity experiments are important for determining if a water may cause adverse effects to receiving system biota. Responses (i.e. mortality) of *Ceriodaphnia dubia* Richard and *Pimephales promelas* Rafinesque can be used to evaluate the toxicity of treated and untreated simulated FOPW. *C. dubia* and *P. promelas* are sentinel species used as indicators for synergistic, additive, and antagonistic effects of constituents that may occur in simulated FOPW. Untreated and treated simulated FOPWs were collected from series inflow, after the oil/water separator, and after each wetland cell. USEPA (1994b) method 600/4-91/002 for estimating toxicity of receiving waters was modified by changes in the composition of moderately hard water used in the method. *C. dubia* and *P. promelas* mortality was measured in 7-day static/renewal toxicity experiments (n=20 *P. promelas* per treatment and n=10 *C. dubia* per treatment: USEPA 2002c). Statistical analysis of the responses of *C. dubia* and *P. promelas* was conducted using SAS 9.2 (SAS Institute 2002) to determine if differences existed between a laboratory control and untreated and treated

simulated fresh PW. Differences were determined by performing Chi-Square Analysis ( $\alpha = 0.05$ ; SAS Institute 2002). The null hypothesis, untreated simulated fresh PW equivalent in toxicity to treated simulated fresh PW, was rejected by the Chi-square analysis (p <0.0001). There is a significant difference between the responses (i.e. mortality) of *C. dubia* and *P. promelas* to untreated and treated simulated fresh PW. All of the *C. dubia* and *P. promelas* died in the untreated simulated fresh PW within 24 hours. After 7 days, nine *C. dubia* and eight *P. promelas* survived in water treated by wetland series 1, and eight *C. dubia* and eight *P. promelas* survived in water treated by wetland series 2.

One-way ANOVA and Post-hoc Tukey's procedure ( $\alpha$ =0.05) were used to evaluate early seedling growth of corn (Zea maya), cabbage (Brassica oleracea), and soybean (Phaseolus vulgaris) seeds with control (municipal) water (n=30 for each seed group), untreated (n=30 for each seed group) and treated simulated FOPW (n=30 for each seedling group). For early seedling growth, differences in the root and shoot length between the seeds watered with treated and untreated simulated FOPW were significant ( $\alpha$ =0.05). The control and treated seedlings were not significantly different ( $\alpha$ =0.05) in root and shoot length. All of the seedlings in the control and treated groups were green and grew twice as fast as the untreated groups. For example, after Day 3 the average soybean height for the control and treated groups was approximately 5.8 cm, while the average soy bean height for the untreated group was approximately 3.9 cm. Only six out of twenty-five untreated cabbage seedlings grew, but were yellow and wilted. Because of their smaller size, the cabbage seeds may have been more sensitive to the constituents than the other seeds. Survival rate was greater for the corn and soybean seedlings than for the cabbage. All of the corn and soybean untreated seedlings grew, but the leaf tips turned yellow and eventually brown. The treated groups continued to grow during the experiment and appeared green and healthy. Green, healthy seedlings were grown from the treated simulated PW, but not from the untreated simulated FOPW. Rate of seedling growth was greater for the treated water than for the untreated water.

An experiment was conducted to discern possible differences in AVS concentrations in the hydrosoil with changes in seasonal temperature. Production of sulfides in the wetland can indicate the presence and activity of dissimilatory sulfate reducing microbial assemblages. Following the method of Leonard and others (1996), acid-volatile sulfides (AVS) were measured as an indirect indicator of dissimilatory sulfate reduction in the pilot-scale CWTSs. A statistical analysis using a 2-sample T-test ( $\alpha$ =0.05) was done to determine if production of AVS was temperature-dependent. The analysis indicated no difference (p-value= 0.31) between the AVS concentrations produced between the winter and summer months. The consistent removal of divalent metals through the winter and summer months may be attributed to the steady production of sulfides by dissimilatory sulfate reducing microbial assemblages or storage of sulfides in the sediment.

#### Nutrient Amendment

The purpose of this research was to gain an understanding of Se-sequestering biogeochemical processes and evaluate the effects of a nutrient amendment on those biogeochemical processes, hydrosoil conditions (redox, pH, and organic matter percentage), Se accumulation, and Se-reducing microbial colonies in the hydrosoil of CWTSs. The two pilot-scale CWTSs [nutrient amended (AquaSmart<sup>TM</sup>) vs. unamended (control)] used in this study were designed and constructed to reduce aqueous Se concentrations in simulated produced water containing low molecular weight organics and metalloids. The specific objectives of this study were: (i) measure hydrosoil conditions, Se-reducing microbial colony forming units (CFUs), and Se accumulation vertically through the hydrosoil; (ii) investigate the potential for Se-sequestering biogeochemical processes (reduction, sorption/complexation) operating in the hydrosoil at various depths; and (iii) evaluate the effect of a nutrient amendment on hydrosoil conditions, microbial CFUs, Se accumulation, and potential for Se-sequestering biogeochemical processes in a pilot-scale CWTS cell designed to treat Se in PW.

This study utilized the first cell from two of the pilot-scale CWTSs (unamended and nutrient amended) designed and constructed in Phase II to treat low molecular weight organics and metalloids in simulated produced water. In the nutrient amended cell and unamended cell, detritus zone grab samples (7cm intervals) and sediment zone samples (3cm intervals) were collected (Figure 45). The nutrient amended cell's vertical hydrosoil profile consisted of three 7-cm thick detritus zone samples (0-21 cm) and five 3-cm thick sediment zone samples (21-36 cm) while the unamended cell's profile contained two 7-cm thick detritus zone samples (0-14 cm) and five 3-cm thick sediment zone samples (14-29 cm). To accomplish the first objective, redox potential was measured in-situ in the hydrosoil before sampling, and pH and organic matter percentage were measured in each sample. Se concentrations were determined for each sample using a microwave digestion procedure (CEM procedures: Microwave Sample Preparation Note, App. Note: OS-14) followed by analysis using an ICP-MS (EPA 200.8). Se-reducing CFUs were measured from pore water samples collected at each sampling interval of the hydrosoil in both cells. The second objective was accomplished by comparing the hydrosoil conditions and Se concentrations within the nutrient amended cell and unamended cell to a literature review of the hydrosoil conditions in which Se-sequestering biogeochemical processes operate (Table 61). Additionally, vertical CFU counts in each cell were used to provide insight into the Se-sequestering biogeochemical processes that may be occurring vertically through the hydrosoil. The third objective was accomplished by comparing hydrosoil conditions, vertical CFU count and vertical Se concentration of the nutrient amended cell with the hydrosoil conditions, vertical CFU count and vertical Se concentration of the unamended cell.

The detritus zone in the nutrient amended pilot-scale system was 7 cm thicker than that of the unamended system. The hydrosoil of each cell was near or within the targeted redox potential range (-150 to +50mV) for dissimilatory Se reduction (Table 62 and Figure 46). The pH was circum-neutral ranging from 6.2-6.8 in the nutrient amended cell and 6.2-6.5 in the unamended cell (Table 62 and Figure 47). The percent of organic matter decreased

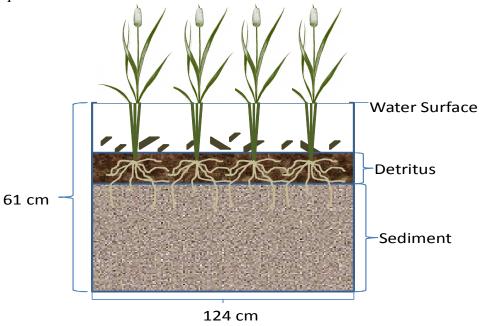
with depth in the hydrosoil in the first cells of both pilot-scale CWTSs (88% to 1% in the nutrient amended cell and 79% to 1% in the unamended cell) (Figure 48). In each cell, the percentage of organic matter in the detrital zone was greater than in the sediment zone (Table 62). Se concentrations in each cell decreased with depth (Figure 49). Concentrations of Se were greater in the detrital zone (76-830 ppm in the nutrient-amended cell and 138-569 ppm in the unamended cell) than in the sediment zone (0.9-212 ppm in the amended cell and 0.5-7 ppm in the unamended cell) (Table 62). Se-reducing microbes were present throughout the hydrosoil in both cells (Figure 50; Table 63).

Measured hydrosoil conditions were compared to hydrosoil conditions under which Se sequestering biogeochemical processes operate (Table 61). In the pilot-scale CWTS cells studied, the detritus zone and intervals of the sediment zone possessed favorable conditions for dissimilatory Se reduction and sorption/complexation (Table 61 and 62). Additionally, based on the high concentrations of Se measured in the detritus zone in the nutrient amended and unamended cells, Se sequestering biogeochemical processes are operating in the detritus zone. The percent of organic matter and Se concentrations in both cells decreased with depth and showed a similar trend suggesting that Se concentration may be influenced by organic matter. The presence of Se-reducing microbes in the hydrosoil of both cells indicates that dissimilatory Se-reduction can occur throughout the hydrosoil, but has the potential to operate at a greater rate in the hydrosoil where the number of CFUs is the greatest.

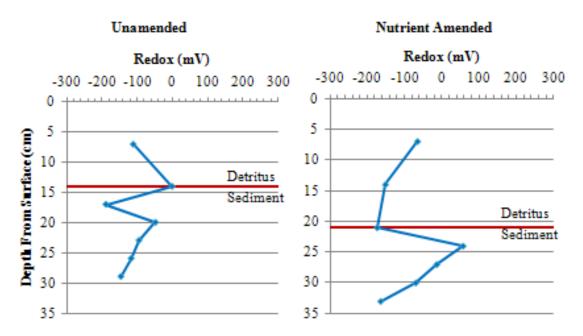
Redox potential in the hydrosoil of both cells ranged from -150 mV to +50 mV, but differed vertically between the nutrient amended cell and unamended cell (Table 62; Figure 46). There was little difference in hydrosoil pH between the nutrient-amended cell and the unamended cell. Organic matter percent was greater in the nutrient amended cell than in the unamended cell (example: 79% at 7 cm in the unamended vs. 88% at 7 cm in the nutrient amended) (Table 62; Figure 48). Comparison of Se concentrations between the cells showed that the detritus zone of the nutrient amended cell contained higher concentrations of selenium than the detritus zone of the unamended cell (830 ppm vs. 569 ppm). Se concentration and organic matter percent in both cells decreased downward through the detritus zone, which contained higher Se concentrations and percent organic matter than the sediment zone (Figure 48 and 49; Table 62). Se-reducing CFUs were greater in hydrosoil of the nutrient amended cell than in hydrosoil of the unamended cell (22,000CFUs/mL pore water) (Figure 50; Table 63).

The greater concentrations of Se in the detritus zone compared to the sediment zone show the importance of the detritus zone for Se-sequestering biogeochemical processes and Se accumulation. Furthermore, the higher Se concentrations in the detritus zone of the nutrient amended cell as compared to the unamended cell may be attributed to the higher percent organic matter in the detritus zone of the nutrient amendment cell compared to the detritus zone of the unamended cell. The nutrient amendment (AquaSmart<sup>TM</sup>) used in this study contained organic carbon, fermented yeast, and additional nutrients. In addition to providing a carbon source to promote Se-reducing microbial activity, nutrients contained in the AquaSmart<sup>TM</sup> amendment may have promoted *T. latifolia* growth within the nutrient amended cell and therefore increased the amount of *T. latifolia* litter falling onto the

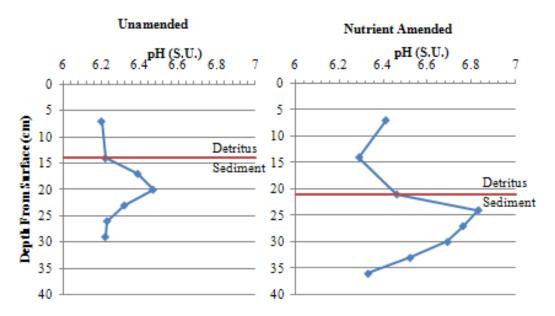
hydrosoil creating a thicker detritus zone. Selenium concentration, percent organic matter, and Se-reducing CFUs were greater in the nutrient amended cell, indicating a positive influence of the nutrient amendment on biogeochemical processes of Se accumulation and sequestration.



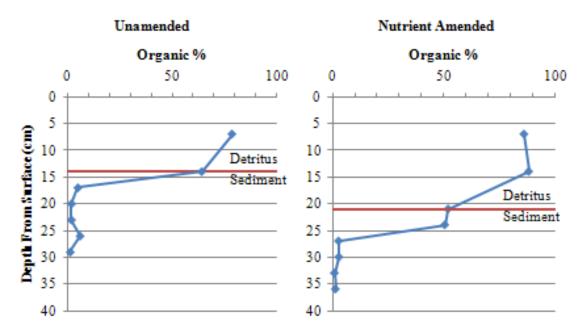
**Figure 45**. Cross-section of a pilot-scale CWTS cell showing hydrosoil layers. Based on pilot-scale CWTS cell construction, observation, and deconstruction.



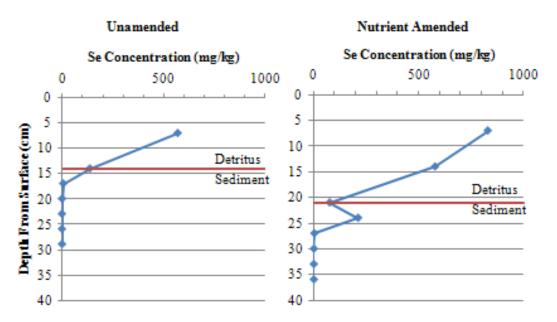
**Figure 46**. Redox with depth through the hydrosoil in the first cell (cell one) of each pilot-scale CWTS.



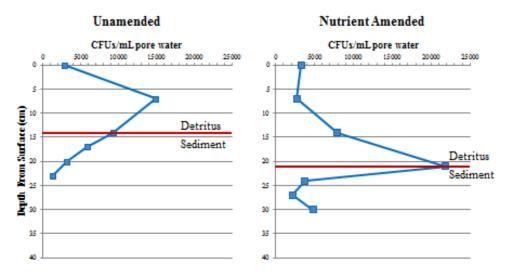
**Figure 47**. pH with depth through the hydrosoil in the first cell (cell one) of each pilot-scale CWTS.



**Figure 48**. Organic matter percentage with depth through the hydrosoil in the first cell (cell one) of each pilot-scale CWTS.



**Figure 49**. Se concentrations with depth through the hydrosoil in the first cell (cell one) of each pilot-scale CWTS.



**Figure 50**. Se-reducing microbial colony forming units (CFUs) with depth through the hydrosoil in the first cell (cell one) of each pilot-scale CWTS.

**Table 61**. Prevalent hydrosoil conditions for Se-sequestering biogeochemical processes within CWTSs.

Process	Organic Matter	рН	Redox Potential
Dissimilatory Se Reduction	High (> 0.1%)	Circum- Neutral (6.5-8)	Reducing (-150 to +50mV)
Sorption/ Complexation (organic & inorganic)	High (> 6%)	Acidic- Neutral (3-7)	Reducing-Oxidizing (-400 to +700mV)

(Trudinger and Swaine 1979, Shamberger 1983, de Souza et al. 1999, Selinus et al. 2005, Zhang et al. 2008, Rodgers and Castle 2008, Kadlec and Wallace 2009, Spacil 2010)

**Table 62**. Measured hydrosoil conditions and Se concentration with depth in nutrient amended and unamended CWTS cells.

	Depth (cm)	Redox (mV)	pH (S.U.)	Organic Matter (%)	Total Se (mg/kg)
Nutrient Amended Cell					
	0-7	-65	6.41	86.08	830.19
	7-14	-151	6.29	88.28	578.67
	14-21	-173	6.46	52.04	76.33
	21-24	57	6.83	50.57	211.91
	24-27	-14	6.76	2.89	3.28
	27-30	-70	6.69	3.08	1.17
	30-33	-164	6.52	1.04	0.89
	33-36	-161	6.33	1.45	0.97
Unamended Cell					
	0-7	-111	6.20	78.72	569.07
	7-14	-2	6.22	64.26	137.61
	14-17	-187	6.39	4.94	6.77
	17-20	-49	6.47	1.92	1.12
	20-23	-95	6.32	1.89	0.54
	23-26	-117	6.23	6.05	0.65
	26-29	-145	6.22	1.34	0.71

**Table 63**. Measured Se-reducing microbial colony forming units (CFUs) with depth in nutrient amended and unamended CWTS cells.

	Depth	CFUs/mL pore water
Nutrient Amended Cell		_
	0-7	2700
	7-14	8000
	14-21	22000
	21-24	3800
	24-27	2200
	27-30	4800
<b>Unamended Cell</b>		
	0-7	15000
	7-14	9300
	14-17	5800
	17-20	3200
	20-23	1300

### Oil-Water Separator

PW containing increased mass loads of O & G provided the opportunity to evaluate the performance of a pilot-scale oil water separator (OWS) followed by a specifically designed constructed wetland treatment system (i.e., pilot-scale OWS and wetland cells, termed hybrid-constructed wetland system) for achieving desired treatment goals. The pilot-scale hybrid CWTS consisted of an OWS and four subsurface-flow cells, each with a 24-hr hydraulic retention time. The OWS was based on an American Petroleum Institute (API) design; which targeted the fraction of oil that can be separated by physical means. The OWS was designed to take advantage of the difference in specific gravity between oil and water. Dimensions of the OWS were 122 cm long by 61 cm wide by 33 cm deep. Treatment performance of the OWS and CWTS cells was assessed by measuring O & G concentration in aqueous samples collected at the inflow of the OWS, outflow of the OWS, between each CWTS cell, and outflow from the CWTS.

Data from this experiment indicate that a hybrid CWTS produced a measureable decrease in outflow concentration of O & G (<1.4 mg/L) compared to a CWTS not containing an OWS (9.9-14.6 mg/L), with both systems operating at an inflow concentration of 100 mg/L O & G (Figures 49 and 50). Both systems effectively decreased the concentration of O & G to below irrigation and livestock watering guidelines. Removal efficiencies (%) and removal rate coefficients (*k*) for O & G were greater in the hybrid CWTS than in the CWTS only containing wetland cells (Table 64). This study has demonstrated that treatment performance in a CWTS is dependent on initial concentrations of O & G; however, when increased mass loads of O & G are encountered, an OWS in a hybrid

CWTS can contribute to effective treatment.

The effect of increased O & G mass loading by decreasing system HRT on treatment performance was investigated. Each series (SSF and FWS) was loaded with O & G at 100 mg/L targeted inflow concentration with an inflow rate providing a 4-day HRT (24-hr for each wetland cell), followed by loading at 100 mg/L O & G inflow concentration with a 2-day HRT (12-hr for each wetland cell). Nominal O & G mass loading was 10 mg/min at 100 mg/L inflow concentration and 4-day HRT, and 20 mg/min at 100 mg/L inflow concentration and 2-day HRT.

For all cells in both series, redox potential was lower at ~20 mg/min mass loading than at ~10 mg/min mass loading (Table 65). For example, redox potential in the first cell of each series ranged from -130 (SSF) to -147 mV (FWS) for 20 mg/min loading and from 115 (SSF) to 98 mV (FWS) for 10 mg/min loading. Oxidizing conditions (>-50 mV; Rodgers and Castle, 2008) were maintained in cell 4 in both series for all loadings of O & G. For O & G, in both the SSF and FWS series, removal extent was greater, removal rate coefficient was similar, and removal efficiency was less at ~20 mg/ min loading compared to ~10 mg/min loading (Table 66). Outflow concentrations of all four metals increased after mass loading was increased from ~10 to 20 mg/min, which is attributed to shorter HRT during ~20 mg/min loading (Table 67). Rate coefficients increased for nickel (1.04 d<sup>-1</sup>) and zinc (1.27 d<sup>-1</sup>) at ~20 mg/min loading compared to ~10 mg/min loading (0.913 d<sup>-1</sup> for nickel, 1.03 d<sup>-1</sup> for zinc). Lower rate coefficients for iron (0.452 d<sup>-1</sup>) and manganese (0.891 d<sup>-1</sup>) were observed at ~20 mg/min loading, compared to 0.786 d<sup>-1</sup> (iron) and 1.22 d<sup>-1</sup> (manganese) observed at ~10 mg/min loading.

A change from oxidizing (>-50 mV) to reducing (≤-50 mV) conditions in the hydrosoil occurred in both the SSF and FWS series as O & G loading increased from ~10 to 20 mg/min. Wetland cells exposed to the greatest mass loading of O & G (i.e. cell 1 in both series) experienced the greatest decreases in reduction-oxidation potential through the duration of the experiment. The change from oxidizing to reducing conditions in the hydrosoil with increased mass loading occurring in both the SSF series (cells 1 & 2) and FWS series (cells 1, 2, and 3) resulted in decreased removal efficiency of O & G from the water column. Removal of nickel and zinc was enhanced in both the SSF and FWS series with the change from oxidizing to reducing conditions coincident with increased mass loading. Results of this study demonstrate the importance of conditions, such as redox potential, in promoting biogeochemical processes for effective treatment in CWTSs and are applicable to the design and construction of full-scale systems.

**Table 64.** Differences in treatment performance between hybrid CWTS and non-hybrid CWTS.

Sampling Period	Parameter	Conc. (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
1	Influent	97.2		
1	Hybrid Extent	<1.4	98.5	1.13
2	Influent	101.2		
2	Hybrid Extent	<1.4	98.6	0.872
3	Influent	102.1		
3	Non-hybrid Extent	9.9	90.3	0.583
4	Influent	103.5		
4	Non-hybrid Extent	14.6	85.9	0.490

**Table 65.** Redox potential (mV) at different mass loadings of O & G for FWS and SSF series.

berreb.						
	1	2	3	4	5	6
O & G-inflow (mg/L)	98.7	96.9	101.2	99.3	103.5	98.6
O & G loading (mg/min)						
FWS series	10.28	10.10	10.55	10.35	21.57	20.55
SSF series	8.91	8.75	9.14	8.97	18.69	17.81
Redox Potential (mV)						
FWS series						
Cell 1	98	69	37	-91	-110	-147
Cell 2	20	-30	51	-10	-51	-91
Cell 3	49	-15	43	19	-67	-49
Cell 4	72	83	87	75	65	72
SSF series						
Cell 1	115	73	-5	10	-105	-130
Cell 2	67	-34	37	-24	-62	-71
Cell 3	-47	-23	63	39	22	-32
Cell 4	118	81	115	87	97	81

**Table 66.** O & G removal extents, efficiencies, and rate coefficients for all 6 sampling periods at different loadings of O & G.

Sampling Period	Series	O & G Loading (mg/min)	HRT (days)	Inflow Conc. (mg/L)	Extent* (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
1	SSF	8.91	4	98.7	7.9	92.0	0.631
	<b>FWS</b>	10.28	4	98.7	5.5	94.4	0.722
2	SSF	8.75	4	96.9	8.5	91.2	0.608
	<b>FWS</b>	10.10	4	96.9	6.7	93.1	0.668
3	SSF	9.14	4	101.2	9.9	90.2	0.581
	<b>FWS</b>	10.55	4	101.2	8.1	92.0	0.631
4	SSF	8.97	4	99.3	11.8	88.1	0.533
	<b>FWS</b>	10.35	4	99.3	14.6	85.3	0.479
5	SSF	18.69	2	103.5	30.8	70.2	0.606
	<b>FWS</b>	21.57	2	103.5	29.8	71.2	0.623
6	SSF	17.81	2	98.6	31.2	68.4	0.575
	<b>FWS</b>	20.55	2	98.6	35.3	64.2	0.514

<sup>\*</sup>Extent = concentration in series outflow

**Table 67.** Metal removal extents, efficiencies, and rate coefficients for sampling periods 4 and 6.

	Sampling	, , , , , ,	O & G	HRT	Inflow	Extent*	Removal	Rate Coef.
Metal	Period	Series	Loading	(days)	Conc.	(mg/L)	(%)	$(d^{-1})$
	1 CHOU		(mg/min) (uays)		(mg/L)	(IIIg/L)	(70)	(u )
Nickel	4	SSF	8.97	4	1.360	0.035	97.4	0.913
		<b>FWS</b>	10.35	4	1.360	0.220	83.8	0.455
	6	SSF	17.81	2	1.291	0.163	87.4	1.04
		<b>FWS</b>	20.55	2	1.291	0.417	67.7	0.565
Zinc	4	SSF	8.97	4	5.89	0.094	98.4	1.03
		<b>FWS</b>	10.35	4	5.89	0.103	98.3	1.01
	6	SSF	17.81	2	5.33	0.423	92.1	1.27
		<b>FWS</b>	20.55	2	5.33	0.611	88.5	1.08
Iron	4	SSF	8.97	4	0.431	0.019	95.7	0.786
		<b>FWS</b>	10.35	4	0.431	0.053	87.7	0.523
	6	SSF	17.81	2	0.420	0.170	59.5	0.452
		<b>FWS</b>	20.55	2	0.420	0.142	66.2	0.542
Manganese	4	SSF	8.97	4	1.314	0.010	99.2	1.22
-		<b>FWS</b>	10.35	4	1.314	0.150	88.6	0.542
	6	SSF	17.81	2	1.202	0.202	83.2	0.891
		<b>FWS</b>	20.55	2	1.202	0.426	64.5	0.518

<sup>\*</sup>Extent = concentration in series outflow

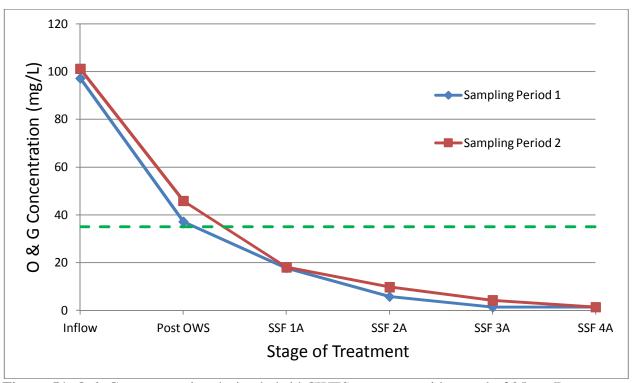
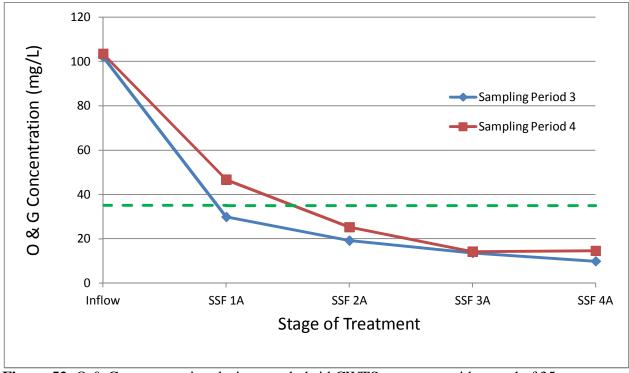


Figure 51. O & G concentration during hybrid CWTS treatment with a goal of 35 mg/L (green dashed line). 35 mg/L = reuse criteria for irrigation and livestock watering.



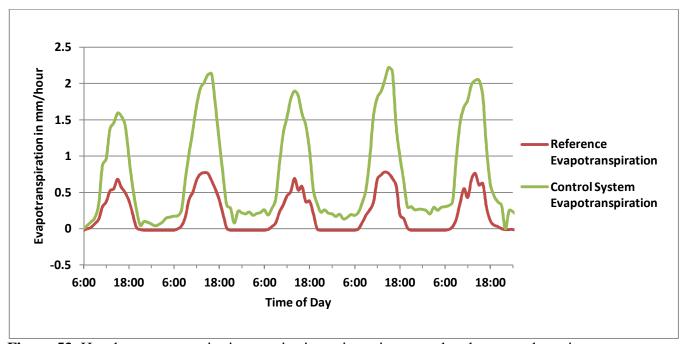
**Figure 52.** O & G concentration during non-hybrid CWTS treatment with a goal of 35 mg/L (green dashed line). 35 mg/L = reuse criteria for irrigation and livestock watering.

### Evapotranspiration and Tracer Tests

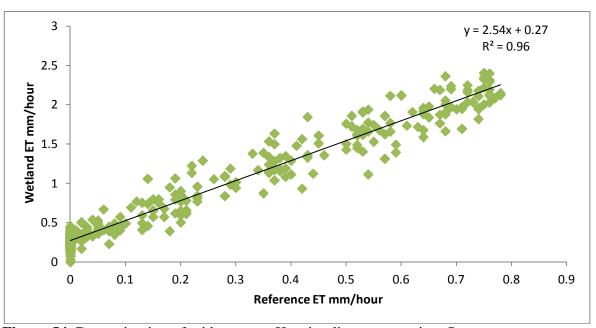
Evapotranspiration serves as the single source of water loss in CWTSs through the combined effects of open water evaporation and plant transpiration. The decrease in flow associated with evapotranspiration can alter performance by enriching constituents and decreasing the system hydraulic retention time. In order to determine potential effects of evapotranspiration on treatment performance, it is necessary to determine the amount of water lost during treatment. For this reason, evapotranspiration was monitored in a pilot-scale CWTS (post-RO control system) serving as a lysimeter and compared with reference evapotranspiration. Evapotranspiration was calculated using a simple mass balance in which the difference between the system inflow and outflow rates was assumed to be due to evapotranspiration. Inflow rates were measured using a stopwatch and graduated cylinder while outflow rates were measured using a digital rain gauge with an onboard data logger. Reference evapotranspiration was calculated with the FAO Penman Monteith equation (Allen et al, 1998) using data collected with an onsite weather station capable of recording temperature, wind speed and direction, barometric pressure, humidity, and solar radiation.

Results for measured CWTS evapotranspiration and reference evapotranspiration are shown in Figure 53. Results indicate a *Typha latifolia* (cattail) crop coefficient (K<sub>c</sub>) of 2.53 for data collected mid-summer during peak plant growth (Figure 54), which is within the range of published K<sub>c</sub> values measured using small stands of cattails (Snyder and Boyd, 1987; Rozkosny et al, 2006; Kadlec and Wallace, 2009). K<sub>c</sub> values are expected to change with seasonal variation in plant density and growth.

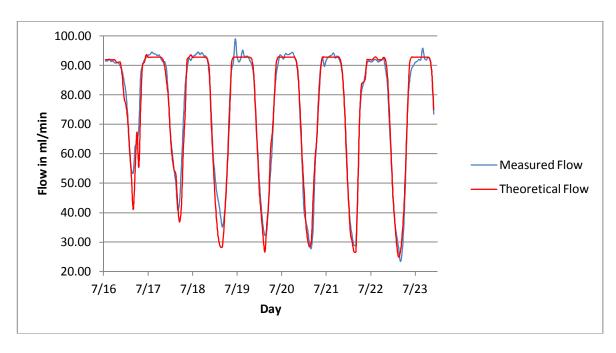
Theoretical lysimeter outflow determined using the Penman Monteith equation and midsummer peak  $K_c$  is shown along with measured lysimeter outflow in Figure 55. This comparison graphically demonstrates application of the Penman Monteith equation to estimating evapotranspiration in the pilot-scale CWTSs used in this investigation.



**Figure 53.** Hourly evapotranspiration monitoring using rain gauge data logger and on-site weather station.



**Figure 54.** Determination of mid-summer  $K_c$  using linear regression. System evapotranspiration can be estimated by multiplying reference evapotranspiration by  $K_c$  (2.54) and adding the open water evaporation term (0.27).



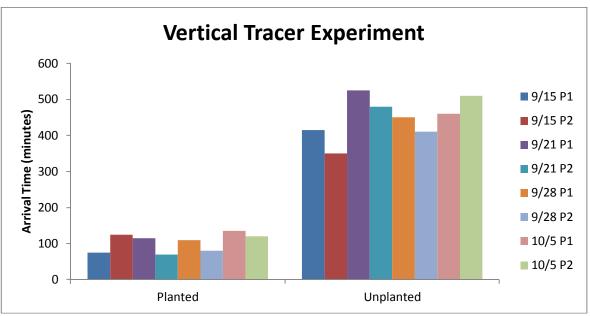
**Figure 55.** Comparison of theoretical lysimeter outflow vs. measured lysimeter outflow during mid-July showing application of the Penman Monteith equation for estimating water loss in pilot-scale CWTSs.

Plant transpiration may play a role in establishing a vertical hydraulic gradient within CWTS hydrosoil. This vertical hydraulic gradient may be a crucial component in initiating an advective flow path required for targeted constituents to reach the hydrosoil, where certain redox-driven reactions may occur (Kadlec, 1999; Martin et al., 2003; Kadlec and Wallace, 2009). For this reason, tracer tests were performed to determine the effect of evapotranspiration on vertical flux of constituents. Eight 20-L buckets were prepared as bench-scale CWTSs by filling each with approximately 5 L of sandy sediment collected from 18-Mile Creek located near Clemson, SC and approximately 8 *T. latifolia* plants. The systems were watered and fertilized regularly for a period of 9 months to allow for plant maturation. Surface water conductivity data collected from each system were used to formulate tracer solutions containing amounts of dissolved sodium chloride needed to yield conductivity ten times greater than that of the surface water in each system. This tracer solution concentration was selected to allow an accurate resolution of tracer arrival time.

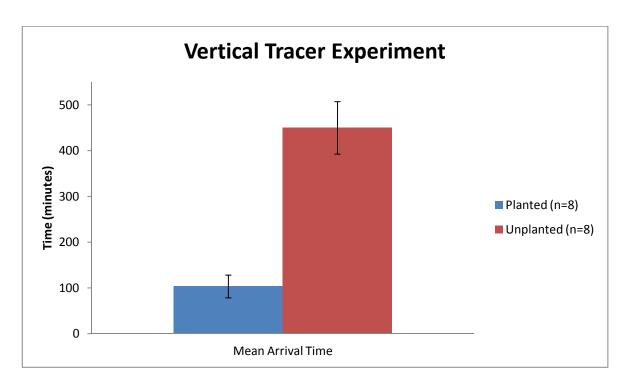
Eight individual tracer tests were performed by placing a pair of stainless steel conductivity probes 5 cm below the surface of the hydrosoil of a system. Prepared tracer solution was then added by a FMI® QG400 pump at a rate of 200mL/min. After 5 minutes, a second QG400 pumping at a rate of 200mL/min was connected to the system at a height of 25 cm above the hydrosoil to remove excess tracer solution, maintaining a constant head. This method allowed tracer solution to be added gently, preventing vertical flow disturbances while still allowing circulation of the tracer solution.

Soil conductivity was measured at 5-minute intervals until elevated conductivity was detected for a minimum of 3 consecutive measurements at each probe. *T. latifolia* in 4

randomly selected systems was trimmed to a height of 35 cm above the hydrosoil (or 5 cm above the waterline) just prior to the tracer test. Mean arrival times for the planted and unplanted cells were compared using Welch's t-test to determine if plant transpiration significantly alters flow through the hydrosoil in the systems ( $\alpha$ =0.05). Results indicated a significant difference (p= 1.2x10<sup>-8</sup>) in tracer arrival time between the planted cells and the cells containing no plants (Figures 56 and 57) indicating that plant evapotranspiration can affect vertical transport of constituents.



**Figure 56.** Arrival times for each probe during the 4 tracer tests performed on the planted systems and 4 on the unplanted systems. Lower arrival times in the planted systems indicate a more rapid transport of the tracer solution.



**Figure 57.** Mean arrival times of the 4 tracer tests performed on the planted systems and 4 on the unplanted systems. Error bars show standard deviation. The lower mean arrival time in the planted systems indicate a more rapid transport of the tracer solution.

## Design and Construction of Demonstration Wetland Treatment System

A demonstration CWTS was designed and constructed to treat CBMPW from the Blue Creek field of the Black Warrior basin in central Alabama. Site-specific produced water samples were characterized to determine COCs targeted for treatment. Three water samples were collected from an on-site holding tank where produced water from multiple wells is stored prior to disposal (i.e. WS-20). The samples were analyzed using standard methods (APHA, 2005), and the results compared with surface water quality criteria from the Clean Water Act Section 304(a) to identify COCs. These criteria are recommended water quality criteria for the protection of aquatic life and human health in surface water for various beneficial uses. Comparison of analyzed constituents with water quality criteria (Table 68) yielded the following COCs: ammonia, barium, cadmium, iron, manganese, nitrate, and selenium. Because results indicate that composition of the produced water analyzed varies with time, the demonstration CWTS was designed to promote conditions required for treatment of both site-specific COCs identified from the water samples analyzed and additional generic COCs identified in produced waters characterized in Task 2. Conditions required for removal of ammonia, cadmium, iron, manganese, nitrate, oil and grease, and selenium identified during Phase II (Tables 20-23, 58, 61, and 62) were targeted in designing the demonstration system. Oxidizing conditions (DO greater than 2.0 mg/L) were targeted in some of the demonstration CWTS cells to promote microbial nitrification of ammonia, biodegradation of oil and grease, and precipitation of iron and

manganese. Reducing conditions (DO less than 2.0 mg/L) were targeted in the remaining cells to promote microbial denitrification of nitrate, sulfide complexation with divalent metals through dissimilatory sulfate reduction, and dissimilatory selenium reduction. Barium was not targeted for pilot-scale treatment during Phase II; however, removal through chemical precipitation can occur under the conditions observed (pH between 2 and 12) in the pilot-scale CWTSs when sulfate is present (Brookings, 1988).

The demonstration CWTS consists of sixteen 100-gallon Rubbermaid<sup>®</sup> containers arranged in four series with four containers as wetland cells in each series. Each cell was filled to a depth of approximately 25 cm with sand and then planted with cattails (*Typha latifolia* and *Typha angustifolia*) collected from a pond located near the site. Connections between cells were made using 2.5-cm poly-vinyl chloride (PVC) pipes to allow gravity flow through each series. Water is pumped from a 5,678-L polypropylene carboy holding tank to the first cell in each series by FMI<sup>®</sup> piston pumps. Each pump was calibrated for a hydraulic retention time of 24 hours per cell. The 24-hour HRT was selected based on rate coefficients determined from the pilot-scale experiments. The outflow pipes from each cell were connected to a small basin, from which a sump pump moves the outflow water to a second 5,678-L polypropylene carboy holding tank for return to the existing waste stream (Figures 58-61).

The demonstration CWTS contains two pairs of replicated series, each pair receiving different hydrosoil amendments. Dried yeast in the form of AquaSmart was added to the hydrosoil of cells 2 and 3 of the first two series (series A and B; approximately 10 kg per cell) to serve as a source of organic carbon to promote reducing conditions and to provide nutrients for both the plants and microbes. Shredded hardwood mulch (approximately 10 kg per cell) was added to the hydrosoil of cells 2 and 3 of the second two series (series C and D) to serve as a source of organic carbon and promote reducing conditions. Cells 1 and 4 in each of the four series were designed as oxidizing cells and therefore did not receive any organic amendment. The oxidizing and reducing cells are capable of treating constituents that are not currently identified as COCs, but that may be present in the future at concentrations exceeding water quality criteria. Municipal (fresh) water was introduced initially to the wetland series to allow for growth of plants and microbes prior to introduction of produced water.

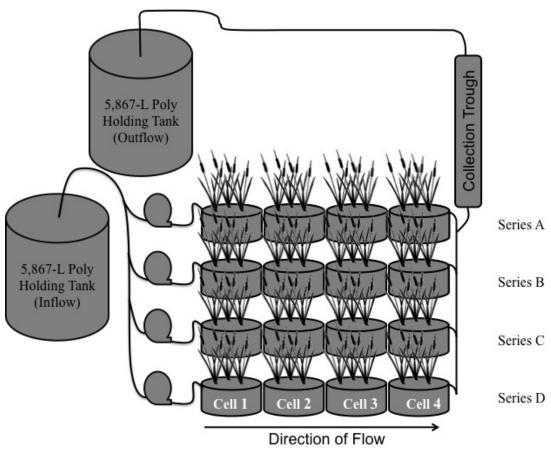
Table 68. Characterization of untreated CBMPW samples collected from the WS-20 tank at demonstration site and

surface water quality criteria. Constituent concentrations are mg/L unless noted.

Parameters <sup>1</sup>	6/7/2011 Sample	5/29/2012 Sample	6/11/2012 Sample	Water Quality Criteria <sup>2</sup>
Conductivity (µm/cm)	7860	11100	6950	NA
pH (S.U.)	8.49	7.98	8.35	NA
Alkalinity (as CaCO <sub>3</sub> )	496	406	450	NA
Hardness (as CaCO <sub>3</sub> )	226	504	468	NA
Ammonia (as N)	2.8	2.1	2.0	1.4
Nitrate	NM	12.2	NM	10
Oil and Grease	<2.8	<2.8	NM	10
Sulfate	NM	4.6	NM	NA
Total Dissolved Solids	NM	6420	6850	NA
Total Suspended Solids	NM	10	10	NA
Al	< 0.045	0.130	< 0.045	NA
As	0.00074	< 0.053	< 0.053	0.15
Ba	29.6	13.8	11.0	1.0
Be	< 0.0003	NM	< 0.0003	0.004
Br	NM	6.80	NM	NA
Ca	93.6	72.2	62.7	NA
Cd	< 0.0034	< 0.001	0.0315	0.00025
Cl	NM	3740	NM	NA
Co	< 0.007	NM	< 0.007	NA
Cr	< 0.0061	< 0.0061	< 0.0061	0.011
Cu	< 0.003	< 0.003	< 0.003	0.009
Fe	< 0.0062	0.35	1.6	1.0
Hg	< 0.0025	NM	NM	NA
Mg	53.7	29.0	28.0	NA
Mn	< 0.0014	0.06	< 0.0014	0.05
Na	NM	1900	25.6	NA
Ni	< 0.015	0.01	< 0.015	0.61
Pb	< 0.042	< 0.0015	< 0.0015	0.0025
Se	0.0032	0.019	NM	0.005
Sr	6.81	NM	2.08	NA
V	< 0.0075	NM	< 0.0075	NA
Zn	<0.0018	0.020	< 0.0018	0.12

<sup>&</sup>lt;sup>1</sup>Metal concentrations are acid soluble concentrations.
<sup>2</sup>Current water quality criteria from the Clean Water Act Section 304(a), chronic toxicity values used where available NA - No suggested criteria from Clean Water Act Section 304(a)

NM - Not measured.



**Figure 58.** Schematic diagram of demonstration CWTS. Produced water is pumped from the 5,867-L polypropylene carboy holding tank through the 4 series to an outflow holding tank. Cells of each series are arranged for gravity flow. Cells 1 and 4 of each series were designed to promote oxidizing conditions, and cells 2 and 3 were designed to promote reducing conditions with the addition of AquaSmart<sup>TM</sup> (series A and B) or mulch (series C and D).



Figure 59. Filling the demonstration CWTS cells with sand.



**Figure 60.** Broadleaf and narrowleaf cattails (*Typha latifolia* and *Typha angustifolia*) were collected from a nearby pond and transplanted to the demonstration CWTS cells at a density of 30 plants per cell.



**Figure 61.** Demonstration CWTS after planting was completed on 5/31/12. Municipal water was introduced initially to the four series to allow growth and maturation of the plants and microbes.

# Measure Treatment Performance of Demonstration Constructed Wetland System (Task 8)

Produced water was introduced to the system on September 3, 2012 (14 weeks after construction), and samples were collected from the inflow and outflow of each cell on 9/6/12, 9/20/12, 10/4/12, 10/18/12, 11/1/12, 11/15/12, 11/29/12, 12/13/12, 1/10/13, 1/24/13, 2/7/13, 2/21/13, 3/14/13, and 3/28/13 for a total of fourteen sampling periods, numbered 1 through 14, respectively. Samples from were analyzed for various constituents (Table 2) and explanatory parameters (Tables 69-82). Of the COCs identified initially in design of the demonstration wetland treatment system, ammonia, Ba, Cd, Fe, and Mn concentrations in the PW inflow exceeded the water quality criteria (WQC) during one or more sampling periods. Se and nitrate concentrations did not exceed WQC during any of the sampling periods.

Removal efficiencies, removal rate coefficients, and removal extents of ammonia, Ba, Cd, Fe, and Mn in the demonstration system were calculated (Tables 83-87) to determine efficiency and effectiveness of the design for each cell, which was based on biogeochemical treatment processes. Removal efficiency was calculated using the following equation:

Removal efficiency (%) = 
$$\left(\frac{[C]_0 - [C]}{[C]_0}\right) \times 100$$
 Eqn. 10

where [Co] is concentration (mg/L) of a COC in the inflow and [C] is concentration (mg/L) in the outflow. Removal rate coefficients were calculated using the standard approach of first-order rate kinetics (Horner et al. 2012; Johnson et al. 2008b). The first-order reaction follows:

$$k = \frac{-\ln([C]/[C]_0)}{t}$$
 Eqn. 11

where k is the rate coefficient (day<sup>-1</sup>), [Co] is the initial inflow COC concentration, [C] is the final outflow COC concentration, and t (days) is the HRT for the system. Comparing treatment goals with calculations of performance parameters will assess success of the demonstration system.

Ammonia concentration was less than the WQC (1.4 mg/L) in both inflow and outflow during the first three sampling periods (Tables 69-71). During the subsequent sampling periods (Tables 72-82), ammonia was present in concentrations above the WQC, and concentrations decreased in each series to less than the WQC (Figure 62, Table 83). Nitrate concentration was measured in samples collected only during sampling period 6 (Figure 63), and the nitrate concentrations measured in the outflow were greater than what would be produced if all the ammonia in the inflow were converted to nitrate. This indicates that the ammonia in the inflow was being removed as well as additional ammonia produced in the system.

During sampling periods 1 through 5, Ba concentration in outflow was less than the WQC of 1.0 mg/L (Figures 64, Tables 69-73). Over 90% of the inflow concentration of Ba was

removed during each of these sampling periods (1 through 5) (Table 84). During sampling periods 6 and 7 (Tables 74 and 75), no Ba removal occurred, which was interpreted as due to a lack of sulfate in the wetland cells (Figure 65, Table 84). Sulfate concentration measured during sampling period 6 indicated that the amount of sulfate available was insufficient for decreasing Ba concentrations. Gypsum was added to all wetland cells during sampling period 7 on 11/29/12 as a source of sulfate. After the addition of gypsum, removal increased during sampling periods 8-10 (Figure 66, Table 84).

Cd concentrations decreased to below WQC (0.00025 mg/L) from the produced water by treatment in the wetland cells during the first two sampling periods (Figure 67, Table 85). After the second sampling period, Cd was not present in the PW in concentrations greater than the WQC.

Fe concentration was less than the targeted performance goal of 1.0 mg/L in both inflow and outflow for sampling periods 1-3 and 7-14 (Tables 69-71 and 75-82). During sampling periods 4, 5, and 6, Fe concentration in the inflow was greater than 1.0 mg/L with decrease in concentration from inflow to outflow in all series (Figure 68, Table 86).

Mn concentration exceeded the WQC (0.05 mg/L) in inflow during the first sampling period, and no removal occurred in the wetland cells (Table 69 and 87). Decrease in concentrations of Mn occurred during the second sampling period in series B and C (Table 70 and 87). During sampling period 4, Mn was present in the inflow and decreased in concentration to below WQC in all wetland series (Table 72). Mn concentration was below the WQC in sampling periods 3, 5-7, and 11-12. During sampling periods 8-10, Mn concentrations were below the WQC of 0.05 mg/L in inflow and outflow, and removal occurred in all series (Tables 76-78 and 87). During sampling periods 13-14, inflow Mn concentration was 0.0659 and 0.0771 mg/L, respectively, and decreased to below WQC in all series.

Results indicate that the demonstration CWTS can reduce concentrations of COCs as they vary through time. The system successfully achieved consistent removal extents although inflow concentrations differed by orders of magnitude. Plant growth (Figure 69) and explanatory parameters measured in the wetland treatment cells during all sampling periods were favorable for decreasing concentrations of COCs in the produced water, with the exception of Ba during sampling periods 6 and 7 as discussed above. After the addition of gypsum to all wetland cells, Ba removal increased during the remaining sampling periods, indicating that the targeted concentration can be achieved by properly adjusting wetland conditions. Upon the conclusion of sampling to measure treatment performance, the demonstration system was disassembled on May 6, 2013.

**Table 69.** COC concentrations and explanatory parameters for Sampling Period 1 (9/6/12)

	Performance Parameters						Explanatory Parameters						
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	Cadmium (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.3	7.7	0.2758	0.1244	0.000108		nd	6710	nd	nd	nd	208	1620
$A1^a$	1.1	6.4	0.2201	1.114	0.000032		2.85	6570	7.58	-137	26.4	420	1620
A2	1.5	3.5	0.5943	1.333	< 0.000025		1.3	4490	7.69	-233	27.2	320	1800
A3	1.6	1.4	0.8435	1.503	< 0.000025		2.34	2608	7.02	-325	26.5	130	1260
A4	2.3	0.45	0.6978	2.272	< 0.000025		2.53	1240	7.22	60	26.4	76	1260
B1	0.5	8.1	0.4869	0.7397	0.000037		3.33	6690	6.86	61	27	208	1620
B2	0.4	3.9	0.6422	1.204	0.000031		4.02	4386	6.99	-201	26.4	156	1260
В3	0.4	1.2	0.3977	3.082	< 0.000025		3.6	2630	7.02	-94	25.8	114	1440
B4	0.6	0.13	0.3139	2.658	0.000034		4.1	1354	6.65	151	25.6	78	1080
C1	0.5	6.5	0.1494	1.11	0.000154		2.3	6690	7.45	142	27.6	198	1620
C2	0.4	2.8	0.1517	2.15	0.000056		1.9	4565	7.1	-276	27.1	150	1440
C3	0.4	0.67	0.0915	0.5587	0.000063		3.75	2630	7.27	-216	26.6	104	1440
C4	0.3	0.26	0.0846	0.6731	0.00003		3.3	1343	7.05	-125	26.2	46	900
D1	0.4	6.8	0.0767	0.3225	0.000029		2.1	6590	7.57	-109	26.6	200	1440
D2	0.4	2.7	0.0375	1.113	0.000033		2.29	4601	6.98	-142	25.7	162	1620
D3	0.4	0.55	0.0783	1.874	0.00004		1.59	2565	6.97	-232	26.7	70	1080
D4	0.3	0.21	0.1755	1.019	0.000107		1.12	1354	6.96	178	26.4	64	900

Se below detection limit (bdl) (0.0002 mg/L).

<sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 70.** COC concentrations and explanatory parameters for Sampling Period 2 (9/20/12)

	Performance Parameters								Explanat	ory Parame	Explanatory Parameters						
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	Cadmium (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )				
Inflow	0.1	6.1	0.1006	0.0200	0.000281		nd	7970	nd	nd	nd	432	1260				
$A1^a$	< 0.03	5.0	0.0548	0.0344	0.000061		4.65	7700	7.71	-10	18.2	550	1080				
A2	0.8	3.6	0.2437	0.2638	0.000035		4.2	6780	7.46	-415	17.5	490	1260				
A3	< 0.03	1.7	0.368	0.2174	0.000101		6.91	6730	7.47	-385	17.5	396	1440				
A4	0.1	0.54	0.2032	0.0372	< 0.000025		4.38	6870	7.39	106	17.7	416	1440				
B1	< 0.03	5.1	0.0186	0.0050	0.000036		6.13	7970	7.97	59	18.7	422	1440				
B2	< 0.03	3.7	0.0203	0.0071	0.000027		7.08	7970	7.82	-225	18.3	418	1260				
В3	< 0.03	1.1	0.0245	0.0115	0.000025		3.7	7770	7.68	-243	17.7	422	1260				
B4	< 0.03	0.43	0.0138	0.0028	< 0.000025		4.39	7670	7.58	130	17.6	402	1440				
C1	< 0.03	5.2	0.0392	0.0196	0.00014		7.46	7830	7.61	111	19.8	430	1260				
C2	< 0.03	3.2	0.0388	2.830	0.00007		8.9	7400	7.49	-346	19	450	1440				
C3	< 0.03	1.9	0.0111	0.0242	0.000052		2.9	6560	7.65	-216	18.1	362	1440				
C4	< 0.03	0.31	0.0093	0.0146	0.000027		2.8	6050	7.22	80	18.1	340	900				
D1	< 0.03	5.2	0.0444	0.0144	0.000036		2.56	7400	7.83	89	18.3	368	1260				
D2	< 0.03	2.6	0.0260	0.0209	0.000088		4.27	6010	7.79	-170	18	274	1080				
D3	< 0.03	1.1	0.0337	0.0892	< 0.000025		1.4	7010	7.77	-213	17.3	212	900				
D4	< 0.03	0.24	0.0162	0.0541	< 0.000025		3.2	5680	7.48	143	17.5	216	1080				

Se bdl (0.0002 mg/L).

<sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 71.** COC concentrations and explanatory parameters for Sampling Period 3 (10/4/12)

_	Performance	Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	0.5	9.3		nd	9760	nd	nd	nd	456	2160
$A1^a$	0.1	7.9		4.2	9650	7.71	-22	24.9	446	1800
A2	1.0	4.1		6.67	9520	7.73	-221	25.2	428	1980
A3	0.9	1.6		6.29	9010	7.07	-320	24.7	410	1980
A4	0.1	0.28		4.26	8420	7.24	122	25.6	394	1800
B1	0.1	7.8		4.25	9360	7.55	305	24.9	432	1800
B2	0.1	4.2		4.55	9520	7.6	-227	25	422	1800
В3	< 0.03	1.9		4.66	9570	7.28	-240	23.7	420	1800
B4	< 0.03	0.17		4.21	9520	7.18	120	24	446	1800
<b>C</b> 1	0.1	7.4		3.06	9680	7.11	124	24.3	430	1800
C2	< 0.03	3.3		7.08	9700	7.39	-350	23.7	440	1800
C3	< 0.03	1.9		6.81	9660	7.67	-212	24.1	462	1980
C4	< 0.03	0.82		5.11	9690	7.77	90	24.5	450	1800
D1	0.1	7.5		4.87	9580	7.89	74	24.1	448	1980
D2	< 0.03	4.5		6.37	9690	7.68	-154	22.7	448	1800
D3	< 0.03	1.4		3.2	9650	7.07	-230	22.5	458	1800
D4	< 0.03	0.39		3.7	9760	7.47	168	23.9	442	1800

Cd bdl (0.000025 mg/L), Fe bdl (0.0062 mg/L), Mn bdl (0.0014 mg/L), Se bdl (0.0002 mg/L).

<sup>&</sup>lt;sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 72.** COC concentrations and explanatory parameters for Sampling Period 4 (10/18/12)

Performance Parameters					Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )	
Inflow	3.3	17.0	2.44	0.169		nd	18300	nd	nd	nd	430	6660	
$A1^{a}$	2.6	13.0	1.32	0.104		3.85	24100	6.96	189	26.8	380	5400	
A2	2.4	7.4	0.389	< 0.0014		7.35	19200	7.24	-347	27	318	5220	
A3	2.4	3.7	0.247	< 0.0014		7.01	16100	7.23	-331	27	264	5040	
A4	1.4	1.2	0.088	< 0.0014		6.71	17070	7.18	181	27.2	280	6120	
B1	3.2	11.3	1.399	0.554		3.8	17900	6.41	211	26.8	400	5940	
B2	2.5	6.7	0.324	0.113		5.3	17050	6.77	-228	23.7	312	5580	
В3	1.8	2.9	0.091	< 0.0014		4.36	18100	7.41	-238	23.2	248	4500	
B4	1.3	0.91	0.016	< 0.0014		3.46	17950	7.18	155	24	254	4860	
C1	3	15.9	1.107	0.341		2.4	23100	7.11	108	24.3	398	6300	
C2	2.3	10.9	0.117	0.051		2	24300	7.39	-394	23.7	326	5220	
C3	1.3	5.7	0.033	< 0.0014		3.65	22300	7.67	-216	24.1	236	5400	
C4	0.9	0.59	0.028	< 0.0014		2.71	18800	7.77	172	24.5	186	3240	
D1	3.2	14.6	2.29	0.21		4.8	19060	7.89	98	24.1	400	5400	
D2	3	10.7	0.351	0.006		5.4	17530	7.68	-174	22.7	344	5220	
D3	1.8	6.6	0.074	< 0.0014		3.28	18500	7.07	-209	22.5	258	3960	
D4	1.3	0.42	0.017	< 0.0014		3.69	17100	7.47	163	23.9	198	2880	

nd- not determined
Cd bdl (0.00025 mg/L), Se bdl (0.0002 mg/L).

a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 73.** COC concentrations and explanatory parameters for Sampling Period 5 (11/1/12)

	Perfori	mance Param	neters		Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )			
Inflow	2.3	13.2	1.48	nd	9120	nd	nd	nd	102	2070			
A1 <sup>a</sup>	1.9	11.6	0.43	7.73	11650	8.14	157	16.1	516	1440			
A2	1.2	8.1	0.269	11.4	10360	8.28	-319	15.3	506	2700			
A3	0.8	4.7	0.115	13	10510	7.88	-204	14.3	498	2700			
A4	0.1	1.4	0.047	14.3	11200	8.02	210	14.5	480	2340			
B1	2.2	10.3	0.907	2.06	12940	8.25	226	17.3	516	2160			
B2	1.9	7.9	0.167	4.9	10380	7.76	-219	13.1	508	2340			
В3	1.8	5.7	0.109	10.2	10170	8.01	-219	12.4	504	1980			
B4	1	1.1	0.02	5.91	11520	7.84	162	13.9	490	1800			
<b>C</b> 1	1.5	11.6	0.98	5.6	12600	6.54	329	15.9	538	1980			
C2	1.5	7.7	0.519	3.23	12440	6.82	-268	16.8	516	1980			
C3	1.9	3.1	0.496	4.1	13270	7.02	-195	14.9	520	2520			
C4	0.9	0.9	0.331	2.9	12010	6.5	242	15.9	490	2340			
D1	2	9.9	0.476	6.64	10510	7.89	161	15.9	520	1800			
D2	1.9	6.2	0.248	4.1	11040	7.68	-196	16.8	540	1980			
D3	1.5	2.6	0.14	2.33	11450	7.07	-154	14.9	510	1980			
D4	0.9	0.61	0.104	3.35	11170	7.47	172	15.9	504	1800			

Cd bdl (0.000025 mg/L), Mn bdl (0.0014 mg/L), Se bdl (0.0002 mg/L). a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 74.** COC concentrations and explanatory parameters for Sampling Period 6 (11/15/12)

	Perfor	mance Paran	neters		Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )			
Inflow	2.5	28.6	1.524	nd	13920	nd	nd	nd	470	3860			
$A1^a$	1.9	28.3	0.38	11.19	18410	7.79	197	20.4	464	3780			
A2	1.3	27.3	0.38	11.4	14460	7.98	-300	18.9	468	3420			
A3	1.2	29.7	0.589	13	14550	8.43	-189	18.8	430	3960			
A4	1.1	28.8	0.119	12.2	14420	8.31	257	18.1	402	3780			
B1	1.8	29.4	1.706	4.5	18600	7.19	252	19.5	472	3600			
B2	1.7	27.4	0.272	10.6	14500	8.25	-215	18	470	3420			
В3	1.6	29.7	0.172	6.75	16010	7.55	-218	17.2	452	3420			
B4	1.2	29.6	0.07	8.37	14830	7.56	181	17.3	454	3960			
C1	1.9	30.5	0.611	9.87	18290	7.09	259	19.5	486	4320			
C2	1.6	28.2	0.418	10.1	15970	7.27	-244	19.5	470	5040			
C3	1.4	26.5	0.386	10.07	15800	6.68	-186	19.3	454	3600			
C4	1.2	25.6	0.123	9.8	14540	8.27	253	18.8	460	4680			
D1	1.8	26.9	1.742	6.6	18500	7.07	168	19.3	470	4140			
D2	1.7	26.2	0352	10.8	17020	7.03	-100	18.4	460	3780			
D3	1.4	26.1	0.255	2.53	14870	6.71	-25	17.7	468	3780			
D4	1.2	25.0	0.144	12.7	14430	8.41	184	18.4	446	3420			

Cd bdl (0.000025 mg/L), Mn bdl (0.0014 mg/L), and Se bdl (0.0002 mg/L).

<sup>&</sup>lt;sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 75.** COC concentrations and explanatory parameters for Sampling Period 7 (11/29/12)

	Perforr	nance Parar	neters		Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )			
Inflow	2.7	12.6	0.262	nd	9670	nd	nd	nd	456	2160			
$A1^a$	1.8	13.0	0.132	7.73	10760	7.98	202	14.7	446	1800			
A2	1.5	13.2	0.156	11.4	10400	7.8	-229	15.4	428	1980			
A3	1.2	14.5	0.126	13	10830	7.98	-200	13.4	410	1980			
A4	0.8	14.6	0.109	12.2	10900	7.95	342	12	394	1800			
B1	1.8	13.5	0.179	4.5	10190	6.91	257	14.2	432	1950			
B2	1.6	13.7	0.118	10.6	10560	7.4	-220	12.3	422	1890			
В3	1.3	14.2	0.088	10.2	10800	6.82	-227	12.8	420	1800			
B4	0.9	13.9	0.055	5.91	12800	7.21	172	11.6	446	1800			
C1	1.9	14.2	0.232	5.6	14300	6.54	262	14.8	430	1800			
C2	1.5	14.3	0.178	10.1	13080	6.78	-243	15.3	440	1800			
C3	1.4	14.2	0.137	10.07	10100	6.84	-205	14.1	462	1980			
C4	0.8	15.5	0.089	9.8	10680	7.56	262	12.1	450	1800			
D1	1.5	14.2	0.254	6.6	10600	6.73	245	13.5	448	1980			
D2	1.2	12.9	0.042	10.8	13300	7.05	-197	13.2	448	1800			
D3	1.1	14.1	0.023	6.6	12400	7.24	44	12.5	458	1800			
D4	0.8	14.0	0.013	9.8	10260	7.31	139	12.9	442	1800			

Cd bdl (0.000025 mg/L), Mn bdl (0.0014 mg/L), and Se bdl (0.0002 mg/L). <sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 76.** COC concentrations and explanatory parameters for Sampling Period 8 (12/13/12)

	Performance Parameters					Explanatory Parameters								
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )		
Inflow	2.9	7.5	0.7842	0.0299		nd	6800	nd	nd	nd	490	2160		
$A1^{a}$	1.2	7.4	0.1299	0.0175		6.3	7380	7.98	181	13.7	452	1440		
A2	1.1	5.8	0.1933	0.0239		8.1	7280	7.37	-320	13.4	476	1440		
A3	0.8	3.8	0.2512	0.0275		8.9	7430	7.98	-299	12.4	430	2160		
A4	0.4	1.8	0.2168	< 0.0014		10.4	7470	7.01	257	12	462	1800		
B1	1.2	7.4	0.1696	0.0114		7.1	7030	6.91	252	12.2	472	1440		
B2	1.1	5.5	0.1068	0.0099		9.8	7300	7.4	-215	11.3	470	1980		
В3	0.8	3.4	0.1314	0.0035		10.2	7350	6.82	-218	12.8	452	1080		
B4	0.5	1.9	0.0990	< 0.0014		8.6	7390	7.04	180	11.6	484	1080		
C1	1.6	7.7	0.2120	0.0237		6.5	9300	6.54	260	13.8	486	2160		
C2	1.2	6.7	0.2352	0.0202		10.4	10000	6.74	-272	13.3	470	2520		
C3	0.6	4.8	0.2857	0.0171		9.5	7130	6.84	-183	12.1	454	1800		
C4	0.2	2.9	0.1481	< 0.0014		9.9	7200	7.56	213	11.1	514	2340		
D1	1.2	7.4	0.1149	0.0189		8.9	9210	6.42	171	12.5	470	1800		
D2	1	7.1	0.3626	0.0196		10.7	11900	7.05	-200	12.2	460	1980		
D3	0.9	5.8	0.0566	< 0.0014		8.6	7240	6.24	-47	11.5	468	2340		
D4	0.3	3.1	0.0554	< 0.0014		9.2	7500	7.31	174	11.9	506	1620		

nd- not determined
Cd bdl (0.00025 mg/L), Se bdl (0.0002 mg/L).

a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 77.** COC concentrations and explanatory parameters for Sampling Period 9 (1/10/13)

		Perform	nance Param	neters			Exp	olanatory l	Parameters	S	
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.6	7.4	0.1316	0.0341	nd	8100	nd	nd	nd	490	1080
$A1^a$	1.1	7.1	0.1171	0.1237	6.7	9380	6.86	181	13	452	2160
A2	0.8	5.7	0.1043	0.1023	9.4	10000	7.37	-396	12.2	476	2520
A3	0.5	3.9	0.0929	0.0329	10	9500	7.56	-379	12	430	1800
A4	0.1	1.1	0.0912	0.0274	10.2	8870	8.1	277	11.8	462	2340
B1	1.4	6.7	0.468	0.421	6.65	8900	7.01	255	11.7	472	1800
B2	1	5.2	0.1082	0.1423	10.6	11600	7.16	-226	11.6	470	1980
В3	0.7	3.2	0.0717	0.0513	10.2	9020	7.04	-238	11.2	452	2340
B4	0.3	1.5	0.0444	0.0079	7.91	9010	7.6	170	11.6	484	1620
C1	1.3	7.3	0.3882	0.0861	6.3	13900	6.77	255	11.7	456	1440
C2	0.9	5.5	0.1809	0.1643	10.1	12800	6.74	-308	11.4	446	1440
C3	0.5	4.3	0.1543	0.0471	9.9	9990	6.7	-283	12	428	2160
C4	0.1	3.1	0.0675	< 0.0014	9.6	8850	7.99	220	11.5	410	1800
D1	1.1	7.3	0.1268	0.1094	9.3	12400	6.42	218	11.8	394	1440
D2	0.8	6.3	0.104	0.0643	10.8	10200	6.68	-267	11.5	432	1980
D3	0.6	4.4	0.0561	0.0089	8.6	10190	6.7	-81	11.8	422	1080
D4	0.2	3.4	0.0060	0.0111	9.9	9750	6.68	274	11.3	420	1080

nd- not determined
Cd bdl (0.00025 mg/L), Se bdl (0.0002 mg/L).

a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 78.** COC concentrations and explanatory parameters for Sampling Period 10 (1/24/13)

		Perform	nance Param	eters			Exp	olanatory l	Parameters	3	
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.7	6.2	0.3554	0.0385	nd	7300	nd	nd	nd	460	2080
$A1^{a}$	0.9	5.7	0.3114	0.0221	8.1	8200	6.9	202	14.6	442	2130
A2	0.7	4.5	0.3067	0.0575	11.2	7790	7.8	-276	15.2	476	2420
A3	0.5	3.1	0.2255	0.0556	10.3	7910	7.6	-242	14.7	419	2100
A4	0.2	2.4	0.1308	0.0377	8.6	7300	6.9	342	11.6	458	2140
B1	1.4	5.6	0.3577	0.0664	5.4	8900	7.59	256	15.1	490	1900
B2	1.1	4.6	0.2548	0.0221	6.48	8300	6.86	-220	13.5	480	1980
В3	0.7	4.5	0.2093	0.0043	7.1	7800	6.7	-238	11.9	452	2340
B4	0.2	2.7	0.0559	< 0.0014	6.41	8080	7.34	128	10.9	460	1820
C1	1.3	5.8	0.4600	0.0454	6.4	8070	6.6	262	14.6	398	1640
C2	0.9	4.8	0.3208	0.0364	6.01	7810	7.9	-244	13.1	431	1540
C3	0.6	2.6	0.2463	0.0193	5.7	7780	6.86	-186	12.4	428	1960
C4	0.2	1.4	0.1144	0.0335	7.13	7840	6.12	262	12	450	2000
D1	1.1	5.9	0.1846	0.0283	6.3	7730	6.96	245	11.5	394	1740
D2	0.8	3.7	0.1537	0.0329	6.12	7850	7.8	-200	11.2	432	1880
D3	0.6	1.7	0.0701	< 0.0014	7.4	7890	6.63	-47	12.1	439	1180
D4	0.3	1.6	0.0371	< 0.0014	8.7	8000	6.87	184	11.7	427	1000

nd- not determined Cd bdl (0.000025 mg/L), Se bdl (0.0001 mg/L). a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 79.** COC concentrations and explanatory parameters for Sampling Period 11 (2/7/13)

	Performance Parameters						Expla	natory Par	ameters		
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)		DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.5	5.4	0.5258		nd	6830	nd	nd	nd	514	1980
$A1^a$	1.2	5.2	0.5082		9.6	7080	7.9	202	14.6	452	1260
A2	2.6	4.2	0.3580		13.2	7200	7.8	-226	15.2	490	1260
A3	1.2	4.2	0.2145		12.3	7270	7.6	-227	14.7	422	1980
A4	0.6	2.7	0.2035		9.6	7190	6.9	344	11.6	462	1620
B1	1.9	4.6	0.6010		4.4	7310	6.59	282	15.1	472	1260
B2	1.8	4.4	0.6834		5.48	7600	6.86	-229	13.5	470	1800
В3	1.7	3.9	0.3252		6.94	7240	7.7	-189	11.9	450	900
B4	1.3	2.7	0.2382		6.41	7400	7.34	120	10.9	484	900
C1	1.4	5.2	0.5442		6.4	8200	6.6	137	14.6	466	1980
C2	1.5	4.7	0.3066		6.78	8020	6.9	-195	13.1	470	2340
C3	1.2	4.1	0.2327		5.7	7640	6.86	-200	12.4	454	1620
C4	1	2.9	0.1610		8.13	7180	7.67	260	12	511	2160
D1	1.3	5.2	0.5377		6.3	8600	6.46	185	11.5	470	1620
D2	1.2	4.9	0.3413		6.12	7200	6.84	-212	11.2	460	1800
D3	0.8	4.7	0.2671		7.4	7330	6.63	-158	12.1	453	2160
D4	0.4	3.3	0.1473		7.7	7260	6.8	139	11.7	497	1440

nd- not determined

Cd bdl (0.000025 mg/L), Mn bdl (0.0014 mg/L), Se bdl (0.0002 mg/L). a Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 80.** COC concentrations and explanatory parameters for Sampling Period 12 (2/21/13)

	Performa	ance Paramet	ters		F	Explanato	ry Parame	eters		
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	2	42.1	0.5589	nd	5980	nd	nd	nd	453	1990
$A1^a$	1.5	34.6	0.4719	8.1	6120	7.6	251	12.5	441	1800
A2	0.9	8.9	0.4007	9.8	6310	7.77	-230	13.6	429	1980
A3	0.8	5.7	0.4645	11.3	6470	7.58	-211	12.8	410	1980
A4	1.2	2.4	0.3387	9.6	6200	7.91	341	10.2	394	1800
B1	2.5	38.9	0.692	6.1	6920	6.96	300	11.3	432	1800
B2	1.5	24.4	0.3675	7.48	7100	7.2	-249	9.9	421	1800
В3	1.5	7.8	0.3333	8.71	6650	7.82	-169	10.4	420	1800
B4	1.3	2.1	0.324	6.2	6810	7.1	220	10.7	446	1800
C1	2.3	40.9	0.391	7.4	6420	6.94	187	12.6	444	1800
C2	1.6	32.8	0.3843	7.58	6130	7.48	-205	10.1	440	1800
C3	1.4	13.2	0.2953	6.1	6880	6.89	-240	11.4	462	1980
C4	1.2	2.4	0.2439	10.3	7120	7.31	278	11.5	450	1800
D1	1.9	40.8	0.3583	8.2	6200	6.22	235	11.8	445	1980
D2	1.4	36.4	0.3474	7.75	6510	7.15	-232	10.2	448	1800
D3	0.7	21.2	0.2851	7.4	6600	6.74	-198	9.1	458	1800
D4	0.8	3.9	0.1852	6.93	6830	6.98	163	10.7	426	1800

nd- not determined

Cd bdl (0.000025 mg/L), Mn bdl (0.0014 mg/L), Se bdl (0.0002 mg/L).

<sup>&</sup>lt;sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 81.** COC concentrations and explanatory parameters for Sampling Period 13 (3/14/13)

		Perform	nance Param	neters			Exp	olanatory l	Parameters	S	
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.5	7.5	0.5386	0.0659	nd	7810	nd	nd	nd	508	1880
$A1^{a}$	1.3	7.3	0.525	0.0864	8.6	8120	7.23	260	12.7	460	1260
A2	0.7	7.2	0.5714	0.0761	9.4	8440	8.21	-211	11.6	490	1260
A3	0.2	6.1	0.5044	0.0634	10.3	7900	7.89	-220	9.8	422	1980
A4	0.1	2.9	0.3061	0.0448	9.7	7630	7.7	321	10.2	461	1620
B1	1.6	6.9	0.4652	0.0771	7.2	7760	6.9	320	9.3	472	1260
B2	1.2	5.9	0.4336	0.0581	6.98	8030	7.29	-219	9.9	470	1800
В3	0.9	4.6	0.416	0.0612	8.25	7860	7.35	-178	10.5	438	900
B4	0.4	2.1	0.3229	0.0431	6.29	8290	7.27	223	9.1	484	900
C1	1.3	7.4	0.5251	0.0713	7.8	7860	6.99	198	11.6	466	1980
C2	1	6.6	0.4447	0.0702	8.78	7740	8	-180	9.4	472	2340
C3	0.7	5.6	0.3785	0.0528	6.61	7700	7.12	-232	10.4	449	1620
C4	0.2	3.1	0.2607	0.0382	9.64	8080	7.3	196	10.5	498	1860
D1	1.3	6.9	0.4122	0.0791	8.4	7530	6.67	220	9.8	470	1620
D2	0.8	6.5	0.4074	0.0671	7.65	7140	7.33	-255	10.8	460	1800
D3	0.2	6.4	0.3828	0.0365	7.3	7890	6.8	-209	10.1	453	1960
D4	0.1	3.9	0.2479	0.0279	5.9	7990	7.2	180	10	483	1640

nd- not determined

Cd bdl (0.000025 mg/L), Se bdl (0.0002 mg/L).

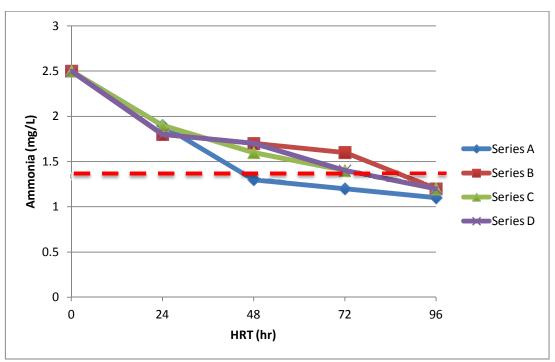
<sup>a</sup> Sampling location format is series (A), cell (1), etc., (Figure 1).

**Table 82.** COC concentrations and explanatory parameters for Sampling Period 14 (3/28/13)

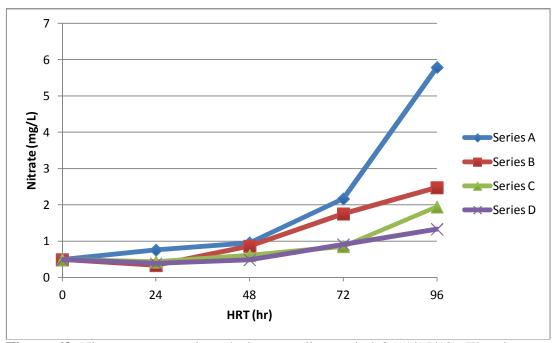
		Perform	nance Param	neters			Exp	olanatory l	Parameters	S	
Sampling Location	Ammonia (mg/L)	Barium (mg/L)	Iron (mg/L)	Manganese (mg/L)	DO (mg/L)	Cond. (µS/cm)	pH (S.U.)	Redox (mV)	Temp (°C)	Alkalinity (mg/L as CaCO <sub>3</sub> )	Hardness (mg/L as CaCO <sub>3</sub> )
Inflow	1.8	7.7	0.4390	0.0771	nd	8140	nd	nd	nd	430	1160
$A1^a$	1.5	7.2	0.4688	0.1090	7.62	8250	7.5	276	13.7	450	1180
A2	0.7	6.1	0.4114	0.0789	9.41	8290	7.82	-280	12.6	475	1200
A3	0.3	5.3	0.4129	0.0732	9.88	8390	7.59	-254	11.3	366	1440
A4	0.1	2.1	0.3704	0.0509	8.3	8470	7.62	335	11.6	416	1440
B1	1	7.1	0.4141	0.1799	7.58	8250	6.87	315	10.3	422	1440
B2	0.9	5.6	0.4747	0.0785	6.6	8340	7.88	-270	11.7	418	1260
В3	0.2	4.3	0.4113	0.0735	7.25	8430	7.1	-161	12	422	1260
B4	< 0.03	1.9	0.3386	0.0338	7.11	8410	6.79	240	11.5	402	1440
C1	1.4	7.2	0.451	0.0790	7.6	8290	6.9	222	11.6	433	1260
C2	0.7	6.1	0.4532	0.0638	8.9	8350	7.8	-183	10.4	450	1440
C3	0.2	4.7	0.3064	0.0442	5.61	8380	7.45	-262	11.2	362	1420
C4	< 0.03	2.5	0.2397	0.0215	7.64	8390	7.67	176	11.5	340	900
D1	1.1	7.2	0.4492	0.0646	8.23	8310	6.82	280	11.8	368	1260
D2	0.7	6.4	0.4167	0.0417	7.6	8370	7.2	-238	10.8	374	1080
D3	0.1	5.9	0.3667	0.0209	6.3	8440	7.34	-212	11.1	312	900
<u>D4</u>	< 0.03	3.9	0.2189	0.0186	6.2	8420	7.18	212	11.3	280	1020

nd- not determined
Cd bdl (0.00025 mg/L), Se bdl (0.0002 mg/L).

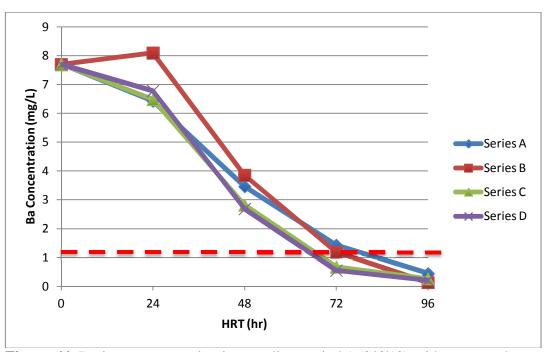
a Sampling location format is series (A), cell (1), etc., (Figure 1).



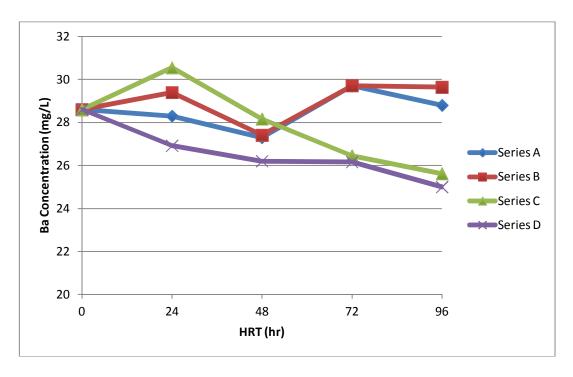
**Figure 62.** Ammonia concentration during sampling period 6 (11/15/12) with a targeted performance goal of 1.4 mg/L (red dashed line).



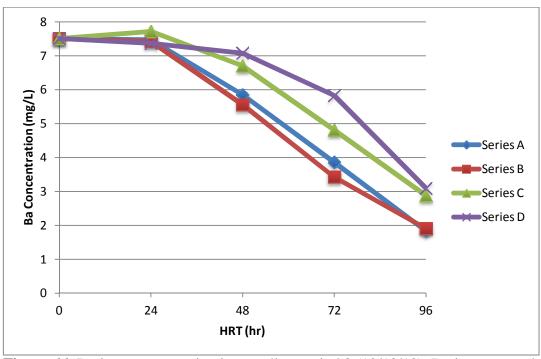
**Figure 63.** Nitrate concentrations during sampling period 6 (11/15/12). The nitrate concentrations measured in the outflow were greater than what would be produced if all the ammonia in the inflow were converted to nitrate, indicating that ammonia in the inflow was removed as well as additional ammonia produced in the system.



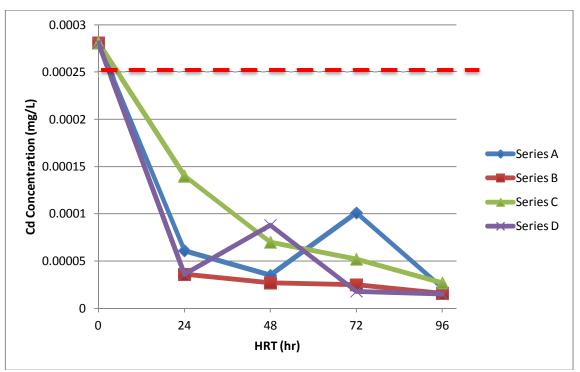
**Figure 64.** Barium concentration in sampling period 1 (9/6/12) with a targeted performance goal of 1.0 mg Ba/L (red dashed line).



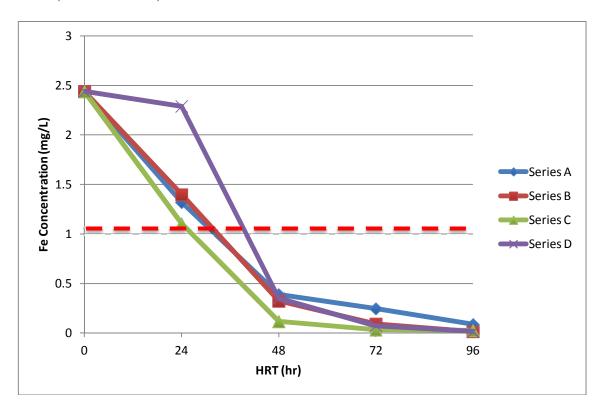
**Figure 65.** Barium concentration in sampling period 6 (11/15/12). Lack of decrease in Ba concentration is attributed to an insufficient amount of sulfate available, which was confirmed by measurement of sulfate concentration. Gypsum was added as a source of sulfate to all wetland cells on 11/29/12.



**Figure 66.** Barium concentration in sampling period 8 (12/13/12). Barium removal increased after the addition of gypsum to wetland cells on 11/29/12.



**Figure 67.** Cd concentration during sampling period 2 (9/20/12) with a goal of 0.00025 mg Cd/L (red dashed line).



**Figure 68.** Fe concentration during sampling period 4 (10/18/12) with a goal of 1.0 mg Fe/L (red dashed line).

**Table 83.** Ammonia concentrations measured in CWTS inflow and outflows, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the outflow (4-day HRT).

			Ammonia	
	<del>-</del>	Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
	Inflow	1.3		
ing 11	Series A outflow	2.3	nr	nr
Sampling Period 1	Series B outflow	0.6	53.8	0.193
Sampling Period 1	Series C outflow	0.3	76.9	0.367
	Series D outflow	0.3	76.9	0.367
-0	Inflow	0.1		
ing d 2	Series A outflow	0.1	0.0	0.000
Sampling Period 2	Series B outflow	< 0.03	>70.0	>0.301
	Series C outflow	< 0.03	>70.0	>0.301
	Series D outflow	< 0.03	>70.0	>0.301
<b>5</b> 0	Inflow	0.5		
ing d 3	Series A outflow	0.1	80.0	0.402
Sampling Period 3	Series B outflow	< 0.03	>94.0	>0.703
	Series C outflow	< 0.03	>94.0	>0.703
	Series D outflow	< 0.03	>94.0	>0.703
<b>5</b> 0	Inflow	3.3		
Sampling Period 4	Series A outflow	1.4	57.6	0.214
npl rio	Series B outflow	1.3	60.6	0.233
Sar Pe	Series C outflow	0.9	72.7	0.325
	Series D outflow	1.3	60.6	0.233
-0	Inflow	2.3		
Sampling Period 5	Series A outflow	0.1	95.7	0.784
Sampling Period 5	Series B outflow	1	56.5	0.208
Sar Pe	Series C outflow	0.9	60.9	0.235
	Series D outflow	0.9	60.9	0.235
50	Inflow	2.5		
ling d 6	Series A outflow	1.1	56.0	0.205
npli riod	Series B outflow	1.2	52.0	0.183
San Per	Series C outflow	1.2	52.0	0.183
	Series D outflow	1.2	52.0	0.183
50	Inflow	2.7		
ling d 7	Series A outflow	0.8	70.4	0.304
mp] rio	Series B outflow	0.9	66.7	0.275
San Per	Series C outflow	0.8	70.4	0.304
	Series D outflow	0.8	70.4	0.304

Table 83. (continued)

			Ammonia	
	_	Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
<b>-</b> 0	Inflow	2.9		
Sampling Period 8	Series A outflow	0.4	86.2	0.495
npl rio	Series B outflow	0.5	82.8	0.439
Sampling Period 8	Series C outflow	0.2	93.1	0.669
	Series D outflow	0.3	89.7	0.567
	Inflow	1.6		
ing 19	Series A outflow	0.1	93.8	0.693
Sampling Period 9	Series B outflow	0.3	81.3	0.418
	Series C outflow	0.1	93.8	0.693
	Series D outflow	0.2	87.5	0.520
LO	Inflow	1.7		
Sampling Period 10	Series A outflow	0.2	88.2	0.535
ldu jod	Series B outflow	0.2	88.2	0.535
Sar	Series C outflow	0.2	88.2	0.535
<b>5</b> , <b>1</b>	Series D outflow	0.3	82.4	0.434
	Inflow	1.5		
Sampling Period 11	Series A outflow	0.6	60.0	0.229
Sampli: Period	Series B outflow	1.3	13.3	0.036
San Per	Series C outflow	1	33.3	0.101
• 2	Series D outflow	0.4	73.3	0.330
	Inflow	2		
Sampling Period 12	Series A outflow	1.2	40.0	0.128
Samplir Period	Series B outflow	1.3	35.0	0.108
Sar Per	Series C outflow	1.2	40.0	0.128
-, -	Series D outflow	0.8	60.0	0.229
<b>50</b>	Inflow	1.5		
ing 13	Series A outflow	0.1	93.3	0.677
ildu iod	Series B outflow	0.4	73.3	0.330
San Peri	Series C outflow	0.2	86.7	0.504
	Series D outflow	0.1	93.3	0.677
	Inflow	1.8		
ing 14	Series A outflow	0.1	94.4	0.723
ldu iod	Series B outflow	< 0.03	>98.3	>1.024
Sampling Period 14	Series C outflow	< 0.03	>98.3	>1.024
	Series D outflow	< 0.03	>98.3	>1.024

**Table 84.** Barium concentrations measured in CWTS inflow and outflows, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the outflow (4-day HRT).

			Barium	
		Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
	Inflow	7.7		
ing 1.1	Series A outflow	0.45	94.2	0.710
Sampling Period 1	Series B outflow	0.13	98.3	1.020
Sar Pe	Series C outflow	0.26	96.6	0.847
	Series D outflow	0.21	97.3	0.900
	Inflow	6.1		
ng 2	Series A outflow	0.54	91.1	0.606
Sampling Period 2	Series B outflow	0.43	93.0	0.663
	Series C outflow	0.31	94.9	0.745
	Series D outflow	0.24	96.1	0.809
	Inflow	9.3		
Sampling Period 3	Series A outflow	0.28	97.0	0.876
Sampling Period 3	Series B outflow	0.17	98.2	1.000
San Per	Series C outflow	0.82	91.2	0.607
<b>0</b> 1 · ·	Series D outflow	0.39	95.8	0.793
	Inflow	17		
Sampling Period 4	Series A outflow	1.2	92.9	0.663
Sampling Period 4	Series B outflow	0.91	94.6	0.732
San Per	Series C outflow	0.59	96.5	0.840
• •	Series D outflow	0.42	97.5	0.925
	Inflow	13.2		
ing 15	Series A outflow	1.4	89.4	0.561
Sampling Period 5	Series B outflow	1.1	91.7	0.621
Sar Pe	Series C outflow	0.9	93.2	0.671
- 2	Series D outflow	0.61	95.4	0.769
<b>-</b> 0	Inflow	28.6		
ing d 6	Series A outflow	28.8	nr	nr
Samplii Period	Series B outflow	29.6	nr	nr
Sampli Period	Series C outflow	25.6	10.5	0.028
	Series D outflow	25	12.6	0.034
<b>5</b> 0	Inflow	12.6		
ling d 7	Series A outflow	14.6	nr	nr
mp] rio	Series B outflow	13.9	nr	nr
= =	Series C outflow	15.5	nr	nr
	Series D outflow	14	nr	nr

 Table 84. (continued)

	,		Barium	
	<del>-</del>	Conc. (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
	Inflow	7.5		
Sampling Period 8	Series A outflow	1.8	76.0	0.357
npl rioc	Series B outflow	1.9	74.7	0.343
Sar Pe	Series C outflow	2.9	61.3	0.238
	Series D outflow	3.1	58.7	0.221
	Inflow	7.4		
ing d 9	Series A outflow	1.1	85.1	0.477
Sampling Period 9	Series B outflow	1.5	79.7	0.399
	Series C outflow	3.1	58.1	0.218
	Series D outflow	3.4	54.1	0.194
p0 C	Inflow	6.2		
Sampling Period 10	Series A outflow	2.4	61.3	0.237
mp rio	Series B outflow	2.7	56.5	0.208
Sa Pe	Series C outflow	1.4	77.4	0.372
	Series D outflow	1.6	74.2	0.339
<u>-</u> a	Inflow	5.4		
Sampling Period 11	Series A outflow	2.7	50.0	0.173
Samplii Period	Series B outflow	2.7	50.0	0.173
Si Pe	Series C outflow	2.9	46.3	0.155
	Series D outflow	3.3	38.9	0.123
₩ 2	Inflow	42.1		
Sampling Period 12	Series A outflow	2.4	94.3	0.716
amp eric	Series B outflow	2.1	95.0	0.750
S. P.	Series C outflow	2.4	94.3	0.716
	Series D outflow	3.9	90.7	0.595
13 13	Inflow	7.5		
Sampling Period 13	Series A outflow	2.9	61.3	0.238
am) eric	Series B outflow	2.1	72.0	0.318
N G	Series C outflow	3.1	58.7	0.221
	Series D outflow	3.9	48.0	0.163
$\mathfrak{g}_4$	Inflow	7.7		
plin d 1	Series A outflow	2.1	72.7	0.325
San Per	Series B outflow	1.9	75.3	0.350
	Series C outflow	2.5	67.5	0.281
	Series D outflow	3.9	49.4	0.170

**Table 85.** Cadmium concentrations measured in CWTS inflow and outflows, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the outflow (4-day HRT). Cadmium concentrations were above the WQC in only the first two sampling periods.

			Cadmium	
	•	Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
	Inflow	0.000108		
ing 11	Series A outflow	< 0.000025	>76.9	>0.366
Sampling Period 1	Series B outflow	0.000034	68.5	0.289
Saı Pe	Series C outflow	0.00003	72.2	0.320
	Series D outflow	0.000107	0.9	0.002
	Inflow	0.000281		
ing 12	Series A outflow	< 0.000025	>91.1	>0.605
Sampling Period 2	Series B outflow	< 0.000025	>91.1	>0.605
Sar Per	Series C outflow	0.000027	90.4	0.586
-	Series D outflow	< 0.000025	>91.1	>0.605

**Table 86.** Iron concentrations measured in CWTS inflow and outflows, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the outflow (4-day HRT). Sampling period 3 is not included due to concentration of iron less than the WQC in inflow and outflow.

			Iron	
		Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
	Inflow	0.2758		
Sampling Period 1	Series A outflow	0.6978	nr	nr
Sampli Period	Series B outflow	0.3139	nr	nr
Sar Pe	Series C outflow	0.0846	69.3	0.295
	Series D outflow	0.1755	36.4	0.113
	Inflow	0.1006		
ing 12	Series A outflow	0.2032	nr	nr
Sampling Period 2	Series B outflow	0.0138	86.3	0.496
Sampling Period 2	Series C outflow	0.0093	90.8	0.596
	Series D outflow	0.0162	83.9	0.456
	Inflow	2.44		
Sampling Period 4	Series A outflow	0.088	96.4	0.831
ampling Period 4	Series B outflow	0.016	99.3	1.257
San Per	Series C outflow	0.028	98.9	1.117
<b>J</b> 1	Series D outflow	0.017	99.3	1.242
	Inflow	1.48		
ing 15	Series A outflow	0.047	96.8	0.862
Sampling Period 5	Series B outflow	0.02	98.6	1.076
Sampling Period 5	Series C outflow	0.331	77.6	0.374
<b>J</b> 1	Series D outflow	0.104	93.0	0.664
	Inflow	1.524		
ing 16	Series A outflow	0.119	92.2	0.637
ldr joi:	Series B outflow	0.07	95.4	0.770
Sampling Period 6	Series C outflow	0.123	91.9	0.629
<b>J</b> 1	Series D outflow	0.144	90.6	0.590
	Inflow	0.262		
ing 17	Series A outflow	0.109	58.4	0.219
ampli Period	Series B outflow	0.055	79.0	0.390
Sampling Period 7	Series C outflow	0.089	66.0	0.270
	Series D outflow	0.013	95.0	0.751

 Table 86. (continued)

non	Iron	
Conc. Remov		
(mg/L) (%)	$(d^{-1})$	
Inflow 0.7842		
$\stackrel{60}{=} \stackrel{\infty}{=} \stackrel{\text{Series A outflow}}{=} 0.2168 \qquad 72.4$	0.321	
Series A outflow 0.2168 72.4 $\odot$ Series B outflow 0.0990 87.4 Series C outflow 0.1481 81.1	0.517	
$\stackrel{\overline{\overline{Z}}}{\sim} \stackrel{\omega}{\sim}$ Series C outflow 0.1481 81.1	0.417	
Series D outflow 0.0554 92.9	0.662	
Inflow 0.1316		
Series A outflow 0.0912 30.7  Series B outflow 0.0444 66.2  Series C outflow 0.0675 48.7	0.092	
Series B outflow 0.0444 66.2	0.272	
$\stackrel{\mathbf{R}}{\sim} \stackrel{\mathbf{G}}{\sim}$ Series C outflow 0.0675 48.7	0.167	
Series D outflow 0.0060 95.4	0.771	
Inflow 0.3554		
Series A outflow 0.3554  Series A outflow 0.1308 63.2  Series B outflow 0.0559 84.2  Series C outflow 0.1144 67.8	0.250	
$\stackrel{\circ}{\mathbf{E}} \stackrel{\circ}{\overset{\circ}{\mathbf{E}}}$ Series B outflow 0.0559 84.2	0.462	
$\overset{\square}{\sim} \overset{\square}{\sim}$ Series C outflow 0.1144 67.8	0.283	
Series D outflow 0.0371 89.6	0.565	
Inflow 0.5258		
Series A outflow 0.2035 61.3  Series B outflow 0.2382 54.7  Series C outflow 0.161 69.4	0.237	
Series B outflow 0.2382 54.7 Series C outflow 0.161 69.4	0.198	
	0.296	
Series D outflow 0.1473 72.0	0.318	
Inflow 0.5589		
Series A outflow 0.3387 39.4	0.125	
Inflow   0.5589	0.136	
Series C outflow 0.2439 56.4 Series D outflow 0.1852 66.9	0.207 0.276	
	0.270	
Inflow 0.5386 Series A outflow 0.3061 43.2	 0.141	
Series A outflow 0.3061 43.2  Series B outflow 0.3229 40.0  Series C outflow 0.2607 51.6	0.141 0.128	
Series C outflow $0.3229$ $40.0$ $0.3229$ $51.6$	0.128	
Series D outflow 0.2479 54.0	0.194	
Inflow 0.439		
$\begin{array}{ccc}  & & & & & & & & & & & \\  & & & & & & &$	0.042	
Series B outflow 0.3386 22.9	0.065	
Series A outflow 0.3704 15.6 Series B outflow 0.3386 22.9 Series C outflow 0.2397 45.4	0.151	
Series D outflow 0.2189 50.1	0.174	

**Table 87.** Manganese concentrations measured in CWTS inflow and outflows, removal efficiencies (%), and removal rate coefficients (d<sup>-1</sup>). Removal efficiencies and removal rate coefficients were calculated using measured concentrations in the outflow (4-day HRT). Manganese concentrations were below detection limit during sampling periods 3-7, and are omitted from the table.

		Manganese		
	<del>-</del>	Conc.	Removal	Rate Coef.
		(mg/L)	(%)	$(d^{-1})$
Sampling Period 1	Inflow	0.1244		
	Series A outflow	2.272	nr	nr
	Series B outflow	2.658	nr	nr
	Series C outflow	0.6731	nr	nr
	Series D outflow	1.019	nr	nr
	Inflow	0.0200		
ling d 2	Series A outflow	0.0372	nr	nr
Sampling Period 2	Series B outflow	0.0028	86.0	0.492
	Series C outflow	0.0146	27.0	0.079
	Series D outflow	0.0541	nr	nr
	Inflow	0.169		
ling d 4	Series A outflow	< 0.0014	>99.2	>1.198
Sampling Period 4	Series B outflow	< 0.0014	>99.2	>1.198
	Series C outflow	< 0.0014	>99.2	>1.198
	Series D outflow	< 0.0014	>99.2	>1.198
- 0	Inflow	0.0299		
ling d 8	Series A outflow	< 0.0014	>95.3	>0.766
Sampling Period 8	Series B outflow	< 0.0014	>95.3	>0.766
Sa Pe	Series C outflow	< 0.0014	>95.3	>0.766
	Series D outflow	< 0.0014	>95.3	>0.766
50	Inflow	0.0341		
Sampling Period 9	Series A outflow	0.0274	19.7	0.055
mp erio	Series B outflow	0.0079	76.8	0.366
Sa. Pe	Series C outflow	< 0.0014	>95.9	>0.798
	Series D outflow	0.0111	67.4	0.281
	Inflow	0.0385		
ling 110	Series A outflow	0.0377	2.3	0.006
Sampling Period 10	Series B outflow	< 0.0014	>96.4	>0.829
Sa Pe	Series C outflow	0.0335	13.0	0.035
	Series D outflow	< 0.0014	>96.4	>0.829

 Table 87. (continued)

		Manganese		
	•	Conc. (mg/L)	Removal (%)	Rate Coef. (d <sup>-1</sup> )
Sampling Period 13	Inflow	0.0659		
	Series A outflow	0.0448	32.0	0.096
	Series B outflow	0.0431	34.6	0.106
	Series C outflow	0.0382	42.0	0.136
	Series D outflow	0.0279	57.7	0.215
	Inflow	0.0771		
ling 1 14	Series A outflow	0.0509	34.0	0.104
Sampling Period 14	Series B outflow	0.0338	56.1	0.206
	Series C outflow	0.0215	72.1	0.319
	Series D outflow	0.0186	75.9	0.355



**Figure 69.** Demonstration wetland treatment system on 9/7/12, 14 weeks after construction.

# Apply Results to Water Management and Technology Replication (Task 9)

Application to Various Geologic Basins and Locations

A strategic literature review and analysis of field-collected water samples completed during Task 2 revealed that produced water composition varies from basin to basin and even from well to well within the same basin. Therefore, targeted constituents considered for treatment during the pilot-scale and demonstration phases of the project (Phases II and III) consisted of constituents that could potentially limit beneficial use of produced water at many locations. These targeted constituents, identified as COCs during Task 3, were addressed for treatment by specifically designed pilot-scale CWTSs in Phase II and included dissolved solids, metals, metalloids, organic compounds, and nitrogenous compounds.

Results from Phase II demonstrated successful treatment of the targeted COCs found in produced water including Fe, Mn, Zn, Ni, Cd, Cu, Se, ammonia, nitrate, oil and grease, and low molecular weight organics to meet beneficial use criteria. Using the design principles established during Phase II, an on-site demonstration CWTS was constructed to remove Ba, Cd, Fe, Mn, and ammonia from coal-bed methane produced water generated from the Black Warrior basin in Alabama. The demonstration CWTS was successful for treating the COCs, thus renovating the produced water to meet stringent discharge limits and guideline criteria for crop irrigation and livestock watering. Successful transfer of CWTS design principles from pilot-scale to demonstration supports the applicability of experimental results from this study for design of future CWTSs.

CWTSs can be tailored to suit a variety of needs presented in areas that have produced waters of differing geologic origins. For instance, coal-bed methane water from the Powder River basin containing elevated concentrations of metals (e.g. Fe, Cu, Zn, etc.) and Se may be treated with a modular CWTS based on the design principles established in Phase II. Oxidizing CWTS cells can be used to precipitate and sequester Fe as insoluble iron(III) oxyhydroxide. Reducing CWTS cells can be assembled to promote dissimilatory sulfate reduction and sulfide precipitation of Cu and Zn as well as microbial reduction and precipitation of elemental Se. Removal rate coefficients measured in Phase II can be used to predict required treatment capacity for a given hydraulic loading that is site- or situation-dependent.

Additional treatment technologies can be incorporated with CWTSs (forming hybrid systems) to expand treatability and beneficial use options. As demonstrated in Phase II, oil-water separators can be used to pretreat produced water prior to its introduction to CWTSs to remove non-aqueous phase organic compounds and oil. The separated oil can then be collected and sold to recover operational and maintenance costs. Reverse osmosis can be used to remove dissolved salts from brackish and saline produced waters so they may be suitable for treatment by CWTSs and subsequently available for salinity-sensitive beneficial use options such as crop irrigation. Hypersaline produced water from some geologic formations such as the Marcellus Shale and Mississippi Lime may not be suitable for treatment due to the current limitations of desalinization technologies, therefore requiring disposal by deep-well injection.

#### Beneficial Use Selection

Two different approaches can be used as the basis for designing CWTSs: a water-specific approach or a site-specific approach. With a water-specific approach, a CWTS can be designed to target all treatable COCs determined during characterization of produced water samples. For example, the pilot-scale CWTSs used in Phase II and demonstration CWTS used in Phase III were designed to meet the most stringent use criteria for each targeted COC. By targeting the most stringent criteria for each COC, produced water can be treated to permit beneficial use for as many different applications as possible. For instance, treated produced water from the Black Warrior basin in Alabama may be used for crop irrigation, livestock watering, industrial process water, power-plant cooling water, aquifer recharge, habitat restoration, and stream augmentation. A water-specific approach can be appropriate when there is a local demand for water for a variety of uses, when the beneficial use is unknown or may change through time, and/or when the produced water contains COCs that are not effectively removed by CWTSs (e.g. total dissolved solids). Because beneficial use demands are subject to change as a result of several factors including changes in seasons and changes in local land use, treating produced water to meet a multitude of beneficial use options helps ensure that the treated water can be used to supplement both current and future needs.

With a site-specific approach, a CWTS can be designed to treat produced water to meet beneficial use criteria for specific local needs. For example, treating produced water generated in urban areas to meet use criteria for crop irrigation and livestock watering in urban areas is not practical. Instead, the produced water may be treated to meet different use criteria more appropriate for local needs such as industrial process water. A site-specific approach can be more applicable in areas where local needs are not predicted to change over the lifetime of the CWTS.

## CWTS Treatment Cost Comparison

The most feasible approach for treating site-specific produced water depends on both capital and operational costs and the ability of the treatment system to meet guideline criteria for beneficial use. Costs of CWTS construction, operation, and maintenance can vary widely depending on the location. These costs are affected primarily by treatment objectives, local terrain, and bottomliner requirements. Mooney and Murray Gulde (2008) found that costs for constructing CWTSs are commonly 50 to 70% less than for conventional treatment systems, with CWTS operation and maintenance costs between \$0.25 and \$0.80 per 1000 gallons of water treated. At a location in USDOE's Savannah River Site, where nearly 1 million gallons of water per day require treatment, estimated costs for conventional treatment systems for metal removal (ion exchange, chemical precipitation, source removal, etc.) proved to be too high for the water volume and treatment criteria (Nelson and Gladden, 2008). CWTSs were constructed as a low-cost alternative and resulted in operation and maintenance costs of less than \$50,000 per year, which is less than \$0.14 per 1000 gallons of water treated (Nelson and Gladden, 2008). Jackson and Myers (2003) provided cost estimates for disposal methods of produced water, and ALL Consulting (2012) provided information on treatment processes being utilized in the shale gas industry as part of a "Water Treatment Catalog" (Table 84). Cost estimates for different

Table 84. Produced water treatment and disposal methods and estimated costs.

	Estimated cost		
Method	(\$/bbl)	Limitation	Benefits
Surface discharge <sup>1</sup>	0.01-0.08	Stringent standards <sup>3</sup>	Livestock, wildlife, irrigation
Secondary oil recovery <sup>1</sup>	0.05-1.25	Infrastructure	Increase oil production
Shallow reinjection <sup>1</sup>	0.10-1.33	Energy and maintenance	Recharge aquifer if water quality is sufficient for drinking
Evaporation pits <sup>1</sup>	0.01-0.80	Sequestering from wildlife/soil contamination	Simple, passive management strategy
Commercial water hauling <sup>1</sup>	0.01-5.50	Distance, safety issues	-
Disposal wells <sup>1</sup>	0.05-2.65	Strict environmental regulations	-
Freeze-thaw evaporation <sup>1</sup>	2.65-5.00	Regional climate	Water/salt separation
Constructed wetland <sup>1</sup>	0.001-2.00	Land area, retention time requirement	Low cost than many technologies; efficient removal of dissolved and suspended contaminants; low energy requirement
Induced air flotation for de- oiling <sup>1</sup>	0.05	CAPEX	Simple operation; durable <sup>3</sup>
Anoxic/aerobic granular activated carbon <sup>1</sup>	0.083	CAPEX	Simple technology <sup>3</sup>
Thermal evaporation <sup>2</sup>	3.00-5.00	Pretreatment required; not effective above 80,000 mg/L TDS	Effective for PW with 40,000-80,000 mg/L TDS
Reverse Osmosis (RO) <sup>2</sup>	0.11	40 to 65% water recovery	Removes most ions
Electrodialysis <sup>2</sup>	0.15	TDS 4,000-15,000 mg/L	More resistant to membrane fouling than RO
Ion Exchange <sup>2</sup>	0.05-0.20	TDS must be <5,000 mg/L; may require pretreatment	Low energy requirement
UV Light <sup>2</sup>	0.60-0.75	May require use of biocides	Can disinfect PW
Ozone <sup>2</sup>	0.60-0.75	Complex chemistry; toxicity issues	Removes bacteria without producing waste byproduct

bbl = barrel of treated water (1 bbl = 42 gallons; Clark and Veil, 2009)

CAPEX: Capital expenses (includes cost for materials, labor, and interest required to construct and begin operation)

<sup>1</sup>(Jackson and Myers, 2003)

<sup>2</sup>(ALL Consulting, 2012)

<sup>3</sup>(Fakhrul-Razi et al., 2009)

treatment options can be refined by evaluating site-specific factors including water quality, site accessibility, cost of land, and cost of labor.

Produced Water Management Plan and Application to Treatment System Design

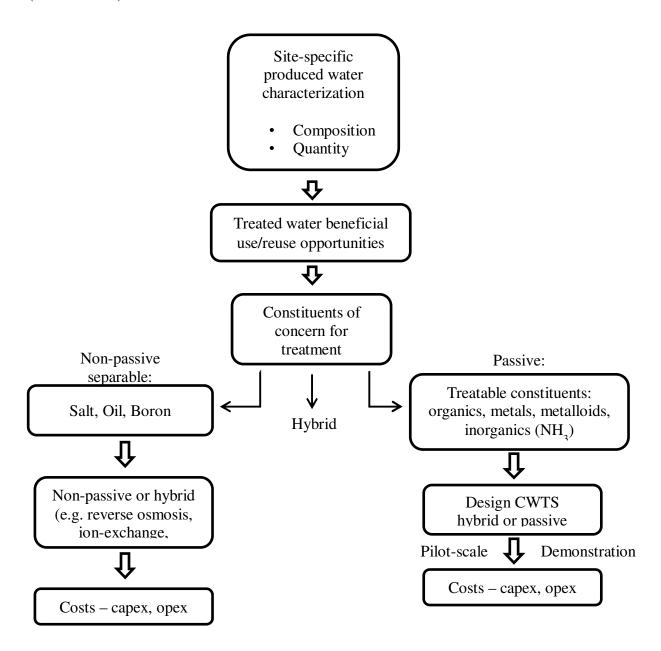
Management of produced water depends on several site-specific factors including produced water quantity and quality, local water needs, and treatment system costs (Figure 70). Constituents of concern limiting beneficial use can be targeted for treatment using a non-passive, passive, or hybrid approach. The non-passive approach can be used to treat COCs that may not be effectively removed with CWTSs (e.g. dissolved salts, boron, etc.) through the use of physical separation technologies (e.g. reverse osmosis, ion-exchange, filtration, etc.). The passive approach can be used to treat COCs that are effectively removed with CWTSs (e.g. metals, metalloids, oil and grease, nitrogenous compounds). A hybrid approach to designing CWTSs consisting of non-passive and passive approaches can be used to remove separable COCs including dissolved salts from produced water prior to introduction in wetland cells to expand treatability and beneficial use options and also lower capital and operational costs.

Because composition of produced waters typically varies from source to source (Tables 4-8), site-specific produced water characterization through sample collection and analysis is necessary to determine composition of the water to be treated. Constituents to be analyzed must include those that may hinder ability of the water to meet beneficial use criteria (Tables 9 and 10). After composition of produced water from the site is determined, COCs) can be identified by comparing constituent concentrations with beneficial use criteria that have been selected based on specific needs of the site location. Constituents that exceed the criteria concentrations are identified as COCs and will require treatment in order for the water to be suitable for use. With proper identification of COCs and design of CWTSs, waters ranging from fresh to hypersaline can be treated (e.g. Johnson et al., 2008b; Kanagy et al., 2008b).

A constructed wetland treatment system should be designed to promote specific biogeochemical pathways by which the COCs can be treated. Pilot-scale CWTS experiments performed during Phase II demonstrated successful design strategies for treating a variety of COCs including divalent metals, metalloids, oil and grease, and ammonia (Figures 2-5). Targeted pathways for treatment of COCs in the pilot-scale CWTS experiments included divalent metal sulfide precipitation through dissimilatory sulfate reduction, metal precipitation through oxidation, reduction of selenite to insoluble elemental selenium, aerobic biodegradation of oil, nitrification of ammonia to nitrate, denitrification of nitrate to nitrogen gas, separation of oil using an oil-water separator, and sorption of ammonia to zeolite.

CWTSs should be designed and built to promote specific environmental and geochemical conditions (Tables 20-23, 58, 61, and 62) in order for the targeted pathways to operate. For example, a CWTS designed to nitrify ammonia to nitrate must provide oxidizing conditions (DO > 2 mg/L), neutral to slightly basic pH (6.6-9.7), and sufficient alkalinity (>100 mg/L as CaCO<sub>3</sub>) in order for nitrification to occur. Pilot-scale experiments achieved

performance when the targeted environmental and geochemical conditions were met (Tables 26-50).



**Figure 70.** Produced water management plan, which is based in part on the methods used in Phases I-III and is intended for management of a site-specific produced water.

Design strategies used in the pilot-scale CWTSs and demonstration CWTS to promote specific conditions that can be applied to design of full-scale CWTSs include plant and soil selection, water-depth selection, addition of amendments, and hydraulic retention time (HRT). These strategies allow conditions within the CWTS to be modified to achieve ranges necessary for the preferred biogeochemical pathways. For example, results of this project show that an oxidizing CWTS can be constructed with *Typha latifolia*, sandy soil, and a water depth of 33 cm or less. A reducing CWTS can be constructed with *Schoenoplectus californicus*, organic-rich soil, and a water depth of 46 cm or greater to promote reduction reactions such as dissimilatory selenium reduction, AquaSmart<sup>TM</sup>, an organic-rich nutrient amendment containing yeast, was added to some of the pilot-scale CWTSs and to the demonstration system. Crushed oyster shells were added to some pilot-scale cells to increase alkalinity and raise pH. An oil-water separator can be included in a CWTS to decrease oil and grease concentrations (Table 64, Figures 51 and 52), and zeolite minerals can be added to increase sorption of ammonia (Figure 30).

In the case of renovating a produced water containing COCs that require different biogeochemical pathways for treatment, a full-scale CWTS can be designed with sequential cells that promote different conditions. For instance, the pilot-scale CWTS for post-RO produced water was designed to promote oxidizing conditions within the first cell for nitrification of ammonia, while the subsequent three cells were designed to promote reducing conditions for denitrification of nitrate (Figure 2). By separating a CWTS into multiple cells, the conditions within each cell can be modified for removal of a variety of COCs. In addition, a CWTS designed with multiple cells allows for convenient sample collection points so that removal rate coefficients can be calculated and performance of individual cells can be evaluated.

By rearranging the integrated first-order rate coefficient equation (Equation 11), removal rate coefficients determined from the pilot-scale CWTS experiments (Tables 51-52, 54-56, 66-67) and confirmed by the demonstration system can be used to predict HRTs required to treat COCs in full-scale CWTSs:

Required Treatment HRT 
$$(t) = \frac{-\ln([c]/[c_o])}{k}$$
 Eqn. 12

where  $[C_o]$  is the concentration (mg/L) of a COC in the inflow, [C] is the targeted treatment concentration (mg/L), k is the first-order rate coefficient (d<sup>-1</sup>), and t is the time (days) and corresponds to HRT of the system.

The HRT calculated using Equation 12 can then be used to determine the surface area or "footprint" of a full-size CWTS for a given inflow rate:

Wetland Surface Area (A) = 
$$\frac{Q \times t}{d}$$
 Eqn. 13

where A is the required surface area of the CWTS (m<sup>2</sup>), Q is the inflow rate (m<sup>3</sup>/d), t is the required HRT (days), and d is the water depth (m).

## Technology Transfer (Task 10)

The following is a list of journal papers and presentations for technology transfer of results from this project:

- Alley, B., Beebe, A., Rodgers, J. H., Jr., and Castle, J. W., "A Comparative Characterization of Produced Water from Conventional and Unconventional Fossil Fuel Resources," <u>American Association of Petroleum Geologists, Program and Abstracts</u>, April 2010.
- Alley, B., Beebe, A., Rodgers, J. H., Jr., and Castle, J. W., "Chemical and Physical Characterization of Produced Waters from Conventional and Unconventional Fossil Fuel Resources," <u>Chemosphere</u>, v. 85, p. 74-82. 2011.
- Alley, B., Beebe, D. A., Rodgers, J. H., Jr., and Castle, J. W., "Chemical and Physical Characterization of Produced Waters from Conventional and Unconventional Fossil Fuel Resources," <u>Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, April 2011.
- Alley, B. L., Rodgers, J. H., Jr., and Castle, J. W., "Performance of Constructed Wetland Treatment Systems Designed to Renovate Fresh Oilfield Produced Waters for Beneficial Uses," <u>Geological Society of America Abstracts with Programs</u>, v. 43, no. 5, p. 579, October 2011.
- Alley, B. L., Rodgers, J. H., Jr., and Castle, J. W., "Renovating Fresh Oilfield Produced Waters for Beneficial Uses: Managing Constructed Wetland Treatment Systems for Performance," <u>Society of Environmental Toxicology and Chemistry (SETAC) North America 32<sup>nd</sup> Annual Meeting Abstract Book, p. 380, November 2011.</u>
- Alley, B. L., Willis, B., Rodgers, J., Jr., and Castle, J. W., "Seasonal Performance of a Hybrid Pilot-Scale Constructed Wetland Treatment System for Simulated Fresh Oil Field-Produced Water," <u>Water, Air, & Soil Pollution</u>, v. 244. DOI 10.1007/s11270-013-1639-5. 2013.
- Alley, B. L., Willis, B., Rodgers, J. H., Jr., and Castle, J. W., "Water Depth and Treatment Performance of Free Water Surface Constructed Wetland Treatment Systems for Simulated Fresh Oilfield Produced Water," <u>Clemson University 20<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, p. 2, April 2012.
- Alley, B. L., Rodgers, J. H., Jr., and Castle, J. W., "Seasonal Performance of a Hybrid Pilot-Scale Constructed Wetland Treatment System for Simulated Fresh Oilfield Produced Water," <u>Clemson University 21<sup>st</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, p. 3, April 2013.
- Beebe, D. A., Castle, J. W., and Rodgers, J. H., Jr., "Clinoptilolite as a Dual Purpose Sorbent Carrier in Constructed Wetland Treatment Systems Designed to Remove Ammonia," <u>Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts</u> with Program, April 2011.

- Beebe, D. A., Song, Y., Castle, J. W., and Rodgers, J. H., Jr., "Pilot Study of Constructed Wetland Treatment Systems Designed for Removal of Ammonia in Oilfield Produced Water," <u>Geological Society of America Abstracts with Programs</u>, v. 43, no. 5, p. 580, October 2011.
- Beebe, D. A., Song, Y., Castle, J. W., and Rodgers, J. H., Jr., "Designing Constructed Wetland Treatments Systems for Ammonia in Produced Water," <u>Society of Environmental Toxicology and Chemistry (SETAC) North America 32<sup>nd</sup> Annual Meeting Abstract Book, p. 377, November 2011.</u>
- Beebe, D. A., Alley, B., Castle, J. W., and Rodgers, J. H., Jr., "Potential for Beneficial Use of Coal-Bed Methane Produced Water in Western Alabama to Augment Water Supplies during Intense Drought," <u>Geological Society of America Abstracts with Programs</u>, v. 44, no. 4, p. 66, April 2012.
- Beebe, D. A., Alley, B., Castle, J. W., and Rodgers, J. H., Jr., "Evaluation of Coal-Bed Methane Produced Water in Western Alabama for Use as a Water Resource during Drought," <u>Clemson University 20<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, p. 4, April 2012.
- Beebe, D. A., Castle, J. W., and Rodgers, J. H., Jr., "Experimental Studies of Evapotranspiration in Small Stand Wetlands," <u>Geological Society of America Abstracts with Programs</u>, v. 44, no. 7, p. 581, November 2012.
- Beebe, D. A., Castle, J. W., and Rodgers, J. H., Jr., "Effects of Evapotranspiration on Water Treatment Performance in Constructed Wetlands," <u>Clemson University 21<sup>st</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, p. 5, April 2013.
- Castle, J. W., Rodgers, J. H., Jr., Alley, B., Spacil, M., Beebe, A., Pardue, M., and Song, Y., "Biogeochemical Processes for Treating Oil and Gas Produced Waters Using Hybrid Constructed Wetland Treatment Systems," <u>American Association of Petroleum Geologists, Program and Abstracts</u>, April 2010.
- Castle, J. W., Rodgers, J. H., Jr., Spacil, M., Alley, B., Horner, J. E, and Pardue, M., "Pilot-Scale Constructed Wetland Treatment Systems for Oil & Gas Produced Waters," 17<sup>th</sup> Annual International Petroleum and Biofuels Environmental Conference Program and Abstracts, p. 47, September 2010.
- Castle, J. W., Rodgers, J. H., Jr., Spacil, M., Horner, J. E, Alley, B., and Pardue, M., "Pilot-Scale Constructed Wetland Systems for Treating Energy-Produced Waters," Ground Water Protection Council Annual Forum, Water & Energy in Changing Climates, Pittsburgh, PA, September 29, 2010.
- Castle, J. W., Rodgers, J. H., Jr., Spacil, M., Alley, B., and Pardue, M., "A Pilot-Scale Study to Apply Biogeochemical Processes of Natural Wetlands to Treating Impaired Waters Using Constructed Wetland Treatment Systems," <u>Geological Society of America Abstracts with Programs</u>, v. 42, no. 5, p. 640, November 2010.
- Castle, J. W., Rodgers, J. H., Jr., Spacil, M. M., and Ritter, C. L., "Treating Selenium

- in Energy-derived Produced Waters Using Constructed Wetland Treatment Systems," <u>Geological Society of America Abstracts with Programs</u>, v. 43, no. 5, p. 578, October 2011.
- Coffey, R. E., Castle, J. W., and Rodgers, J. H., Jr., "A Demonstration Constructed Wetland Treatment System for Unconventional Gas Produced Water," <u>Clemson University 21<sup>st</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, p. 10, April 2013.
- Jurinko, K. N., Ritter, C. L., Castle, J. W., and Rodgers, J. H., Jr., "Biogeochemical Process in a Pilot-Scale Constructed Wetland Treatment System Designed to Remove Metals from Produced Water," <u>Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, April 2011.
- Pardue, M. J., Castle, J. W., and Rodgers, J. H., Jr., "Evaluation of a Pilot-Scale Constructed Wetland System for Treatment of a Specific Oilfield Produced Water," Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program, April 2011.
- Pardue, M. J., Castle, J. W., and Rodgers, J. H., Jr., "A Biogeochemical Approach to Renovation of Energy-Derived Produced Water Using a Pilot-Scale Constructed Wetland Treatment System," <u>Geological Society of America Abstracts with Programs</u>, v. 43, no. 5, p. 580, October 2011.
- Pardue, M. J., Castle, J. W., Huddleston, G. M., and Rodgers, J. H., Jr., "Treatment of Oilfield Produced Water Using a Pilot-Scale Constructed Wetland Treatment System," <u>Clemson University 20<sup>th</sup> Annual Hydrogeology Symposium Abstracts</u> <u>with Program</u>, p. 33, April 2012.
- Ritter, C. L., Jurinko, K. N., Castle, J. W., and Rodgers, J. H., Jr., "Biogeochemical Processes in a Constructed Wetland Treatment System Designed for Removal of Selenium from Energy-Derived Water," <u>Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, April 2011.
- Rodgers, J. H., Jr., Castle, J. W., Horner, J. E, Spacil, M., Eggert, D., Alley, B., Beebe, A., Pham, M. P. T., Song, Y., Myers, J. E., Murray Gulde, C., Huddleston, M., and Mooney, D., "Constructed Wetland Treatment Systems for Renovation of Oilfield Produced Water for Beneficial Use," <u>Clemson University 17<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, April 2009.
- Rodgers, J. H., Jr., and Castle, J. W., "Characteristics of Produced Waters and Biogeochemical Processes for Effective Management Using Constructed Wetland Treatment Systems," Goldschmidt International Conference on Earth, Energy, and the Environment, Geochimica et Cosmochimica Acta, v. 74, issue 12, Supplement 1, p. A876, June 2010.
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- (SETAC) North America 32<sup>nd</sup> Annual Meeting Abstract Book, p. 376, November 2011.
- Spacil, M. M., Rodgers, J. H., Jr., Castle, J. W., and Chao, W. Y., "Performance of a Pilot-Scale Constructed Wetland Treatment System for Selenium, Arsenic, and Low Molecular Weight Organics in Simulated Fresh Produced Water," <u>Clemson University 19<sup>th</sup> Annual Hydrogeology Symposium Abstracts with Program</u>, April 2011.
- Spacil, M. M., Rodgers, J. H., Jr., Castle, J. W., and Chao, W. Y., "Performance of a Pilot-Scale Constructed Wetland Treatment System for Selenium, Arsenic, and Low Molecular Weight Organics in Simulated Fresh Produced Water," <u>Environmental Geosciences</u>, v. 18, p. 145-156. 2011.
- Spacil, M. M., Rodgers, J. H., Jr., Castle, J. W., and Chao, W. Y., "Treatment of Selenium in Produced Water Using a Pilot-scale Constructed Wetland Treatment System," <u>Society of Environmental Toxicology and Chemistry (SETAC) North America 33<sup>rd</sup> Annual Meeting Abstract Book, p. 273, November 2012.</u>

#### CONCLUSION

A wide range of chemical, physical, and risk characteristics was present in the produced waters investigated. Constituents of concern identified as requiring treatment in produced waters prior to reuse or discharge include dissolved salts, metals, metalloids, organics, and post-RO constituents. Dissolved salts, expressed as salinity, conductivity, or TDS in published reports and in data provided by industry, occur over a wide range in produced waters. Cations found in produced water commonly include sodium, calcium, magnesium, and potassium; anions present often include chloride, sulfate and bicarbonate. Reverse osmosis (RO) is sometimes used to reduce the concentration of cations and anions in produced waters. However, specific constituents such as ammonia can pass through RO membranes and require further treatment. Metals that are of concern in produced waters can include any or all of the following: iron, manganese, zinc, cadmium, cobalt, copper, lead, mercury, chromium, nickel, and zinc. Based on our analysis, other constituents of concern identified in produced waters include arsenic and selenium. Produced waters can contain a range of organic constituents from oil and grease to low molecular-weight aromatic hydrocarbons such as benzene, toluene, ethylbenzene, and xylene (BTEX).

Pilot-scale constructed wetland treatment systems were designed and constructed at Clemson University based on fundamental reactions and processes needed to treat (transfer and transform) targeted constituents in produced waters. Separate systems were constructed for representative constituents with specific water compositions selected based on results of the previous tasks. We investigated four waters comparing a range of characteristics or constituents of concern for treatment (metals, metalloids, oil and grease, and ammonia). The pilot-scale systems were designed to produce biogeochemical conditions that result in targeted treatment pathways for constituents that occur in produced waters. Systems were constructed so that the sequence of biogeochemical

conditions can be changed as needed to achieve the most effective treatment.

Performance measurements from the pilot-scale constructed wetland systems indicated effective treatment of targeted constituents in the four types of produced waters investigated. Data from the pilot-scale phase of the project were used to design a demonstration system. In collaboration with Chevron, a site for constructing the demonstration system was selected near Berry, Alabama. This site is within a producing coalbed methane field. The demonstration CWTS was designed and built to promote specific environmental and geochemical conditions in order for targeted treatment pathways to operate.

Specific experiments in support of designing a demonstration system were performed to gain a better understanding of conditions affecting treatment pathways and hybridized design strategies that can be incorporated into constructed wetland treatment, including the demonstration CWTS. The experiments included investigation of a zeolite for sorption of ammonia, the effect of water depth and seasonal variation on conditions and treatment, effect of adding a nutrient amendment, incorporation of an oil/water separator, and evapotranspiration.

Experiments were designed to compare seedling growth of corn (*Zea maya*), cabbage (*Brassica oleracea*), and soybean (*Phaseolus vulgaris*) between untreated and treated simulated produced water. The difference in early seedling growth between the waters was determined to be statistically significant, with higher growth rate in the treated water.

Toxicity tests using *Ceriodaphnia dubia* and *Pimephales promelas* compared the potential between untreated and treated simulated fresh produced water to cause adverse effects to receiving system biota. Statistical analysis of the experimental data indicated a significant difference in the response of *C. dubia* and *P. promelas* between untreated and treated simulated fresh produced water, with greater survival in the treated water.

Evapotranspiration data were collected. Theoretical lysimeter outflow determined using the Penman Monteith equation and mid-summer peak  $Typha\ latifolia$  (cattail) crop coefficient ( $K_c$ ) was compared with measured lysimeter outflow. The results demonstrate application of the Penman Monteith equation to estimating evapotranspiration in the pilot-scale CWTSs used in this investigation. Tracer tests indicate a significant difference in vertical transport of constituents caused by plant transpiration.

The influence of water depth on treatment performance of pilot-scale CWTSs for renovating simulated fresh oilfield produced water was investigated. Divalent metal concentrations were greater in outflow from wetland cells having 15 and 23 cm water depths than from cells having 46 and 56 cm water depths. The cells having 46 and 56 cm water depths provided reducing conditions conducive for dissimilatory sulfate reduction, which contributed to removal of divalent metals from the simulated produced water. Wetland cells with 15 and 23 cm water depths were more effective for removing oil from the simulated produced water than cells with greater water depths because shallower depth promoted conditions favorable for aerobic degradation of oil.

Produced water samples were collected from the demonstration CWTS site in central Alabama and analyzed. Comparison with surface water quality criteria yielded the following COCs to be targeted for treatment: ammonia, barium, cadmium, iron, manganese, nitrate, and selenium. A demonstration CWTS was designed and constructed to promote the biogeochemical conditions required for effective treatment of identified COCs in the produced water. Oxidizing cells (targeted DO greater than 2.0 mg/L) were included in each series to promote microbial nitrification of ammonia and precipitation of iron and manganese. Reducing cells (targeted DO less than 2.0 mg/L) were included to promote microbial denitrification of nitrate, dissimilatory sulfate reduction and metal sulfide formation, and dissimilatory selenium reduction. Performance of the demonstration CWTS was monitored using sampling methods that allow for calculation of removal efficiency, removal rate, and removal extent of constituents of concern. The demonstration CWTS successfully achieved consistent treatment of COCs even as their concentrations in the produced water varied by orders of magnitude.

Design strategies used in the pilot-scale and demonstration CWTSs to promote specific conditions that can be applied to designing full-scale CWTSs include plant and soil selection, water-depth selection, addition of amendments, hydraulic retention time (HRT), and hybrid components. These strategies allow conditions within a CWTS to be modified to achieve ranges necessary for the preferred biogeochemical processes for treating fresh to saline produced waters. In the case of renovating a produced water containing COCs that require different biogeochemical pathways for treatment, a CWTS can be designed with sequential cells that promote different conditions. By incorporating multiple wetland cells in a CWTS, conditions within each cell can be modified for removal of specific COCs. In addition, a CWTS designed with multiple cells allows for convenient sample collection so that biogeochemical conditions of individual cells can be monitored and performance can be evaluated. Removal rate coefficients determined from the pilot-scale CWTS experiments and confirmed by the demonstration system can be used to calculate HRTs required to treat COCs in full-scale CWTSs. The calculated HRTs can then be used to determine the surface area or "footprint" of a full-size CWTS for a given inflow rate of produced water.

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